

1 Degradation of dead cladoceran zooplankton and their contribution to organic carbon cycling in
2 stratified lakes: field observation and model prediction

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27 **KEYWORDS:** zooplankton carcasses; non-predatory mortality; sinking velocities; microbial
28 degradation; Lake Stechlin; simulation modeling

29 **Abstract**

30

31 The contribution of dead zooplankton biomass to carbon cycle in aquatic ecosystems is practically
32 unknown. Using abundance data of zooplankton in water column and dead zooplankton in sediment
33 traps in Lake Stechlin, an ecological-mathematical model was developed to simulate the abundance
34 and sinking of zooplankton carcasses and predict the related release of labile organic matter (LOM)
35 into the water column. We found species-specific differences in mortality rate of the dominant
36 zooplankton: *Daphnia cucullata*, *Bosmina coregoni*, and *Diaphanosoma brachyurum* (0.008, 0.129
37 and 0.020 d^{-1} , respectively) and differences in their carcass sinking velocities in metalimnion (and
38 hypolimnion): 2.1 (7.64), 14.0 (19.5) and 1.1 (5.9) m d^{-1} , respectively. Our model simulating
39 formation and degradation processes of dead zooplankton predicted a bimodal distribution of the
40 released LOM: epilimnic and metalimnic peaks of comparable intensity, ca. $1\text{ mg DW m}^{-3}\text{ d}^{-1}$.
41 Maximum degradation of carcasses up to ca. $1.7\text{ mg DW m}^{-3}\text{ d}^{-1}$ occurred in the density gradient
42 zone of metalimnion. LOM released from zooplankton carcasses into the surrounding water may
43 stimulate microbial activity and facilitate microbial degradation of more refractory organic matter;
44 therefore, dead zooplankton are expected to be an integral part of water column carbon source/sink
45 dynamics in stratified lakes.

46 **Introduction**

47

48 In aquatic ecosystems, decaying organic matter (both autochthonous and allochthonous)
49 including “necromass” resulting from non-consumptive mortality of plants and animals, is
50 considered to be a functionally important part of the carbon cycle (Benbow *et al.*, 2019; Benbow *et*
51 *al.*, 2020) but the contribution of non-consumptive mortality to carbon fluxes is rarely quantified
52 (Benbow *et al.*, 2020). In lentic ecosystems, zooplankton carcasses, which form “lake snow”,
53 represent a large portion of the decaying organic matter (Grossart and Simon, 1998; Benbow *et al.*,
54 2020). However, zooplankton carcasses are often overlooked as a vector of carbon sequestration
55 (Halfter *et al.*, 2021).

56 In general, zooplankton bodies contain high-quality substrates, i.e., proteins, lipids and
57 carbohydrates, while the less labile chitin comprises only ca. 7% of the body mass (Błędzki and
58 Rybak, 2016). A recent study showed that labile dissolved organic matter (DOM) released by
59 zooplankton carcasses contained over 7,000 compounds, many of which were N- and S- rich
60 (Johnston *et al.*, 2021). In freshwater habitats, dead zooplankton account for an average of 7.4 –
61 47.6% of the total zooplankton population (Tang *et al.*, 2014), making zooplankton carcasses a
62 ubiquitous and potentially abundant source of organic carbon, including labile organic matter
63 (LOM). This LOM, released by dead zooplankton, can improve bacterial utilization of recalcitrant
64 organic matter (water humus), including that of terrestrial origin by interactive (“priming”) effects
65 (Kolmakova *et al.*, 2019; Neubauer *et al.*, 2021).

66 There has been an increasing effort to understand the dynamics of zooplankton carcasses in
67 salt- and freshwaters (Tang *et al.*, 2006a; Tang *et al.*, 2009; Tang *et al.*, 2014; Giesecke *et al.*, 2017;
68 Dubovskaya *et al.*, 2018; Kolmakova *et al.*, 2019; Tang *et al.*, 2019; Silva *et al.*, 2020; Daase and
69 Søreide, 2021; Halfter *et al.*, 2021; Neubauer *et al.*, 2021, Diniz *et al.*, 2021). The abundance of
70 zooplankton carcasses is determined by the balance of their production and removal mainly due to
71 sinking and microbial degradation and to a lesser extent ingestion (Elliott *et al.*, 2010; Tang *et al.*,
72 2014; Daase *et al.*, 2014; Halfter *et al.*, 2021). The sinking rates determine how long zooplankton
73 carcasses will remain in the water column and thus how much they will be degraded, and in what
74 depth the maximum release of carcass organic matter occurs. In stratified lakes, the localization of
75 increased LOM input from dead zooplankton in specific water layers, i.e., density layers such as the
76 pycnocline, may determine the extent that interactive effects increase microbial DOM utilization,
77 particularly of low-bioavailable terrestrial DOM.

78 In stratified temperate lakes, for example, the water column is divided into epi-, meta- and
79 hypolimnion, each with distinct biological and physicochemical features, such as microbial
80 community compositions (Grossart and Simon, 1998; Simon *et al.*, 2002; Yue *et al.*, 2021; Rojas-

81 Jimenez *et al.*, 2021), nutrient concentrations, temperature, turbulence, light, pH and dissolved
82 oxygen (Wetzel, 2001). Yet, how these layers may differ in DOM degradation as stimulated by
83 interactive effects remains unclear due to the fact that the effects are often difficult to measure *in*
84 *situ* and may vary widely according to the lake's characteristics (Bengtsson *et al.*, 2018; Wologo *et*
85 *al.*, 2021).

86 The epilimnion has the highest rates of primary production, providing bacteria with labile
87 substrates, and the relatively high temperature and illumination can accelerate organic matter
88 mineralization (Moran and Zepp, 1997; Tranvik and Bertilsson, 2008; Aarnos *et al.*, 2012;
89 Fonvielle *et al.*, 2021). The metalimnion often exhibits increased microbial activities in association
90 with the oxygen minimum zone (Schram and Marzolf, 1994; Kreling *et al.*, 2017). The metalimnic
91 temperature is still high enough to support rapid microbial growth, and the proximity to the
92 hypolimnion provides heterotrophs with limiting nutrients. In addition, the metalimnion receives a
93 high input of fresh organic particles, including zooplankton carcasses, settling from the epilimnion,
94 which accumulate at the pycnocline and sustain high microbial activities there (Grossart and Simon,
95 1998; Simon *et al.*, 2002). Thus, LOM derived from zooplankton carcasses in the epi- and
96 metalimnion may stimulate microbial activities and hence overall interactive effects on
97 allochthonous carbon cycling in these layers.

98 The fluxes of zooplankton carcasses within the water column largely depend on physical
99 (e.g., turbulence and sinking velocity) and chemical (pH, oxygen and nutrient availability)
100 variables, but also biological variables (e.g., food web dynamics, zooplankton community
101 composition, mortality rates and habitat depth) (reviewed in Tang *et al.*, 2014). For example,
102 increased turbulence in the epilimnion increases the probability that zooplankton carcasses remain
103 in a given water layer for a prolonged time period (Kirillin *et al.*, 2012); hence, the carcasses may
104 be subject to increased microbial degradation (Tang *et al.*, 2006a; Tang *et al.*, 2009). Once they
105 have reached the metalimnion, the carcasses leave the turbulence zone and begin to sink rapidly and
106 continue through the hypolimnion, where microbial degradation is expected to occur at a much
107 lower rate due to the lower water temperature. Thus, individual lake features would determine the
108 fraction of organic matter that is microbially degraded or remains on the zooplankton carcasses
109 when settling to the hypolimnion and finally being deposited in the sediments, i.e., contribute to
110 carbon sequestration.

111 To understand the dynamics of the dead zooplankton pool in a stratified lake, sinking and
112 degradation of natural zooplankton carcasses throughout the water column of Lake Stechlin
113 (Germany) were investigated. Based on the studied dynamics in vertical distribution, mortality and
114 sinking rates of the dominant zooplankton species, a simulation model summarizing the
115 contributions of individual species was developed in order to calculate intensity profile of LOM

116 from dead zooplankton throughout the stratified water column. The results of our field and
117 modeling study advance our understanding of the role of dead zooplankton in microbial organic
118 matter degradation and transformation.

119

120 **Method**

121

122 *Study site*

123 Lake Stechlin (53°10' N, 13°02' E) is a deep dimictic oligo-mesotrophic lake in northeastern
124 Germany (Koschel and Adams, 2003) which is characterized by an area of 4.23 km², a mean depth
125 of 23.3 m, a maximum depth of 69.5 m. Lake Stechlin is thermally stratified from April until the
126 end of December. In 2015- 2017, the lake showed anshowed a strong increase in total phosphorus to
127 33 µg L⁻¹ and annual mean chlorophyll-a concentration in surface waters to 3.3 µg L⁻¹ (Scholtysik
128 et al., 2020), which altered its trophic status between the mesotrophic and eutrophic states. Changes
129 in phytoplankton and zooplankton biomass revealed the proliferation of cyanobacteria and the
130 decrease of large-sized zooplankton (*Eudiatomus*, *Eurytemora*) (Selmechy et al., 2019). Aside of
131 the basic physical, chemical and biological parameters, the *in situ* live-dead zooplankton
132 composition (Bickel et al., 2009), microbial zooplankton carcass decomposition (Tang et al., 2006a,
133 2009) and non-predatory mortality of *Bosmina longirostris* (O.F. Müller, 1776) (Dubovskaya et al.,
134 2015) have been investigated in detail. Our study was conducted on 27th July – 1st August 2017 at a
135 20 m deep sampling point in the south-western basin (Fig. 1).

136

137 *Sediment traps and water column sampling*

138 The methods were similar to our previous field experiments estimating zooplankton non-predatory
139 mortality rate (Dubovskaya et al., 2015; Dubovskaya et al., 2018) with the same accuracy and
140 precision of sampling and trap performance. We used custom-made sediment traps of the Håkanson
141 (1984) type, which consisted of a pair of cylindrical collectors without lids (each cylinder was 0.077
142 m in diameter × 0.485 m height). We deployed three sets of 2 sediment traps (at 7.0 ± 0.5 m and
143 14.0 ± 0.5 m depths) for 24 h each of five consecutive days during the summer stratification period
144 in late July - early August 2017. Each set of traps was moored to an anchor and a submerged buoy,
145 which was marked by a small float. Before deployment, the trap cylindrical collectors were filled
146 with water from the corresponding deployment depth pre-screened through a 90-µm mesh. Upon
147 trap retrieval, water from the paired collectors of each trap was pooled and concentrated on a 90-µm
148 mesh. Zooplankton samples were stained with aniline blue for carcass counting (Y in Equation 1;
149 see below).

150 Zooplankton samples were collected daily near the traps at around 9-10 a.m. with an open-
151 close plankton net (90 μm mesh and mouth of 17 cm dia.) from 5 vertical tows: 0-3, 3-6, 6-9, 9-12,
152 and 12-15 m. Samples from layers of 6-9 m and 12-15 m were used to determine carcass abundance
153 at trap exposure depth (y^*) for the upper trap (7 m, epilimnion) and the lower trap (14 m, below
154 thermocline at 8-10 m), respectively.

155 The carcasses accumulation in sedimentation traps and corresponding abundance at trap
156 exposure depths were measured reliably for all studied cladoceran species except for *Chydorus*
157 *sphaericus* (O.F. Müller, 1776) in the metalimnion. Positioning the traps in an area of variable live
158 and dead *C. sphaericus* populations (7 m) led to significant uncertainties in determining the average
159 background concentration of carcasses per day and consequently for sinking velocities
160 calculationcalculations. Since field measurements of *C. sphaericus* sinking velocity at 7 m were not
161 reliable, this species was excluded from our model analysis. Yet, taking into account that *C.*
162 *sphaericus* was not abundant and it amounted to maximal 11% of the total cladoceran biomass, its
163 exclusion should not significantly affect our analysis of the processes of formation, destruction and
164 sinking of zooplankton carcasses in Lake Stechlin.

165

166 *Staining, counting and biomass calculation*

167 To distinguish between live and dead zooplankton, all zooplankton samples were stained
168 within an hour after collection with water-soluble aniline blue (Bickel *et al.*, 2009) and fixed in 10%
169 formalin. Dominant species were counted and measured under a dissecting microscope. These
170 species were Cladocera *Daphnia cucullata* (Sars, 1862), *Diaphanosoma brachyurum* (Liévin,
171 1848), *Chydorus sphaericus* and *Bosmina coregoni* (Baird, 1857). The copepods *Thermocyclops*
172 *oithonoides* (Sars, 1863) and *Eurytemora lacustris* (Poppe, 1887) were not counted in the samples
173 since their abundance was negligible in the epi- and metalimnion and increased only below 12 m.
174 For samples with > 500 individuals, random subsamples were examined until at least 50 individuals
175 of the same species were counted.

176 The biomass of zooplankton was calculated based on a linear size-biomass relationship
177 using power-law formulas (McCauley, 1984). Average sizes were obtained by measuring 200
178 individuals under an inverted microscope (Nikon, Diaphot 300, Japan).

179

180 *Sinking velocities and fluxes of zooplankton carcasses*

181 The vertical flux F_h (ind. $\text{m}^{-2} \text{d}^{-1}$) of carcasses at the trap exposure depth h was measured by
182 sediment traps as

183
$$F_h = \frac{Y}{S}, \quad (1)$$

184 where Y is the number of carcasses accumulated in a sediment trap per day (ind. d⁻¹), $S = 0.0093 \text{ m}^2$
 185 is the input area of the trap. Sinking velocity of carcasses v_h (m d⁻¹) at the depths of trap exposure
 186 was derived from the concentration of carcasses at the exposure depth y_h (ind. m⁻³) and the
 187 calculated vertical flux F_h as

188
$$v_h = \frac{F_h}{y_h}. \quad (2)$$

189 A normalized flux of carcasses \bar{F}_h (d⁻¹) to the number of live zooplankton was calculated as

190
$$\bar{F}_h = \frac{F_h}{\int_0^h N(z) dz}, \quad (3)$$

191 where z is the depth over the layer $0 < z < h$, N is the abundances of live individuals derived from
 192 plankton net tows. If the settling carcasses are not destroyed in the water column, then their
 193 normalized flow should be the same at all depths. However, if the normalized flux decreases with
 194 depth, then this indicates the process of destruction of carcasses within the strata, which has to be
 195 considered when modeling zooplankton mortality and LOM release. It is worth noting that the value
 196 of the normalized flux is an integral value that depends only on the specific mortality and the
 197 number of live zooplankton in the water column above the trap. In the steady state, the flux is not
 198 related to the number of dead zooplankton in the water column or to the form of their vertical
 199 distribution.

