



# Draft Genome Sequences of Nine Environmental Bacterial Isolates Colonizing Plastic

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**ABSTRACT** Here, we report the draft genome sequences of nine bacterial isolates obtained after laboratory incubation of seawater, soil, and wastewater samples with polylactic acid, polyethylene, or polyethylene terephthalate film for 2 weeks. Assuming colonization as a prerequisite of degradation, these strains could contribute to a solution to the global plastic waste problem.

Plastic is omnipresent in our environment, and there is no sustainable recycling solution. Many pollutants can be degraded by microorganisms, and degradation is often induced by colonization of the material by the organism. Aiming to contribute a biological solution to the plastic problem, we isolated nine bacterial strains colonizing plastic. Environmental samples (wastewater [WW], seawater [SW], and soil [S]) were obtained from a wastewater treatment plant (Lyngby-Taarbæk Forsyning A/S, Kongens Lyngby, Denmark [55°48'07.4"N, 12°32'20.8"E]) on 14 September 2018, from a harbor (Hellerup, Denmark [55°43'55.5"N, 12°34'51.0"E]) on 17 September 2018, and from a site near the university campus (55°47'07.8"N, 12°30'49.7"E) on 11 October 2018. On the marine and terrestrial sides, plastic pieces (seawater plastic [SWP] and soil plastic [SP]) were also collected. Microbial communities were removed from plastic by sonication for 5 min, and all environmental samples were inoculated at a starting concentration of 10<sup>6</sup> cells/ml (as determined by SYBR gold staining, filtration, and microscopy) in OECD301 medium (WW, S, and SP samples) or OECD306 medium (SW and SWP samples) (1, 2) with a piece of plastic (18 by 18 mm by 0.05 mm), i.e., poly-L-lactic acid (ME331050; Goodfellow), polyethylene terephthalate (ES301250; Goodfellow), or low-density polyethylene (ET311150; Goodfellow), or a cover glass (18 by 18 mm) (631-1567; VWR) as a control. OECD301 and OECD306 media contain 8.5 mg/liter KH<sub>2</sub>PO<sub>4</sub>, 217.5 mg/liter K<sub>2</sub>HPO<sub>4</sub>, 334 mg/liter Na<sub>2</sub>HPO<sub>4</sub>·2H<sub>2</sub>O, 5 mg/liter NH<sub>4</sub>Cl, 36.4 mg/liter CaCl<sub>2</sub>·2H<sub>2</sub>O, 22.5 mg/liter MgSO<sub>4</sub>·7H<sub>2</sub>O, and 0.25 mg/liter FeCl<sub>3</sub>·6H<sub>2</sub>O in either distilled water (OECD301) or seawater (OECD306) prepared with 2.5% sea salts (Sigma). Cultures were incubated in the dark at 16°C without shaking. After 14 days, the plastic pieces were transferred to fresh OECD301 or OECD306 medium and sonicated for 5 min. The samples were diluted 10-fold and plated onto OECD301 or OECD306 medium with 1.5% agar, 1% polypeptone, and 0.2% yeast extract. After incubation at 16°C for 7 days, colonies that were unique to each plastic type and different from the control samples were isolated by restreaking onto OECD301 or OECD306 1.5% agar plates with 1% polypeptone and 0.2% yeast and incubation for 4 days at 16°C.

Genomic DNA was extracted using the NucleoSpin tissue kit (740952; Macherey-Nagel). Five hundred nanograms of DNA per strain was submitted for sequencing at the Novo Nordisk Center for Biosustainability (Technical University of Denmark, Lyngby, Denmark) using the NextSeq 500/550 midoutput kit v2 (Illumina) for 150-bp paired-end sequencing on a MiSeq Illumina platform. The sequence data were analyzed with KBase v1.8.9 (3). The read quality was assessed using FastQC v0.11.5 (4) and Trimmomatic v0.36 (5) (sliding window size:4; sliding window minimum quality:15; post tail crop length:140; head crop length:10; leading minimum quality: 3; trailing

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**TABLE 1** Genomic features of nine environmental isolates colonizing plastic