200

201 *Model of formation, sinking, microbial degradation and destruction of dead zooplankton*

202 The model was constructed to calculate the amount of LOM release by carcasses into the
 203 water column. The model is modified from the one used to calculate the vertical distribution of live
 204 and dead copepod *Arctodiaptomus salinus* (Daday, 1885) in Lake Shira (Tolomeev *et al.*, 2018), by
 205 adding the depth dependence of carcass sinking rate and microbial degradation.

206 It is assumed that the total amount and the vertical distribution of live zooplankton do not
 207 change significantly over time. Its vertical distribution is approximated by k layers of thickness Δz ,
 208 with corresponding positions $\Delta z \leq z_i \leq k \cdot \Delta z$ and piecewise-homogeneous zooplankton abundances
 209 N_i , with $i = 1$ to k . Dynamics of the carcass abundance for the i -th layer is described by the stationary
 210 equation for vertical advection with a zero-order source (carcasses production) term and a first-order
 211 removal (carcasses destruction) term:

212

$$\frac{\partial v(z)y_i}{\partial z} = mN_i\delta(z-z_i) - D(z)y_i, \quad (4)$$

214 where y and N are abundances of carcasses and live individuals (ind. m⁻³), respectively; m (d⁻¹) is
 215 the specific non-predatory mortality, $v(z)$ is the depth-dependent sinking velocity of carcasses (m d⁻¹),
 216 $D(z)$ is the depth-specific rate of carcass removal or destruction rate (d⁻¹). Here, $\delta(x)$ is the unit
 217 sample function defined as

218

$$\delta(x) = \begin{cases} 1 & \text{if } x < 0 \\ 0 & \text{otherwise} \end{cases}.$$

220

221 By definition, $F = v(z)y$ is the vertical flux of carcasses. The solution of Eq. (4) can be written as,

$$y_i(z) = \delta(z - z_i) \frac{mN_i}{D_i} \left(1 - e^{-\frac{D_i}{v_i}\Delta z} \right) + [1 - \delta(z - z_i)] y_{i,z_i} e^{-\int_{z_i}^z \frac{D(z)}{v(z)} dz} \cdot \frac{v(z)}{v_i}. \quad (5)$$

223 The integrated distribution of the carcass concentrations $y(z)$ depicts the sum of the solutions y_i :

224

$$y(z) = \sum_{i=1}^k y_i(z). \quad (6)$$

226

227 The important factor that influences the rate of carbon cycling in the water column is the
 228 release of LOM by sinking zooplankton carcasses. We separated the model parameterization of the
 229 LOM release rate ($\mu\text{g DW m}^{-3} \text{ day}^{-1}$) in two processes with different time scales: the “microscale”
 230 $I_{DW\rho}(z)$ describes the slow and continuous loss of density due to continuous release of organic
 231 matter by microbial degradation, while the “macroscale” $I_D(z)$ represents the mechanical
 232 fragmentation of carcasses (the stage of entire carcasses disappearance, suggesting their quick
 233 degradation). The amount of LOM released by a carcass is proportional to the change in its density.
 234 The density ρ_o corresponds to the initial body weight of the zooplankter $DW\rho_o$ (μg), and the lower
 235 limit density $\rho_{term} = 1000 \text{ kg m}^{-3}$ corresponds to its complete destruction; i.e., the density at which
 236 the amount of LOM released into the medium is equivalent to the entire body weight of the
 237 zooplankter. The weight of the carcass, depending on ρ , is determined by the formula:

$$DW\rho = DW\rho_o \cdot \frac{\rho - \rho_{term}}{\rho_o - \rho_{term}} \quad (7).$$

239

240 Elliott *et al.* (2010) proposed the empirical equation describing the carcass density
 241 dependence on time and temperature:

242

$$\rho = \rho_o - 3.78 \cdot (1 - e^{-0.329T}) \cdot (\ln(t) + 1.39), \quad (8)$$

243

244

245 where $\rho_0 = \rho(0.25) = 1045$ is the initial carcass density (kg m^{-3}), T - temperature ($^{\circ}\text{C}$) and t - time
246 after death (from 0.25 h onward). Ranging from $t = 0$ to $t = 0.25$ h, the density ρ is not defined,
247 therefore, we will assume that the density of carcasses does not change in the first 0.25 hours after
248 animal death.

249 According to Equation 8, the most intense release of LOM occurs within the first hours after
250 death (logarithmic dependence on time). Zooplankton carcasses within the water column consist of
251 carcasses that have different "ages" from the moment of death and therefore have different rates of
252 LOM release; therefore, to calculate the LOM profile, it is necessary to know not only the total
253 number of carcasses, but also their "age" structure in each layer.

254 The rate of release of LOM from zooplankton carcasses can be determined based on the
255 obtained i -solutions $y_i(z)$ by Equation 5, reflecting the total number and the "age" structure of dead
256 zooplankters. Let us denote the rate of LOM release in the i -solution associated with changes in
257 carcasses' density as $I_{DW\rho_i}(y_i, z)$, whose vertical profile is defined as:

258

$$259 \quad I_{DW\rho}(z) = \sum_{i=1}^k I_{DW\rho_i}(y_i, z) \quad (9)$$

260

261 We will find the vertical distributions $I_{DW\rho_i}$ as discrete values of the LOM release rate in
262 layers \mathbf{z}_{i+n} , where n is an integer $0 \leq n \leq k - i$. The LOM flux in layer \mathbf{z}_i , associated with the
263 change in carcass body weight in each i -solution can be described as follows:

$$264 \quad I_{DW\rho_{i,z_i}} = (DW\rho_0 - DW\rho_{i,z_i}) \cdot y_{i,z_i} \cdot \frac{v_{i,z_i}}{\Delta z} \quad (10)$$

265 where $DW\rho_0$ is initial weight of carcass, $DW\rho_{i,z_i}$ is weight of carcass after time $\Delta t_{i,z_i} = \frac{\Delta z}{v_{i,z_i}}$, in
266 which the carcass sinks to the lower boundary of the layer \mathbf{z}_i . At depth $z > z_i$ the LOM flux is
267 defined as

$$268 \quad I_{DW\rho_{i,z_{i+n}}} = (DW\rho_{i,z_{i+n-1}} - DW\rho_{i,z_{i+n}}) \cdot y_{i,z_{i+n}} \cdot \frac{v_{i,z_{i+n}}}{\Delta z}, \quad (11)$$

269 where n is an integer in the range $1 \leq n \leq k - i$. $DW\rho_{i,z_{i+n-1}}$ and $DW\rho_{i,z_{i+n}}$ are
270 the weight of carcasses at the upper and lower boundaries of the layer \mathbf{z}_{i+n} , respectively. $y_{i,z_{i+n}}$ is
271 the number of carcasses at the depth z_{i+n} , $v_{i,z_{i+n}}$ is the sinking velocity of carcasses in the layer
272 \mathbf{z}_{i+n} .

273 Calculation of the LOM flux profile by Equation 11 requires finding $DW\rho$ corresponding to
274 the densities $\rho_{i,z_{i+n-1}}$ and $\rho_{i,z_{i+n}}$ for each water depth. However, Equation 7 does not allow direct
275 estimate of $\rho_i(z)$ at arbitrary water depths below z_i due to the undefined variables $v(z)$ and $T(z)$.

276 Nevertheless, it is possible to find $\rho_i(z)$ using a recursive procedure calculating $\rho_{i,z_{i+n}}$ from the
 277 value on the previous time step $\rho_{i,z_{i+n-1}}$. With the density of carcasses at the upper boundary of the
 278 current layer known, the reference time t' can be calculated back to the moment of zooplankter'
 279 death assuming constant $v(z) = v_{i,z_{i+n}}$ and $T(z) = T_{i,z_{i+n}}$. We find the time t' by the backward
 280 solution of Equation 8 with regard to t .

$$281$$

$$282 \quad t' = \exp \left[\frac{e^{0.329 \cdot T} \cdot (\rho_0 - \rho_{i,z_{i+n-1}})}{3.78 \cdot (e^{0.329 \cdot T} - 1)} - 1.39 \right] \quad (12)$$

283

284 Then, substituting time $(t' + \Delta t_{i,z_{i+n}})$ into equation 8 as t , we yield $\rho_{i,z_{i+n}}$, and can repeat the cycle
 285 until the lower limit of the simulated depth range is reached. As a result, we can determine all $\rho_{i,z_{i+n}}$
 286 needed to compute $DW \rho_{i,z_{i+n}}$ and thus obtain the depth profile $I_{DW\rho}(z)$ by Equation 9.

287 Another form of release of organic matter by dead zooplankton is the complete destruction of
 288 carcasses, determined by $D(z)$ in Equation 4. In this case, the organic matter enters the environment
 289 in the form of fragments of crustacean bodies. This type of organic matter can also play an important
 290 role in accelerating the carbon cycle of the water column because the particles still contain bacteria
 291 and fungi decomposing them in an attached form. The intensity of the release of organic matter $I_D(z)$,
 292 determined by the destruction of carcasses, is also calculated based on the sum of i -solutions
 293 $y_i(z)$ similarly to the equation 9:

$$294$$

$$295 \quad I_D(z) = \sum_{i=1}^k I_{D_i}(y_i, z). \quad (13)$$

296

297 However, instead of calculating the losses associated with the decrease in carcass weight during
 298 their passage through the water layer, it is necessary to determine the difference in carcass numbers
 299 entering and leaving the water layer. Multiplying the obtained value by the initial carcass weight in
 300 the considered layer and dividing it by the time required for passing the water layer, we obtain value
 301 I_{D_i} . In layer z_i it will be defined as

$$302 \quad I_{D_i,z_i} = (m \cdot N_{i,z_i} t_{i,z_i} - y_{i,z_i}) \cdot DW \rho_0 \cdot \frac{v_{i,z_i}}{\Delta z}, \quad (14)$$

303 and at depth $z > z_i$ as

$$304 \quad I_{D_i,z_{i+n}} = (y_{i,z_{i+n-1}} \cdot \frac{v_{i,z_{i+n-1}}}{v_{i,z_{i+n}}} - y_{i,z_{i+n}}) \cdot DW \rho_{i,z_{i+n-1}} \cdot \frac{v_{i,z_{i+n}}}{\Delta z}, \quad (15)$$

305 here n depicts an integer in the range $1 \leq n \leq k - i$.

306

307 *Use of smoothed zooplankton distributions for the model*

308 Modeling the vertical distribution of dead zooplankton and the resulting LOM flux requires
309 detailed distributions of live and dead zooplankton for each layer z_i as initial input. Distributions of
310 zooplankton were obtained from field observation data. For each day, the distribution data were
311 smoothed by the method of kernel density estimation for heaped and rounded data (Gross, 2015).
312 The obtained profiles of live and dead zooplankters of each species were further used for model
313 calculations.

314

315 *Specific mortality m and sinking velocities of carcasses $v(z)$ at condition $D(z) \approx 0$.*

316 Laboratory experiments indicate that at 20 °C zooplankton carcasses did not reach the late
317 stage of degradation until ca. 78 hours (3.25 days) after death (Tang *et al.*, 2006b). Consequently,
318 zooplankton species with high sinking rates (~ 20 m day⁻¹) will have sufficient time to leave the
319 water column before full disintegration. Under these conditions, we can assume $D(z) \approx 0$ that
320 makes possible calculation of the absolute values of m and $v(z)$ in a steady state system by the
321 established formulas (Dubovskaya *et al.*, 2015):

322

323
$$m = \frac{y^* \cdot v^*}{\bar{N} \cdot h} , v(z) = \frac{m \cdot \bar{N}_z \cdot z}{y(z)} \quad (16)$$

324 where y^* is carcass concentration at trap depth h , v^* is carcass sinking velocities at trap depth, \bar{N} is
325 averaged abundance of live individuals over the layer $0 < z < h$, \bar{N}_z is averaged concentration of
326 live individuals in the water column from the surface to depth z .

327

328 *Specific mortality m and sinking velocities of carcasses $v(z)$ at condition $D(z) \gg 0$.*

329 If the factor depth significantly affects the value of normalized flow $\bar{F}(z)$, then this indicates
330 high rates of carcass removal rate in the water column, and $D(z)$ becomes important for the model.
331 Since the rate of degradation is strongly dependent on temperature, one can expect that in the
332 warmer epilimnion, disintegration of the carcasses will occur more rapidly. However, the carcasses
333 start to break apart only on the late stages of degradation, after the loss of a major portion of their
334 carbon content. The rate of carbon release varies between species, but it usually occurs over several
335 days. For instance, 50% of copepod carcass carbon is mineralized within 6–12 d at a temperature
336 close to 20°C (Franco-Cisterna *et al.*, 2021). In Lake Stechlin, *Daphnia cucullata* carcasses lose 26
337 to 43% C d⁻¹ over a 2-day *in situ* experiment (Tang *et al.*, 2009). Even in the late stage of
338 decomposition when most of the carcasses' internal tissues have disappeared, the chitinous carapace
339 remains relatively intact (Tang *et al.*, 2009). Consequently, despite the high rate of carbon loss by
340 carcasses in the epilimnion, the carcasses even at low rates of sinking (1-2 m d⁻¹) have a high
341 probability to leave the epilimnion before disintegration. Thus, it can be assumed that removal rate

342 of carcasses in the epilimnion zone is insignificant compared to the lower layers. Therefore, we set
 343 D to zero in the upper 7 m of the water column (epilimnion), so that, m and $v(z)$ in the layer 0-7 m
 344 can be found from Equation 16.

345 In order to obtain $D(z)$ below 7 m it is necessary to know $v(z)$ below this depth. For this,
 346 we used the sinking velocities of carcasses measured *in situ* at 7 m and 14 m. Denoting $v_{D0}(z)$ as
 347 the “reference” sinking velocity at $D(z) \approx 0$, and $z_7 = 7$ m and $z_{14} = 14$ m as the depths
 348 corresponding to the measured sinking velocities, the actual sinking rate $v(z)$ passing through z_7
 349 and z_{14} can be approximated linearly as:

$$350 \quad v(z) = K(z) \cdot v_{D0}(z) \text{ for } z_7 < z < z_{14}, \quad (17)$$

$$351 \quad K(z) = b + a \cdot z,$$

352 where b and a are the coefficients of the line passing through $K(z_7) = 1$ and $K(z_{14}) =$
 353 $v_{trap}(z_{14}) / v_{D0}(z_{14})$. Fig. 2 shows an example approximating sinking velocity curve $v(z)$ for
 354 *Daphnia* (data collected on 1st August, 2017) assuming $D(z) \gg 0$, passing through the measured
 355 sinking velocity point at 14 m. Given the distribution $v(z)$ for $D(z) \gg 0$ in the 7-14 m layer, D can
 356 be calculated from Equation 5. It however involves solution of a transcendental equation. As a
 357 workaround, we assume that the destruction rate of carcasses produced in each single layer z_i is
 358 equal to zero (i.e., in layer z_i , only the carcasses degrade, deposited from the upper layers). The
 359 assumption is plausible because a newly formed carcass is unlikely to be immediately destroyed.
 360 Then the destruction rate D_{z_i} is determined as follows:

361

$$362 \quad D_{z_i} \simeq \ln\left(-\frac{y_{i-1} \cdot \frac{v_{i-1}}{v_i}}{m \cdot N \cdot \frac{\Delta z}{v_i} - y_i}\right) \cdot \frac{v}{\Delta z} \quad (18)$$

363

364 All model calculations were implemented in the programming language R, ver. 4.1.1 (R
 365 Core Team, 2021), using the basic R functions and the tidyverse packages (Wickham *et al.*, 2019).
 366 To find the boundaries of the metalimnion, we used the R package rLakeAnalyzer (Winslow *et al.*,
 367 2019).

368

369

370 **Results**

371 *Vertical distribution in abundance of live and dead zooplankton and carcass sinking flux*

372 Cladocerans dominated the zooplankton community, represented by four main species:
 373 *Daphnia cucullata* and *Bosmina coregoni* in the epilimnion; *Diaphanosoma brachyurum* and
 374 *Chydorus sphaericus* in the metalimnion (Fig. 3 - 6). Among these, *D. cucullata* and *D. brachyurum*

375 contributed ca. 50% and 30% of the biomass, respectively (Table 1). The averaged percentages of
376 carcasses among these zooplankton species ranged from 4.6% to 8.9% throughout the water column,
377 0-15 m (Table 1).

378 Abundance of live and dead cladocerans in the water column per m² varied noticeably during
379 the six days of sampling. However, these fluctuations usually did not exceed a factor of two. An
380 exception was the abundance of live *B. coregoni*, which varied by a factor of 6.5 (Fig. 4 C). The
381 distribution of *D. cucullata* and *B. coregoni* populations within the epilimnion zone was patchy. We
382 observed substantial changes in the abundance of live individuals in different layers of the epilimnion
383 over time (Fig.3 A, 4 A). The dynamics and spatial distribution of metalimnetic populations of *D.*
384 *brachyurum* and *C. sphaericus* were more consistent (Fig. 5 A, 6 A).

385 The dynamics of dead and live individuals of *D. cucullata* were closely linked with each
386 other (Fig. 3 A-C). Changes in abundance of carcasses of *B. coregoni* also followed those of living
387 individuals (m⁻²), except on 31st July (Fig. 4 A-C). However, the dynamics of carcasses of both *D.*
388 *brachyurum* and *C. sphaericus* differed substantially from live individuals (Fig. 5 A-C and Fig. 6A-
389 C).

390 Carcass sinking fluxes of *D. cucullata* and *B. coregoni* measured by sediment traps each day
391 were similar at 7 m and 14 m depths, but differed markedly between the sampling time points (Fig.3
392 D, 4 D). In contrast, the sinking flux of *D. brachyurum* carcasses considerably decreased from 7 m
393 to 14 m (about 2.5 times) (Fig. 5 D). Accumulation of *C. sphaericus* carcasses was also lower in the
394 lower traps (Fig.6 D).

395

396

397 *Sinking velocities and normalized fluxes*

398 Distribution of sinking velocities of *B. coregoni*, *D. cucullata*, and *D. brachyurum* at 7 m
399 and 14 m depths, i.e. in the meta- and hypolimnion, revealed a similar pattern (Table 1). Sinking
400 velocities of these cladocerans were significantly lower at 7 m than at 14 m. *B. coregoni* carcasses
401 sinking rate was the highest among all species and reached 14 and 20 m d⁻¹ at 7 and 14 m depths,
402 respectively. Yet, the increase in *B. coregoni* sinking rate at 14 m was not pronounced (ca. 1.5-fold)
403 when compared to *D. cucullata* and *D. brachyurum* carcasses, which were sinking at 1 - 2 m d⁻¹ in
404 the metalimnion, but increased to 6 - 8 m day⁻¹ at 14 m depth (Table 1).

405 Differences in normalized fluxes of *D. cucullata* and *B. coregoni* carcasses between 7 m
406 and 14 m depths were not significant (Table 1), indicating that these carcasses were not
407 substantially degraded in the water column. It is worth noting that the normalized fluxes of *B.*
408 *coregoni* carcasses (0.110-0.134 d⁻¹) were an order of magnitude higher than those of *D. cucullata*

409 (0.006-0.007 d⁻¹). The decrease in the normalized flux of carcasses of *D. brachyurum* from 0.017 d⁻¹
410 at 7 m to 0.003 d⁻¹ at 14 m was significant (Table 1).

411

412 *Simulation results*

413 The non-predatory mortality was highest for *B. coregoni* (0.129 d⁻¹), followed by *D.*
414 *brachyurum* (0.020 d⁻¹) and finally *D. cucullata* (0.008 d⁻¹; see also Table 2). The significantly
415 higher mortality of *B. coregoni* than of other species was consistent with the highest percentage of
416 dead *B. coregoni* in the water column (Table 1).

417 Sinking velocities of dead individuals of these cladoceran species had a similar pattern of
418 vertical distribution (Fig.7). The calculated sinking velocities were minimal in the epilimnion (ca. 1-
419 2 m d⁻¹ for *D. cucullata* and *D. brachyurum* and ca. 6-7 m d⁻¹ for *B. coregoni*), which is consistent
420 with the higher epilimnic turbulence. From the metalimnion to the hypolimnion, carcass sinking
421 rates increased, but in the deeper part of the hypolimnion they tended to stabilize or even decrease
422 (below 12 m for *B. coregoni* and *D. brachyurum*). Sinking velocity of *D. cucullata* carcasses,
423 however, further increased at 14 m. Yet, accuracy of sinking velocity calculations decreased at
424 lower depths as live and dead zooplankton abundances were relatively low in these water layers. It
425 may seem confusing that carcass sinking velocities near the surface were close to zero. In fact, the
426 model does not show the physical sinking velocity of individual carcasses, but rather the resulting
427 settling and rising velocities of the whole population. For example, if turbulent vortices hold the
428 carcass in the surface layer, the resulting sinking velocity would be indeed low.

429 Although the decrease in normalized carcass fluxes from 7 to 14 m for *D. cucullata* and *B.*
430 *coregoni* was statistically insignificant (Table 1), we assume that the degradation of dead
431 zooplankton in the water column was within our methodological accuracy. Nevertheless, we
432 estimated carcass destruction rates (*D*) assuming a proportional increase in sinking velocities
433 between 7 and 14 m, according to the Eqs. 17 and 18. Maximum destruction rate *D* in the 7-14 m
434 water layer were 0.36 d⁻¹ and 0.71 d⁻¹ for *D. cucullata* and *B. coregoni*, respectively, and occurred
435 at the lowest depth, i.e. 14 m (Table 2, Fig.7).

436 For *D. brachyurum*, the normalized carcass flux decreased by as much as 82% between 7
437 and 14 m (Table 1). The large difference between the measured sinking velocity (5.9 m d⁻¹; Table 1)
438 and the calculated velocity (34.9 m d⁻¹; Table 2) at 14 m indicates a high destruction rate of *D.*
439 *brachyurum* carcasses. Our model (without any carcass destruction) and actual sinking velocities of
440 *D. cucullata* and *B. coregoni* carcasses differed only by 1.5 and 1.2 times, respectively, but by
441 almost 6 times for *D. brachyurum* (Table 2). The destruction rate of *D. brachyurum* was estimated
442 to be 3.99 d⁻¹(Table 2).

443 Different species contributed differently both to the formation of total LOM of dead
444 zooplankton, and to its separate fractions $I_{DW\rho}$ and I_D (Table 2). The largest contribution to LOM
445 as $I_{DW\rho}$ was made by *B. coregoni* at 3.87 ± 0.42 mg DW m⁻² (44.3%), due to its high population
446 mortality during the study period and the low rate of carcasses destruction. The highest input of
447 LOM in the form of destroyed carcasses I_D was from *D. brachyurum* at 2.93 ± 0.54 mg DW m⁻²
448 (62.4 %). *D. cucullata*, despite dominating the biomass (50%) of all cladocerans, contributed
449 relatively little LOM: 2.18 ± 0.13 mg DW m⁻² (25.6 %) and 0.54 ± 0.15 mg DW m⁻² (10.1 %) of the
450 fractions $I_{DW\rho}$ and I_D , respectively, because of its low mortality rate during the observation period.

451 The profile $I_{DW\rho}$ showed two maxima, one in epilimnion (2.5 m) formed by *B. coregoni*
452 and *D. cucullata*, and one in metalimnion (7.5 m) formed mainly by *D. brachyurum* (Fig. 8). Both
453 maxima were comparable to each other and yielded an input of LOM of 0.7 - 1.2 mg m⁻³ d⁻¹. Since
454 the highest rate of organics released by the crustacean carcasses has been observed in the first few
455 hours after the animal's death, the profile of LOM influx practically coincided with the distribution
456 of carcasses in the water column (Fig. 8). In other words, the carcasses lost most of their organic
457 matter not far from the place of their formation.

458 Total destruction of carcasses I_D began in the metalimnion and reached maximum values of
459 0.8 mg m⁻³ d⁻¹ approximately at the bottom of the thermocline at 8 m depth and then gradually
460 decreased with increasing depth. A smooth reduction in I_D was achieved by a balance between
461 some increase in carcass destruction rate with sinking time and the decrease in carcass abundance
462 with depth (Fig. 8).

463

464 Discussion

465 We revealed distinct species-specific differences in carcass distribution, sinking and
466 potential release of LOM and microbial degradation in the water column of a stratified lake. In
467 particular, planktonic crustaceans differed in their abundances, mortality rates, sinking velocities,
468 and carcass degradation rates in different water layers, i.e., epi-, meta-, and hypolimnion. We tried
469 to distinguish between the release of organic matter in the beginning and later stages of carcass
470 degradation and relate them to the spatial structure of cladoceran zooplankton using the modeling
471 approach.

472 The analysis of the processes of carcass degradation by means of the model revealed
473 findings that were not obvious. For example, the abundance of *B. coregoni* carcasses was 5 times
474 lower than *D. cucullata* carcasses and it was minimal among all cladoceran species, but the sinking

475 velocities of *B. coregoni* carcasses were the highest, suggesting a short residence time in the water
476 column. The high sinking velocity of *B. coregoni* carcasses in comparison to all other cladocerans
477 was probably due to their relatively higher body density. *B. coregoni*, which had a body size ca. 1.6
478 times smaller than *D. cucullata*, nevertheless had almost the same body weight (1.9 $\mu\text{g DW}$).
479 Therefore, one would expect *B. coregoni* to make a smaller contribution to the total LOM.
480 However, the overall mortality of the *B. coregoni* population was the highest of all species studied,
481 exceeding the mortality of *Daphnia* by ca. 16 times. This compensated for the relatively low
482 abundance of this species and its high carcass sinking velocity, resulting in a large *Bosmina*
483 contribution to the total LOM flow via zooplankton carcasses. The $I_{DW\rho}$ fraction of LOM released
484 by *B. coregoni* was the highest among all species, at 44 %. The abundance of *D. brachyurum* and
485 *D. cucullata* carcasses were of the same magnitude, but *D. brachyurum* contributed 62% (I_D
486 fraction) compared to 10 % for *D. cucullata* to the total LOM, due to the higher destruction rate
487 and the lower sinking velocity of *D. brachyurum* carcasses.

488 It is worth noting that non-predatory mortality values can vary over a wide range for natural
489 cladoceran populations from low during growth and steady-state to high during population decline
490 (Gladyshev et al., 2003; Tang et al., 2014). The development conditions for the *B. coregoni*
491 population during the observation period were likely less favorable than for other species. For
492 instance, the non-predatory mortality rate of a closely related species *Bosmina longirostris* derived
493 from sediment trap data in Lake Stechlin during steady-state of the population (Dubovskaya et al.,
494 2015) was lower: 0.015 d^{-1} . However, non-predatory mortality rates of *Daphnia* measured by
495 sediment traps in Lake Constance was of the wide range 0.002–0.18 d^{-1} (Gries and Güde, 1999) as
496 in our study. This suggests that the contribution of zooplankton carcasses to the overall carbon flux
497 and their ability to stimulate microbial organic matter degradation in each period of the growth
498 season was highly species-specific and depended on the current state of the populations and lake
499 ecosystem. In order to calculate the parameters $I_{DW\rho}$ and I_D of the released LOM, the model has to
500 rely on some assumptions that may need further explanation. One of the most important
501 assumptions made in the model is that there is no significant carcass destruction in the epilimnion.
502 A comparison of the normalized carcass fluxes of *D. cucullata* and *B. coregoni* for the 7 and 14 m
503 water layers shows that their destruction in the upper layers can be indeed neglected.
504 Underestimation of carcass destruction (if any) actually means an underestimation of the non-
505 predatory mortality for the species. However, with fixed sinking velocities, any increase in non-
506 predatory mortality values will lead to an increase in carcass destruction rates at lower depths
507 according to the model. As a result, the correction made may affect the absolute calculated values of
508 natural mortality and thus carcass destruction rates, but does not substantially affect the shape of the
509 vertical profile of LOM release from sinking carcasses. Consequently, our model results concerning

510 occurrence, relative location, and relative magnitude of LOM released from dead zooplankton
511 throughout the water column will be valid regardless of the assumptions made.