Strain	Source	Plastic <sup>c</sup>	Closest strain	Estimated ANI (%) to closest strain	Genome size (bp)	G+C content (%)	Coverage (x)	No. of contigs	N <sub>50</sub> (bp)	No. of genes	SRA accession no.	GenBank accession no.	Assembly accession no.
IB03	WW	PLA	<i>Acidovorax radialis</i> N35 <sup>T</sup>	84.6	4,318,166	63.22	243	103	94,511	4,052	SRR13320098	JAEFCH0000000000	GCA_016406015.1
IB04	WW	PLA	<i>Pseudomonas veronii</i> DSM 11331 <sup>T</sup>	98.1	7,009,445	60.79	164	109	221,072	6,488	SRR13320097	JAEECG0000000000	GCA_016406005.1
IB05	WW	PLA	<i>Paracoccus versutus</i> DSM 582 <sup>T</sup>	77.2	5,294,826	63.23	220	108	95,395	5,052	SRR13320096	JAEECF0000000000	GCA_016405985.1
IB15	SW	PLA	<i>Vibrio gigantis</i> LGP 13 <sup>T</sup>	90.3	5,060,694	44.22	247	171	67,607	4,659	SRR13320095	JAEECE0000000000	GCA_016406055.1
IB21	SW	PE	<i>Alteromonas australica</i> H 17 <sup>T</sup>	80.3	4,538,050	44.43	318	78	134,642	3,961	SRR13320094	JAEECD0000000000	GCA_016405965.1
IB30	SWP	PE	<i>Paraglaeicola chathamensis</i> S118K6 <sup>T</sup>	97.8	5,067,268	44.18	240	118	125,622	4,389	SRR13320093	JAELLT0000000000	GCA_016405925.1
IB36	S	PET	<i>Delftia acidovorans</i> NBRC 14950 <sup>T</sup>	98.3	6,368,012	66.86	209	47	292,370	5,770	SRR13320092	JAEECC0000000000	GCA_016405945.1
IB41	S	PE	<i>Variovorax boronicumullans</i> NBRC 103145 <sup>T</sup>	89.7	6,867,556	67.50	135	49	283,377	6,448	SRR13320091	JAEECB0000000000	GCA_016405905.1
IB48	SP	PE	<i>Flavobacterium anhuiense</i> CGMCC 16859 <sup>T</sup>	89.0	5,603,394	33.47	262	36	663,986	4,730	SRR13320090	JAEFCA0000000000	GCA_016405855.1

<sup>a</sup>PLA, polylactic acid; PE, polyethylene; PET, polyethylene terephthalate.

minimum quality:3; minimum read length:36). The genomes were assembled using SPAdes v3.13.0 (6), and quality and metrics were analyzed using QUAST v4.4 (7). The level of contamination was assessed using CheckM v1.0.8 (8). The assemblies were automatically annotated with NCBI PGAP v5.0 (9). Species phylogeny was analyzed using autoMLST (10) (Table 1). Six strains had <95% estimated average nucleotide identity (ANI) with respect to genomes of type strains (11) and thus could represent novel bacterial species.

**Data availability.** The genome assemblies have been deposited in GenBank under BioProject number [PRJNA666993](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA666993), and detailed information is listed in Table 1.

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## REFERENCES

1. Organisation for Economic Cooperation and Development. 1992. Test no. 301: ready biodegradability, OECD guidelines for the testing of chemicals, section 3. OECD Publishing, Paris, France. <https://doi.org/10.1787/9789264070349-en>.
2. Organisation for Economic Cooperation and Development. 1992. Test no. 306: biodegradability in seawater, OECD guidelines for the testing of chemicals, section 3. OECD Publishing, Paris, France. <https://doi.org/10.1787/9789264070486-en>.
3. Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, Keegan KP, Kondo S, Kumar V, Land ML, Meyer F, Mills M, Novichkov PS, Oh T, Olsen GJ, Olson R, Parrello B, Pasternak S, Pearson E, Poon SS, Price GA, Ramakrishnan S, Ranjan P, Ronald PC, Schatz MC, Seaver SMD, Shukla M, Sutormin RA, Syed MH, Thomason J, Tintle NL, Wang D, Xia F, Yoo H, Yoo S, Yu D. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. *Nat Biotechnol* 36:566–569. <https://doi.org/10.1038/nbt.4163>.
4. Andrews S, Krueger F, Segonds-Pichon A, Biggins L, Krueger C, Wingett S. 2012. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
7. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
8. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
9. Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
10. Alanjary M, Steinke K, Ziemert N. 2019. AutoMLST: an automated web server for generating multi-locus species trees highlighting natural product potential. *Nucleic Acids Res* 47:W276–W282. <https://doi.org/10.1093/nar/gkz282>.
11. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.