512 Our model predicts two peaks of organic matter release—one located in the epilimnion and
513 the other one in the metalimnion—both having different origins. The epilimnion peak of LOM
514 release is associated with the mortality of zooplankton primarily inhabiting the upper water layers
515 (e.g. *Daphnia* and *Bosmina*). It is formed by substances released by fresh zooplankton carcasses
516 during the first hours (up to one day) after death while maintaining carapace integrity. In the model,
517 this fraction is defined through the loss of carcass density $I_{DW\rho}$ and refers to the highly
518 biodegradable carbon fractions in dissolved or partially solid form. This fraction can supply
519 heterotrophs with the energy and nutrients needed to synthesize enzymes capable of degrading less
520 bioavailable material (Neubauer et al., 2021). The deeper metalimnetic peak is associated with
521 carcasses of *D. brachyurum* contributing to a local LOM release $I_{DW\rho}$ enforced by the flux of
522 organic particles (I_D) from the upper water layers. The latter fraction I_D represents the entry of
523 organic matter into the surrounding medium in form of fragmented carcass particles. Obviously, the
524 LOM fraction represented by I_D is of lesser quality, because of the higher proportion of chitin at the
525 late stages of microbial carcass degradation. However, fragments of the disintegrated carcasses
526 spread into a much larger volume of the water column, carrying attached microorganisms and
527 allowing them to get in contact with other organic particles in the water column. Additionally,
528 smaller particles may have lower excess density and, therefore, can be trapped in the density
529 gradient zone (thermocline) and involved in the local carbon cycle. Chitin, being a relatively hard-
530 to-degrade polysaccharide, still contains nitrogen molecules needed by bacteria. Therefore, under
531 conditions of nitrogen deficiency, it promotes development of specialized heterotrophs that can
532 decompose polysaccharides for nitrogen mining (see e.g. Bengtsson et al., 2018). This increases
533 microbial species diversity, which is necessary for a more effective mineralization of organic matter
534 with a low bioavailability in lakes.

535 The depth of formation of the second peak corresponds approximately to the depth of 8 m
536 (intersection of curves $I_{DW\rho}$ and I_D), which is the bottom of the thermocline. With the observed
537 composition and distribution of zooplankton, the most likely zones of carcass-induced increase in
538 interactive effects as determined by the release of LOM from dead zooplankton are localized either
539 in the epilimnion or in the metalimnion zone, i.e. at the thermocline density gradient.

540 The predicted zones of maximum LOM release do not necessarily correspond to the highest
541 microbial carbon mineralization rates in the water column (data of Franco-Cisterna et al., 2021), but
542 may represent the starting point for microbial organic matter remineralization and related interactive
543 (“priming”) effects (Kolmakova *et al.*, 2019; Neubauer *et al.*, 2021). In any case, our model

544 provides reliable estimates for scenarios of carcass LOM release and transfer into the surrounding
545 during carcass sinking throughout the stratified water column. It suggests an important role of
546 sinking zooplankton carcasses for the overall lake carbon cycle. Nevertheless, to better quantify
547 their significance, the formation of microbial aggregates and the interactions of carcass LOM with
548 organic material of different origins (e.g. dead phytoplankton and allochthonous particulates) and
549 their hydrophysical properties, i.e. buoyancy and accumulation at density gradients such as the
550 pycnocline, need to be investigated. The accumulation of carcass LOM and other organic matter in
551 distinct water layers as indicated by our model can potentially accelerate microbial mineralization
552 of both autochthonous and allochthonous organic matter and thus profoundly affect the carbon
553 source and sink functions of stratified lakes.

554

555 **Conclusions**

556 Sinking velocity, mortality rates, and degradation rates of cladoceran carcasses in the
557 stratified lake were species-specific. The model simulation allowed us to describe mechanisms of
558 LOM formation from zooplankton carcasses in a stratified lake and to determine vertical profiles of
559 LOM distribution, which appeared to vary in space and time. The model predicts a bimodal
560 occurrence of LOM release intensity peaks in relation to the presence and sinking of zooplankton
561 carcasses throughout the water column, with epilimnic and metalimnic peaks. The epilimnic LOM
562 peak was primarily formed by microbial LOM release from fresh zooplankton carcasses without
563 destroying carcass integrity. The metalimnic peak, however, comprised of LOM released by both
564 intact, fresh carcasses from the metalimnion but also aged, heavily degraded carcasses from the
565 epilimnion. This carcass LOM mixture potentially increases metalimnic LOM chemodiversity and
566 diversity of the microbial community, and likely intensifies microbial organic matter degradation.
567 Thus, LOM of zooplankton carcasses, in particular when accumulating at density layers such as the
568 pycnocline has the potential to change the carbon sink towards a carbon source function of stratified
569 lakes, which will be further amplified by current global warming.

570

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573

574 **Conflict of interest**

575 Authors declare no conflict of interest.

576

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581

582

583 **References**

584 Aarnos, H., Ylöstalo, P., and Vähätalo, A. (2012) Seasonal phototransformation of dissolved
585 organic matter to ammonium, dissolved inorganic carbon, and labile substrates supporting bacterial
586 biomass across the Baltic Sea. *J. Geophys. Res.: Biogeosci.*, **117**, 1004.

587 Benbow, M. E., Receveur, J. P., and Lamberti, G. A. (2020) Death and Decomposition in
588 Aquatic Ecosystems. *Front. Ecol. Evol.*, **8**, 17.

589 Benbow, M. E., Barton, P. S., Ulyshen, M. D., Beasley, J. C., DeVault, T. L., Strickland, M.
590 S., Tomberlin, J. K., Jordan, H. R., *et al.* (2019) Necrobiome Framework for Bridging
591 Decomposition Ecology of Autotrophically and Heterotrophically Derived Organic Matter. *Bull.*
592 *Ecol. Soc. Am.*, **100**, e01454.

593 Bengtsson, M. M., Attermeyer, K., and Catalán, N. (2018) Interactive effects on organic
594 matter processing from soils to the ocean: are priming effects relevant in aquatic ecosystems?
595 *Hydrobiologia*, **822**, 1–17.

596 Bickel, S. L., Tang, K. W., and Grossart, H.-P. (2009) Use of aniline blue to distinguish live
597 and dead crustacean zooplankton composition in freshwaters. *Freshwater Biol.*, **54**, 971–981.

598 Błędzki, L. A. and Rybak, J. I. (2016) Freshwater crustacean zooplankton of Europe:
599 Cladocera & Copepoda (Calanoida, Cyclopoida) key to species identification, with notes on
600 ecology, distribution, methods and introduction to data analysis. Springer International Publishing.

601 Daase, M., Varpe, Ø., Falk-Petersen, S. (2014) Non-consumptive mortality in copepods:
602 occurrence of *Calanus* spp. carcasses in the Arctic Ocean during winter. *J. Plankton Res.*, **36**, 129–
603 144.

604 Daase, M. and Søreide, J. E. (2021) Seasonal variability in non-consumptive mortality of
605 Arctic zooplankton. *J. Plankton Res.*, **43**, 565–585.

606 Diniz, L. P., França, E. J., Bonecker, C. C., Marcolin, C. R., Melo Júnior, M. (2021). Non-
607 predatory mortality of planktonic microcrustaceans (Cladocera and Copepoda) in neotropical
608 semiarid reservoirs. *Anais da Academia Brasileira de Ciências*, **93**.

609 Dubovskaya, O. P., Tolomeev, A. P., Kirillin, G., Buseva, Z., Tang, K. W., and Gladyshev,
610 M. I. (2018) Effects of water column processes on the use of sediment traps to measure zooplankton
611 non-predatory mortality: a mathematical and empirical assessment. *J. Plankton Res.*, **40**, 91–106.

612 Dubovskaya, O. P., Tang, K. W., Gladyshev, M. I., Kirillin, G., Buseva, Z., Kasprzak, P.,
613 Tolomeev, A. P., and Grossart, H.-P. (2015) Estimating In Situ Zooplankton Non-Predation
614 Mortality in an Oligo-Mesotrophic Lake from Sediment Trap Data: Caveats and Reality Check.
615 *PLoS ONE*, **10**, e0131431.

616 Elliott, D. T., Harris, C. K., and Tang, K. W. (2010) Dead in the water: The fate of copepod
617 carcasses in the York River estuary, Virginia. *Limnol. Oceanogr.*, **55**, 1821–1834.

618 Fonvielle, J. A., Giling, D. P., Dittmar, T., Berger, S. A., Nejtgaard, J. C., Lyche Solheim,
619 A., Gessner, M. O., Grossart, H., *et al.* (2021) Exploring the Suitability of Ecosystem Metabolomes
620 to Assess Imprints of Brownification and Nutrient Enrichment on Lakes. *J. Geophys. Res.:*
621 *Biogeosci.*, **126**.

622 Franco-Cisterna, B., Stief, P., and Glud, R. N. (2021) Temperature effects on carbon
623 mineralization of sinking copepod carcasses. *Mar. Ecol.: Prog. Ser.*, **679**, 31-45.

624 Giesecke, R., Vallejos, T., Sanchez, M., and Teiguie, K. (2017) Plankton dynamics and
625 zooplankton carcasses in a mid-latitude estuary and their contributions to the local particulate
626 organic carbon pool. *Cont. Shelf Res.*, **132**, 58–68.

627 Gladyshev M.I., Dubovskaya, O.P., Gubanov, V.G., and Makhutova O.N. (2003) Evaluation
628 of non-predatory mortality of two *Daphnia* species in a Siberian reservoir. *J. Plankton Res.*, **25**,
629 999-1003.

630 Gries, T. and Güde, H. (1999) Estimates of the nonconsumptive mortality of
631 mesozooplankton by measurement of sedimentation losses. *Limnol. Oceanogr.*, **44**, 459–465.

632 Gross, M. (2015) *Kernelheaping: kernel density estimation for heaped and rounded data*.
633 Freie Universitat Berlin.

634 Grossart, H. and Simon, M. (1998) Significance of limnetic organic aggregates (lake snow)
635 for the sinking flux of particulate organic matter in a large lake. *Aquat. Microb. Ecol.*, **15**, 115–125.

636 Håkanson, L. (1984) Suspension and calibration of a sediment trap. *Schweiz Z Hydrol.*, **46**,
637 171–175.

638 Halfter, S., Cavan, E. L., Butterworth, P., Swadling, K. M., and Boyd, P. W. (2021)
639 “Sinking dead”—How zooplankton carcasses contribute to particulate organic carbon flux in the
640 subantarctic Southern Ocean. *Limnol. Oceanogr.*, Ino.11971.

641 Johnston, S. E. Finlay, K., Spencer, R.G.M., Butman, D.E., Metz, M., Striegl, R., and
642 Bogard M.J. (2021) Zooplankton release complex dissolved organic matter to aquatic environments.
643 *Biogeochemistry*.

644 Kirillin, G., Grossart, H.-P., and Tang, K. W. (2012) Modeling sinking rate of zooplankton
645 carcasses: Effects of stratification and mixing. *Limnol. Oceanogr.*, **57**, 881–894.

646 Kolmakova, O., Gladyshev, M., Fonvielle, J., Ganzert, L., Hornick, T., and Grossart, H.-P.
647 (2019) Effects of zooplankton carcasses degradation on freshwater bacterial community
648 composition and implications for carbon cycling. *Environ. Microbiol.*, **21**, 34–49.

649 Koschel, R. and Adams, D. D. (2003) Preface: An approach to understanding a temperate
650 oligotrophic lowland lake (Lake Stechlin, Germany). *Arch. Hydrobiol., Spec. Issues Advanc.*
651 *Limnol.*, **58**, 1–9.

652 Kreling, J., Bravidor, J., Engelhardt, C., Hupfer, M., Koschorreck, M., and Lorke, A. (2017)
653 The importance of physical transport and oxygen consumption for the development of a
654 metalimnetic oxygen minimum in a lake: Metalimnetic oxygen minimum. *Limnol. Oceanogr.*, **62**,
655 348–363.

656 McCauley, E. (1984) The estimation of the abundance and biomass of zooplankton in
657 samples. *A manual on methods for the assessment of secondary productivity in freshwaters*.
658 Blackwell Sci. Publishers, London, pp. 228–265.

659 Moran, M. A. and Zepp, R. G. (1997) Role of photoreactions in the formation of
660 biologically labile compounds from dissolved organic matter. *Limnol. Oceanogr.*, **42**, 1307–1316.

661 Neubauer, D., Kolmakova, O., Woodhouse, J., Taube, R., Mangelsdorf, K., Gladyshev, M.,
662 Premke, K., and Grossart, H.-P. (2021) Zooplankton carcasses stimulate microbial turnover of
663 allochthonous particulate organic matter. *ISME J*, **15**, 1735–1750.

664 R Core Team (2021) *R: A language and environment for statistical computing*. R
665 Foundation for Statistical Computing, Vienna, Austria.

666 Rojas-Jimenez, K., Araya-Lobo, A., Quesada-Perez, F., Akerman-Sanchez, J., Delgado-
667 Duran, B., Ganzert, L., Zavialov, P. O., Alymkulov, *et al.* (2021) Variation of bacterial
668 communities along the vertical gradient in Lake Issyk Kul, Kyrgyzstan. *Environ. Microbiol.*
669 *Reports*, **13**, 337–347.

670 Scholtysik, G., Dellwig, O., Roeser, P., Arz, H. W., Casper, P., Herzog, C., Goldhammer
671 T., Hupfer M. (2020) Geochemical focusing and sequestration of manganese during eutrophication
672 of Lake Stechlin (NE Germany). *Biogeochemistry*, **151**, 313–334.

673 Schram, M. D. and Marzolf, G. R. (1994) Metalimnetic Oxygen Depletion: Organic Carbon
674 Flux and Crustacean Zooplankton Distribution in a Quarry Embayment. *Transactions of the*
675 *American Microscopical Society*, **113**, 105–116.

676 Selmeczy, G. B., Abonyi, A., Krienitz, L., Kasprzak, P., Casper, P., Telcs, A., Somogyvári,
677 Z., Padisák J. (2019) Old sins have long shadows: climate change weakens efficiency of trophic
678 coupling of phyto- and zooplankton in a deep oligo-mesotrophic lowland lake (Stechlin,
679 Germany)—a causality analysis. *Hydrobiologia*, **831**, 101–117.

680 da Silva, A. J., de Melo, P. A. M. C., Neumann-Leitão, S., and de Melo Júnior, M. (2020)
681 Non-predatory mortality of planktonic copepods in a reef area influenced by estuarine plume.
682 *Marine Environmental Research*, **159**, 105024.

683 Simon, M., Grossart, H., Schweitzer, B., and Ploug, H. (2002) Microbial ecology of organic
684 aggregates in aquatic ecosystems. *Aquat. Microb. Ecol.*, **28**, 175–211.

685 Tang, K., Bickel, S., Dziallas, C., and Grossart, H. (2009) Microbial activities
686 accompanying decomposition of cladoceran and copepod carcasses under different environmental
687 conditions. *Aquat. Microb. Ecol.*, **57**, 89–100.

688 Tang, K. W., Backhaus, L., Riemann, L., Koski, M., Grossart, H.-P., Munk, P., and Nielsen,
689 T. G. (2019) Copepod carcasses in the subtropical convergence zone of the Sargasso Sea:
690 implications for microbial community composition, system respiration and carbon flux. *J. Plankton*
691 *Res.*, **41**, 549–560.

692 Tang, K. W., Hutalle, K. M. L., and Grossart, H. (2006a) Microbial abundance, composition
693 and enzymatic activity during decomposition of copepod carcasses. *Aquat Microb Ecol*, **45**, 219–
694 227.

695 Tang, K. W., Freund, C. S., and Schweitzer, C. L. (2006b) Occurrence of copepod carcasses
696 in the lower Chesapeake Bay and their decomposition by ambient microbes. *Estuarine, Coastal and*
697 *Shelf Science*, **68**, 499–508.

698 Tang, K. W., Gladyshev, M. I., Dubovskaya, O. P., Kirillin, G., and Grossart, H.-P. (2014)
699 Zooplankton carcasses and non-predatory mortality in freshwater and inland sea environments. *J.*
700 *Plankton Res.*, **36**, 597–612.

701 Tolomeev, A. P., Kirillin, G., Dubovskay, O. P., Buseva, Z. F., and Gladyshev, M. I. (2018)
702 Numerical Modeling of Vertical Distribution of Living and Dead Copepods *Arctodiaptomus salinus*
703 in Salt Lake Shira. *Contemp. Probl. Ecol.*, **11**, 543–550.

704 Tranvik, L. and Bertilsson, S. (2008) Contrasting effects of solar UV radiation on dissolved
705 organic sources for bacterial growth. *Ecology Letters*, **4**, 458–463.

706 Wetzel, R. G. (2001) *Limnology: lake and river ecosystems*. 3rd ed. Academic Press, San
707 Diego.

708 Wickham H., Averick M., Bryan J., Chang W., McGowan L.D., François R., Golemund G.,
709 Hayes A., et al. Welcome to the tidyverse. *Journal of Open Source Software*, **4**, 1686. doi:
710 10.21105/joss.01686.

711 Winslow, L., Read, J., Woolway, R., Brentrup, J., Leach, T., Zwart, J., Albers, S., and
712 Collinge, D. (2019) *rLakeAnalyzer: Lake Physics Tools*.

713 Wologo, E., Shakil, S., Zolkos, S., Textor, S., Ewing, S., Klassen, J., Spencer, R. G. M.,
714 Podgorski, D. C., et al. (2021) Stream Dissolved Organic Matter in Permafrost Regions Shows

715 Surprising Compositional Similarities but Negative Priming and Nutrient Effects. *Global*
716 *Biogeochem. Cycles*, **35**.

717 Yue, Y., Cai, L., Tang, Y., Zhang, Y., Yang, M., and Wang, F. (2021) Vertical Distribution
718 of Bacterial Community in Water Columns of Reservoirs With Different Trophic Conditions
719 During Thermal Stratification. *Front. Environ. Sci.*, **9**, 632089.

720 **Table and Figure legends**

721 Table 1. Average length, dry weight, biomass of live and dead zooplankton, sinking
722 velocities and normalized fluxes of dead cladocerans in Lake Stechlin. Significance of differences
723 in values between 7 m and 14 m were assessed by t-test.

724 Table 2. Average natural mortality rate, maximum carcass decomposition rates at 7 - 14 m,
725 and hypothetical sinking velocity at 14 m (model), corresponding to $D \approx 0$.

726 Fig. 1. Map of Lake Stechlin with sampling location.

727 Fig. 2. Example of calculated sinking velocity $v_{D0}(z)$ under the condition $D(z) \approx 0$ and
728 interpolated sinking velocity $v(z)$ by Equation 17 through the actual velocities determined via
729 sediment traps at 7 and 14 m depths for *Daphnia* on 1st August, 2017.

730 Fig. 3. Variations of vertical distributions of live and dead *D. cucullata*, variations of their
731 abundance per m^2 and the carcass sinking flux at 7 and 14 m as estimated via sediment traps.

732 Fig. 4. Variations of vertical distributions of live and dead *B. coregoni*, the variations of
733 their abundance per m^2 and carcass sinking flux at 7 and 14 m as estimated via sediment traps.

734 Fig. 5. Variations of vertical distributions of live and dead *D. brachyurum*, the variations of
735 their abundance per m^2 and carcass sinking flux at 7 and 14 m as estimated via sediment traps.

736 Fig. 6. Variations of vertical distributions of live and dead *C. sphaericus*, the variations of
737 their abundance per m^2 and carcass sinking flux at 7 and 14 m as estimated via sediment traps.

738 Fig. 7. Distributions (5-days averages) of live and dead *D. cucullata*, *B. coregoni*, and *D.*
739 *brachyurum* individuals and their corresponding fitted carcass sinking velocities (v) and degradation
740 rates (D).

741 Fig. 8. Vertical distribution of total biomass of live and dead *D. cucullata*, *B. coregoni*, and
742 *D. brachyurum* in Lake Stechlin (27th July to 1st August, 2017), as well as their corresponding
743 model calculations of LOM release into the water column as I_{DWp} and I_D and their sum.

744

745 Table 1

Parameter	<i>D. cucullata</i>	<i>B. coregoni</i>	<i>D. brachyurum</i>	<i>C. sphaericus</i>
Length (mm) ± SE	0.714 ± 0.010	0.430 ± 0.006	0.723 ± 0.008	0.267 ± 0.003
Dry weight DW (μg) ± SE	1.91 ± 0.072	1.92 ± 0.068	1.35 ± 0.043	0.78 ± 0.032
Biomass and percent contribution in cladoceran biomass (mg m ⁻²)	566.67 ± 45.8 (49.71%)	101.44 ± 15.48 (8.9%)	345.61 ± 44.97 (30.32%)	126.29 ± 6.99 (11.08%)
Percentage of dead of each species	4.82 ± 0.48 %	8.91 ± 0.89 %	5.85 ± 0.88 %	4.59 ± 0.43 %
Measured sinking velocities v , m d ⁻¹				
7 m	2.15 ± 0.40	14.01 ± 0.98	1.10 ± 0.20	17.52 ± 2.40 ^a
14 m	7.64 ± 1.62	19.53 ± 1.92	5.88 ± 1.44	9.31 ± 1.31
t-test	3.30	2.55	3.28	-
Normalized flux \bar{F} , d ⁻¹				
7 m	0.007 ± 0.001	0.134 ± 0.011	0.017 ± 0.004	-
14 m	0.006 ± 0.001	0.110 ± 0.009	0.003 ± 0.001	-
Percentage of flux reduction between water layers	14.3%	17.9%	82.3%	
t-test	0.94	1.72	3.43	-

746 ^a artifact of sinking velocity determination (*C. sphaericus*) at 7 m, explanation provided in the main text. Bolded t-
747 values represent significant differences ($p < 0.05$)

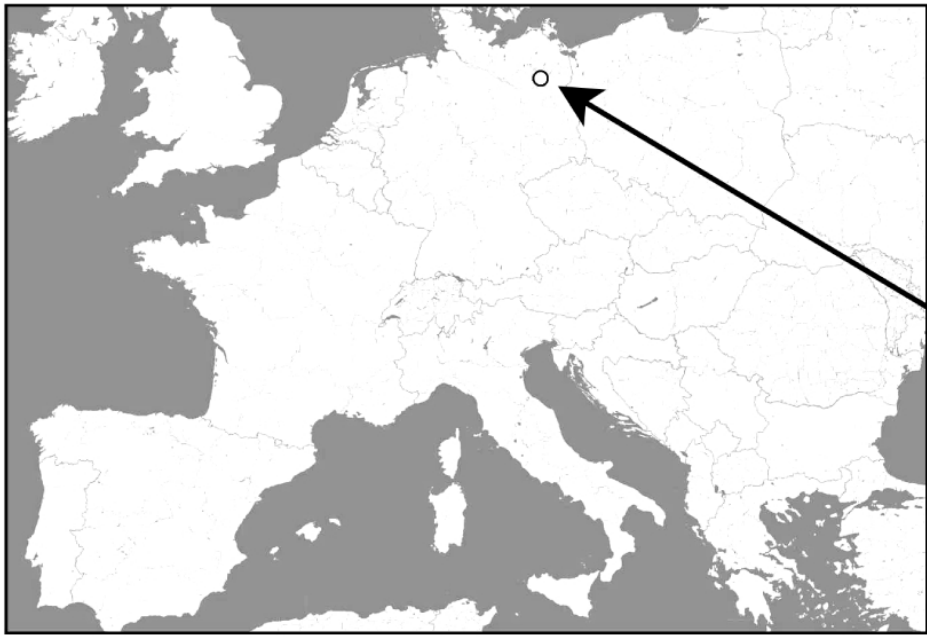
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749 Table 2.

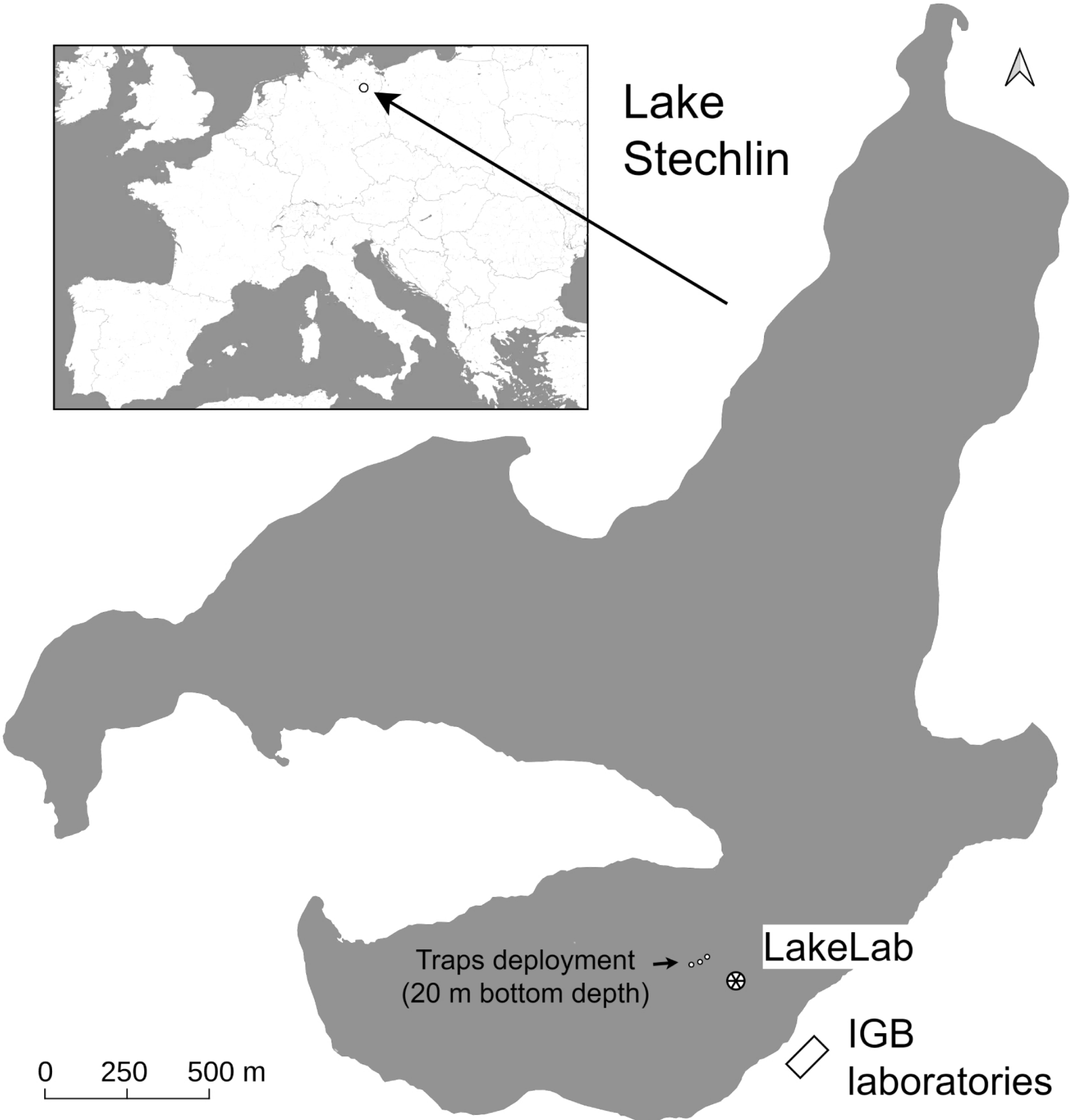
Fitted parameters	<i>D. cucullata</i>	<i>B. coregoni</i>	<i>D. brachyurum</i>
m – specific mortality rate (d^{-1})	0.008 ± 0.001	0.129 ± 0.012	0.020 ± 0.004
D_{max} – maximum specific destruction rate of carcasses in the layer 0 – 14 m (d^{-1})	0.362 ± 0.076	0.709 ± 0.263	3.990 ± 1.109
Hypothetic sinking velocities (m d^{-1}) at 14 m depth, calculated for the case $D \approx 0$	11.8 ± 1.9	23.7 ± 3.6	34.9 ± 9.5
Ratio of hypothetical sinking velocity (for $D \approx 0$) to observed one at 14 m	1.5	1.2	5.9
Contribution of species to total LOM (mg DW m^{-2})			
$I_{DW\rho}$ - fraction	2.18 ± 0.13 (25.6 %)	3.87 ± 0.42 (44.3 %)	2.6 ± 0.32 (30.1 %)
I_D - fraction	0.54 ± 0.15 (10.1 %)	1.57 ± 0.61 (27.5 %)	2.93 ± 0.54 (62.4 %)

750

751



Lake Stechlin



LakeLab

IGB laboratories

Traps deployment (20 m bottom depth)

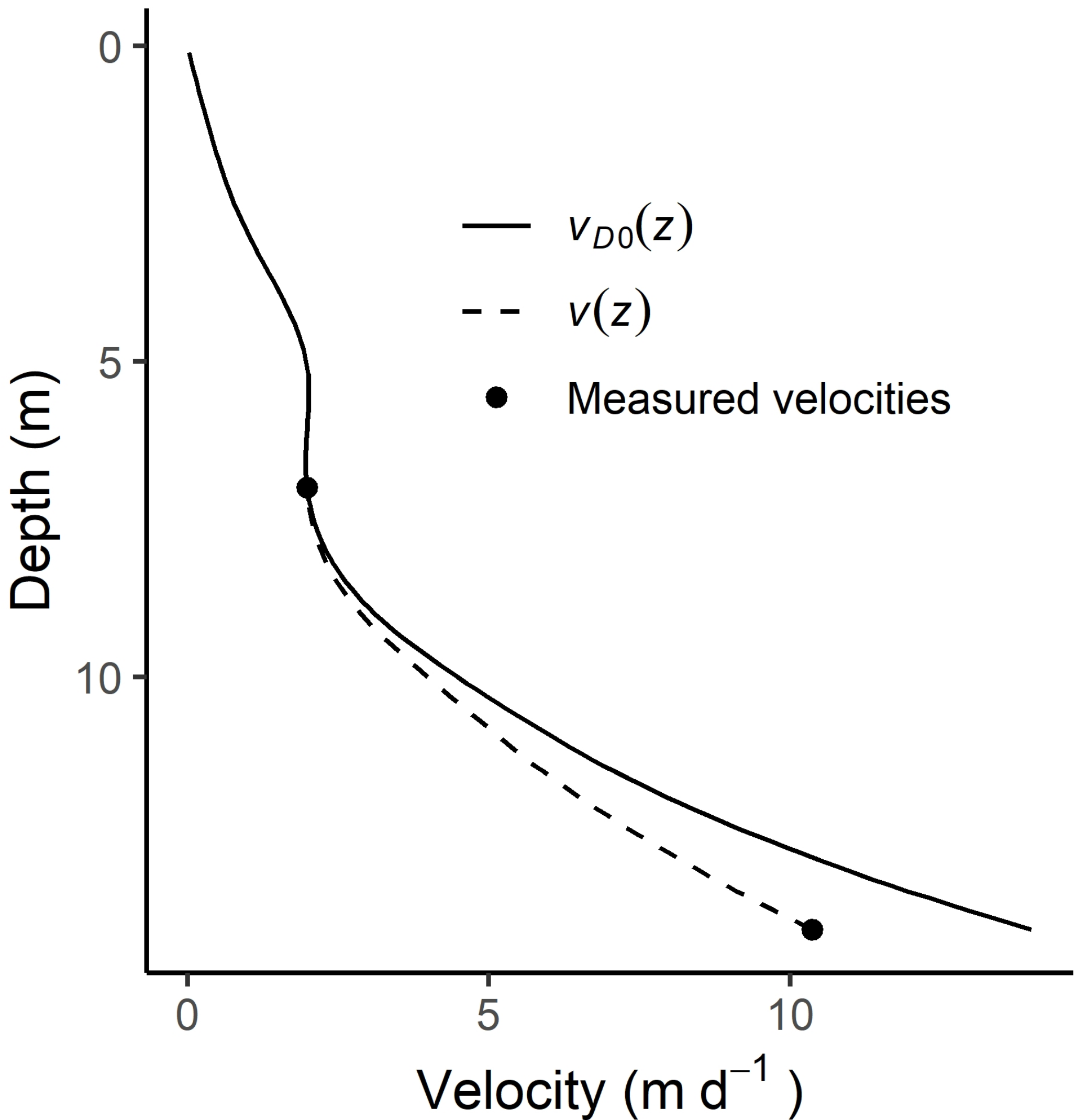
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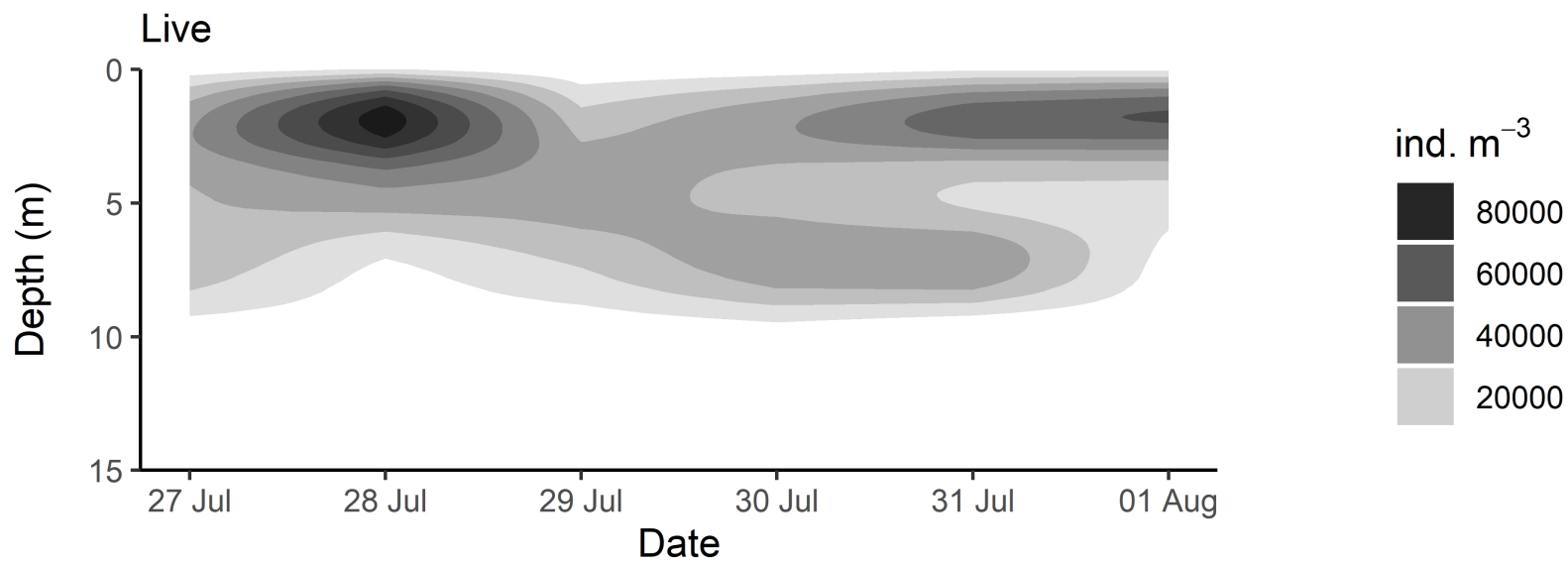
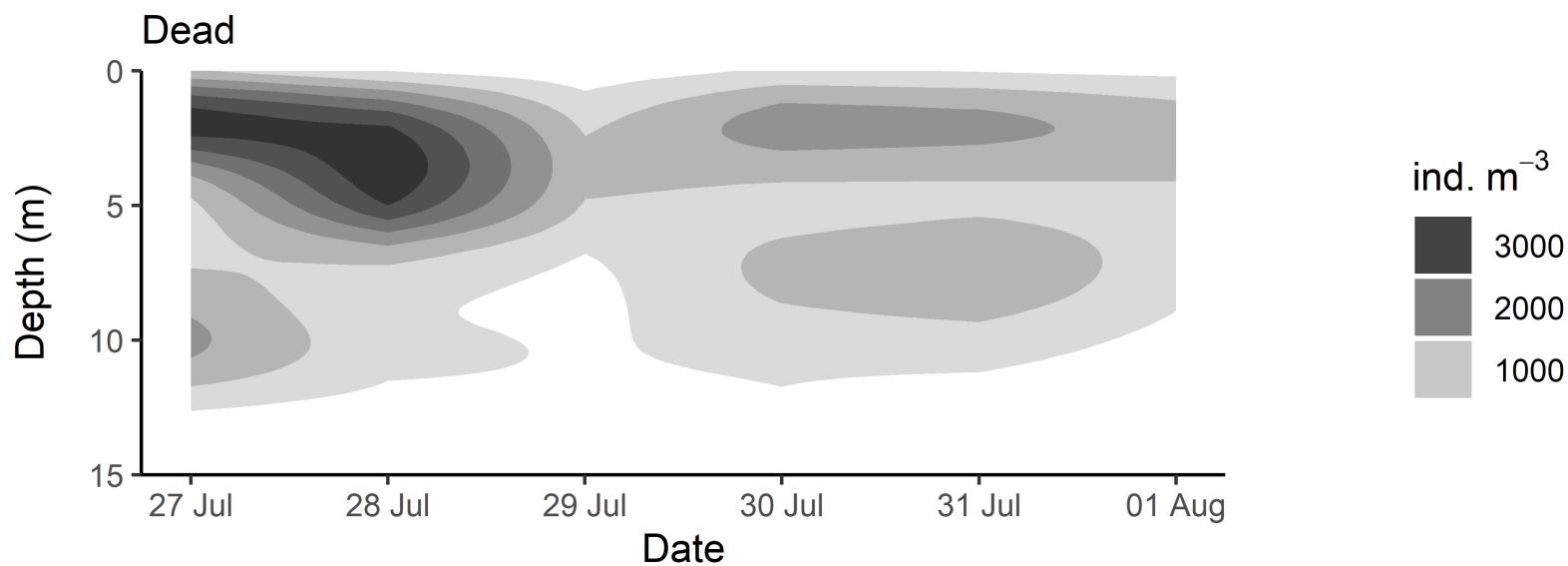
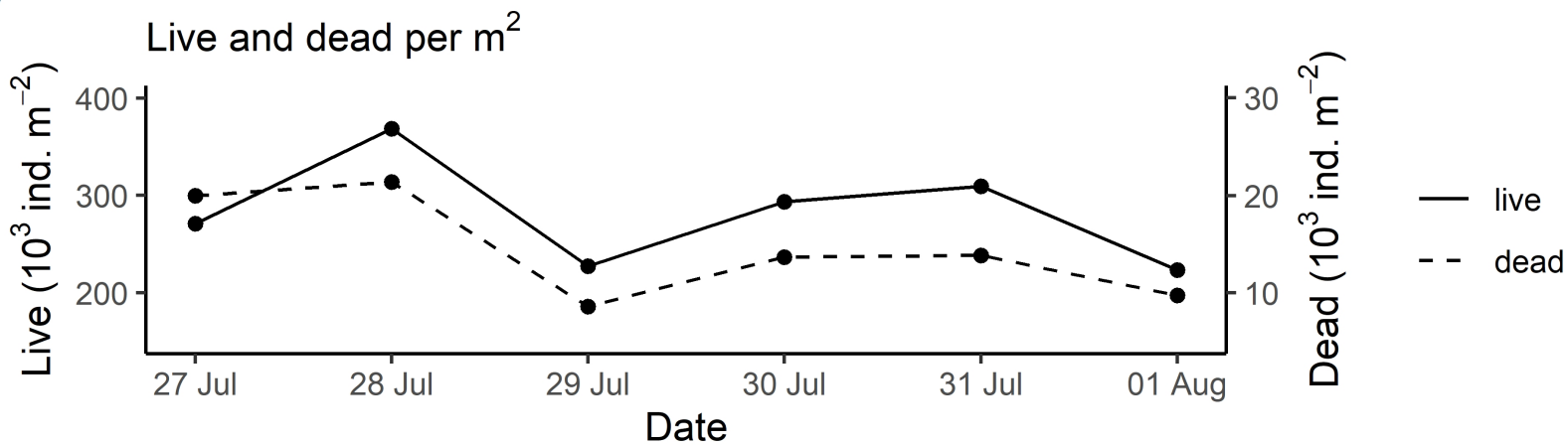
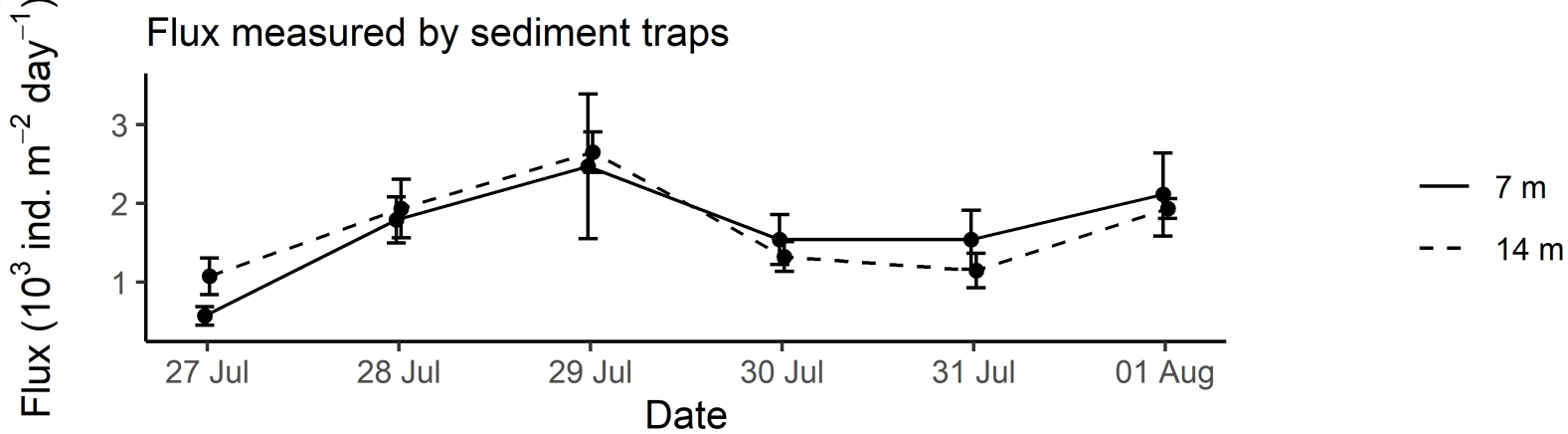
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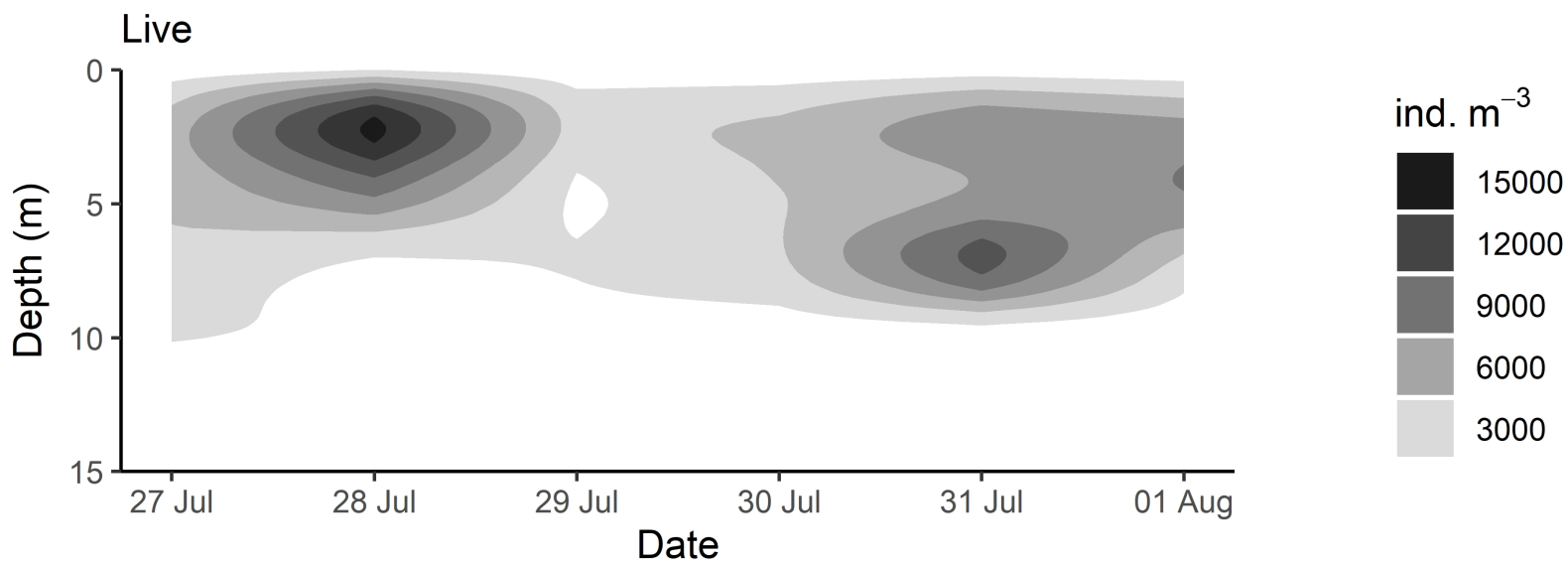
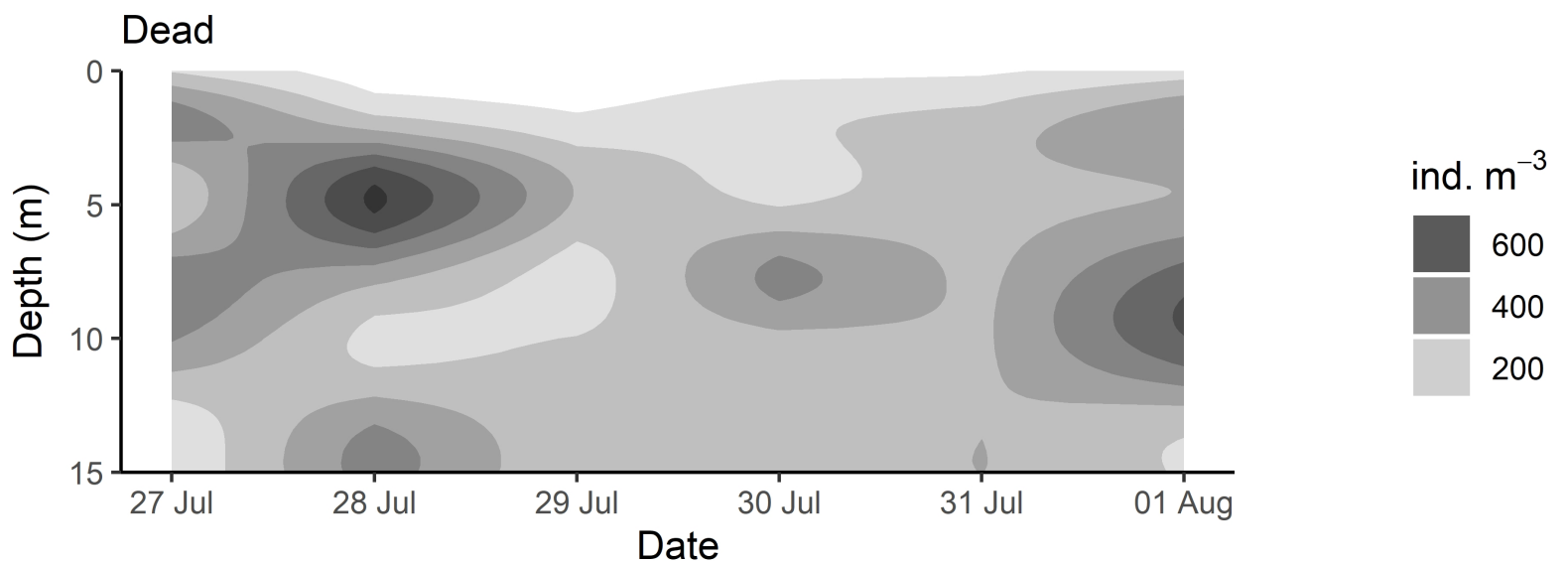
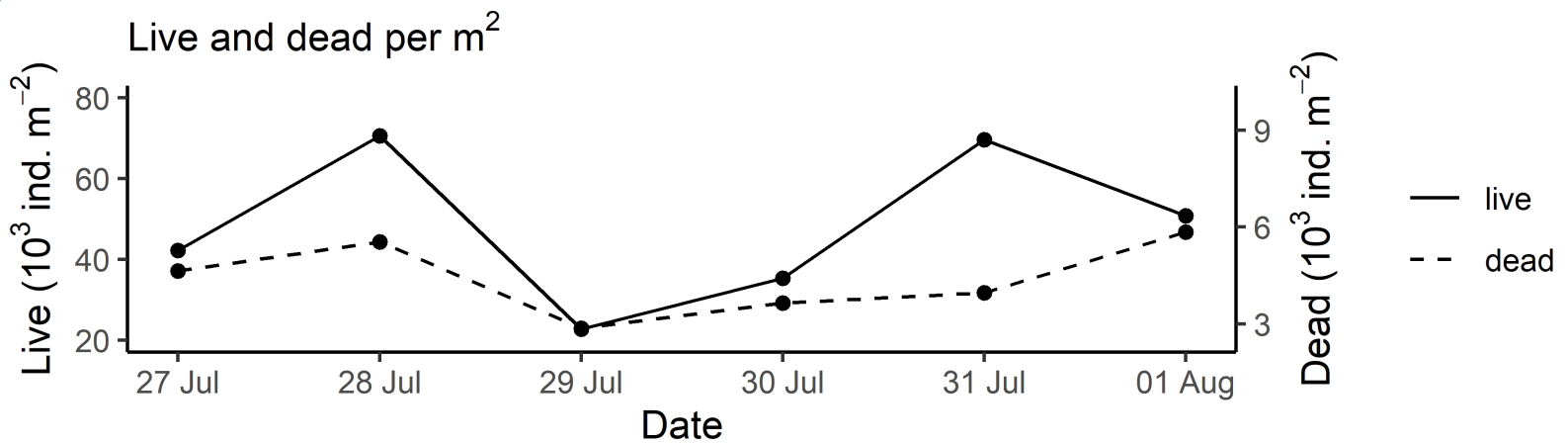
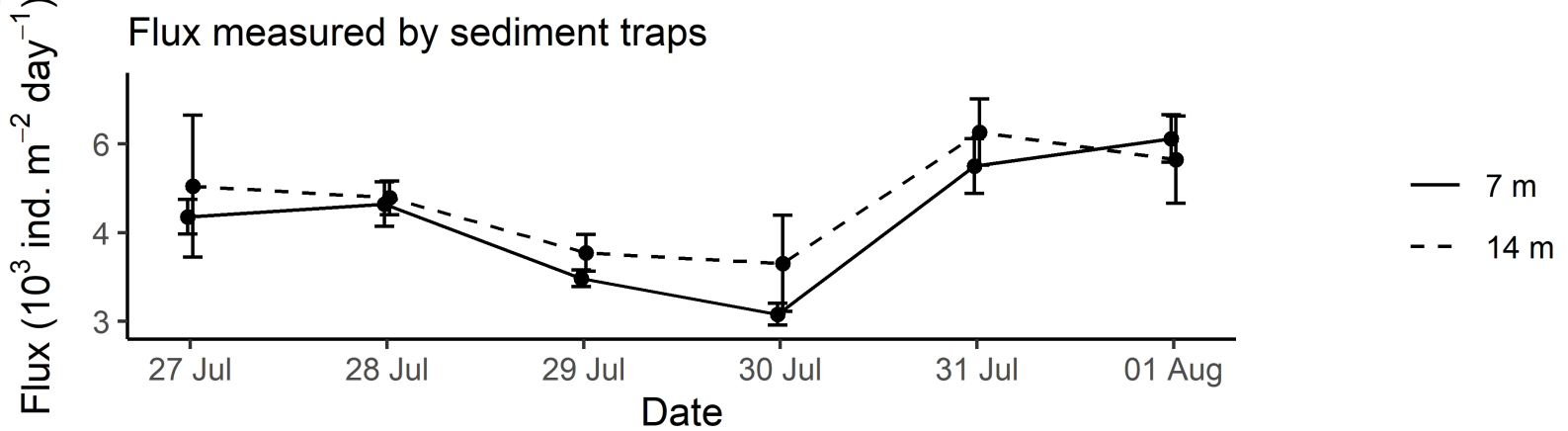
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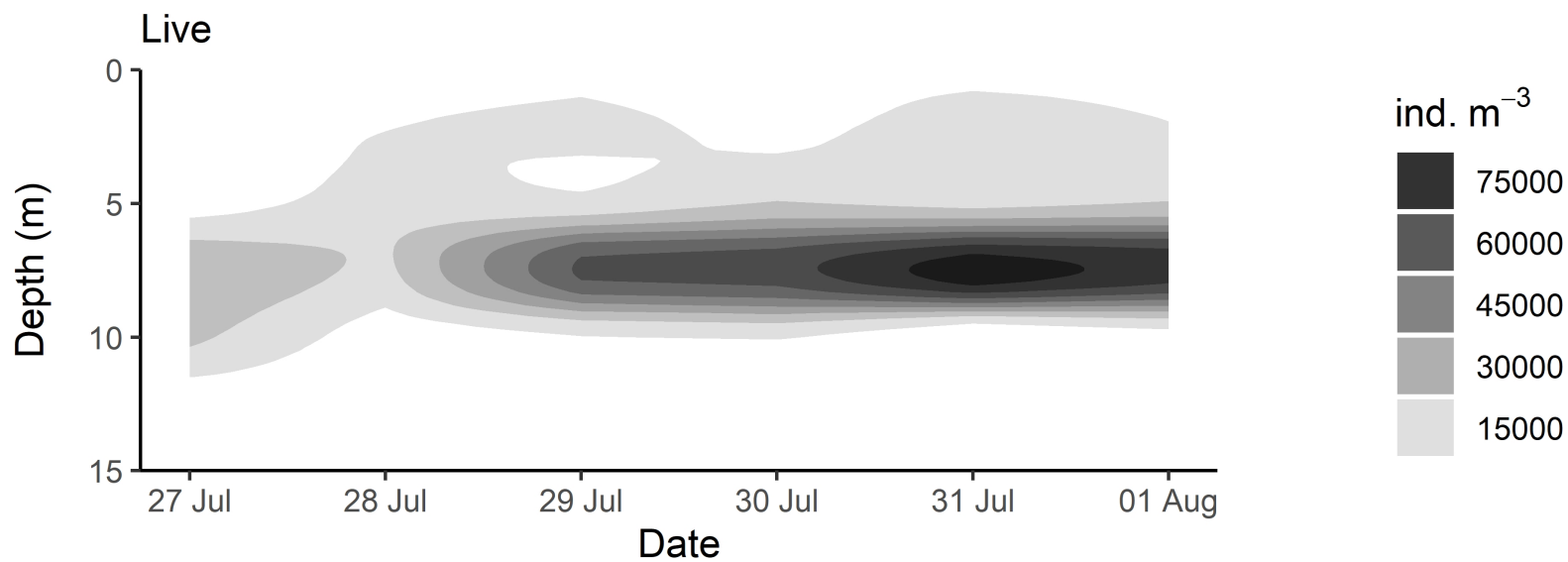
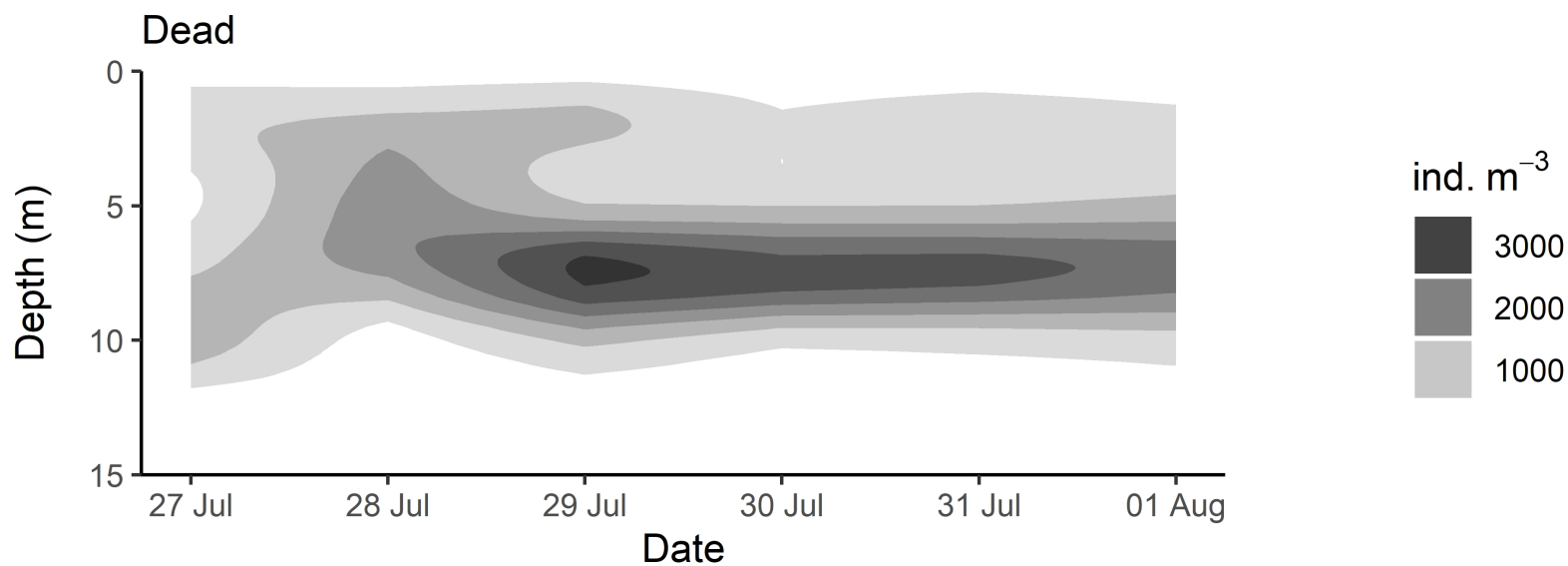
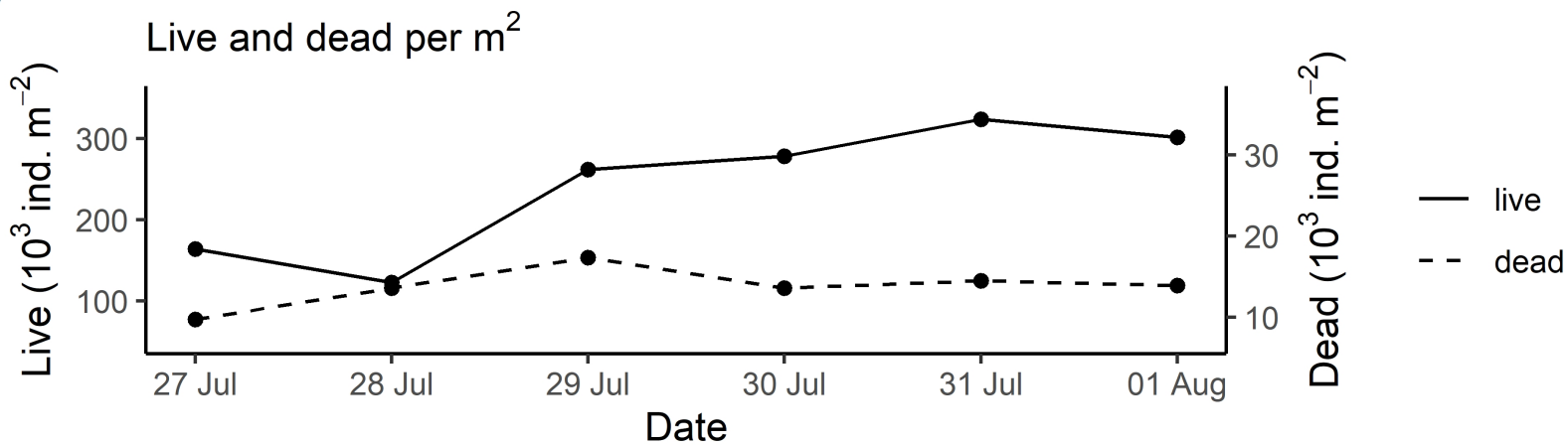
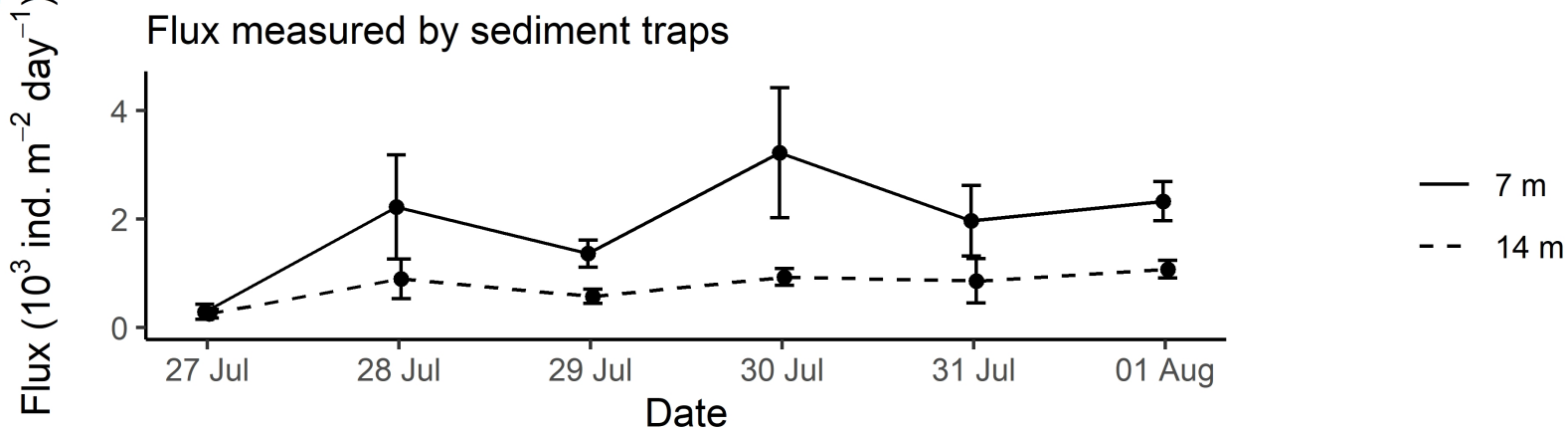
Daphnia cucullata

A**B****C****D**

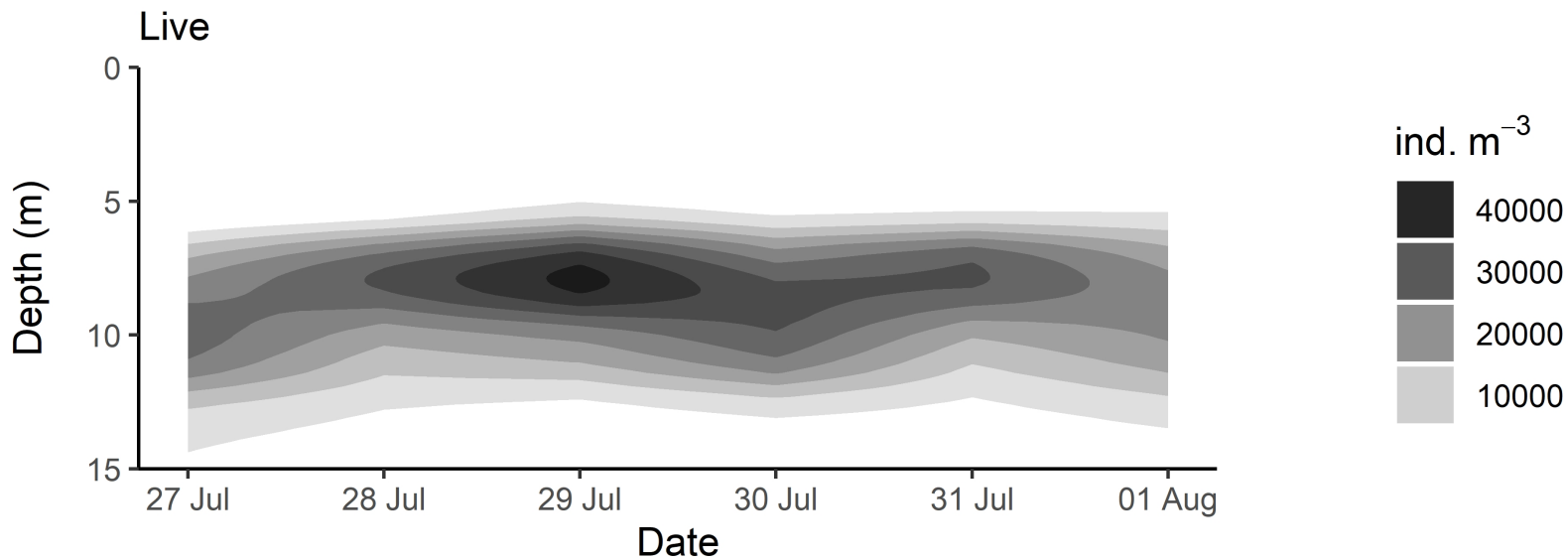
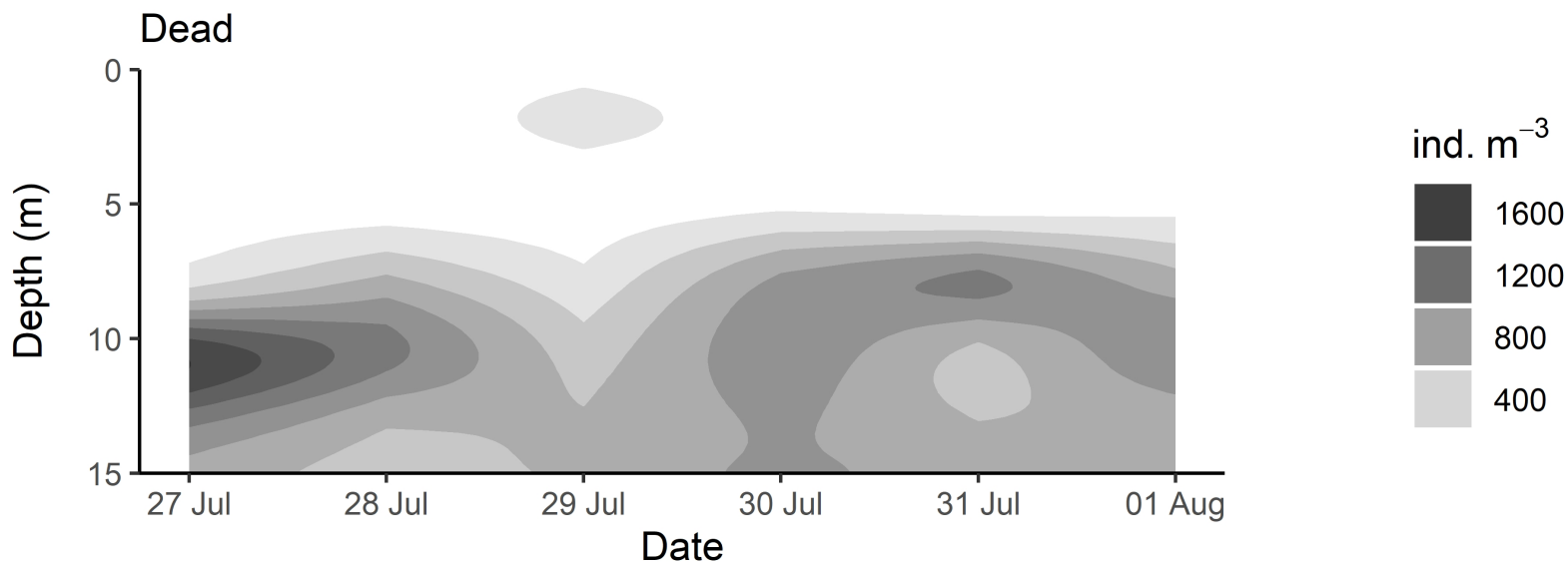
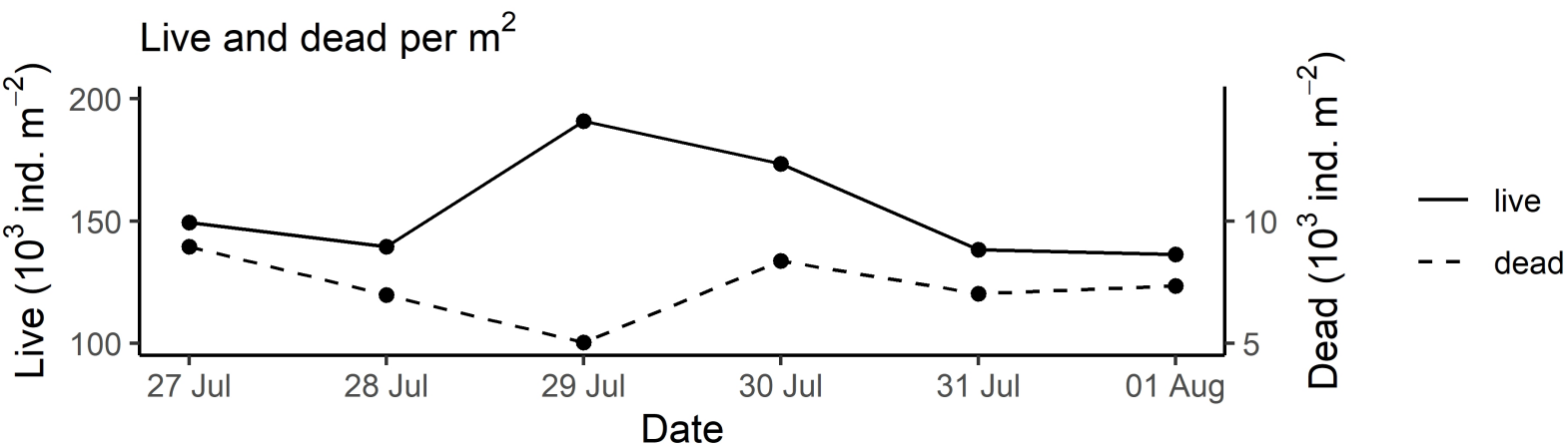
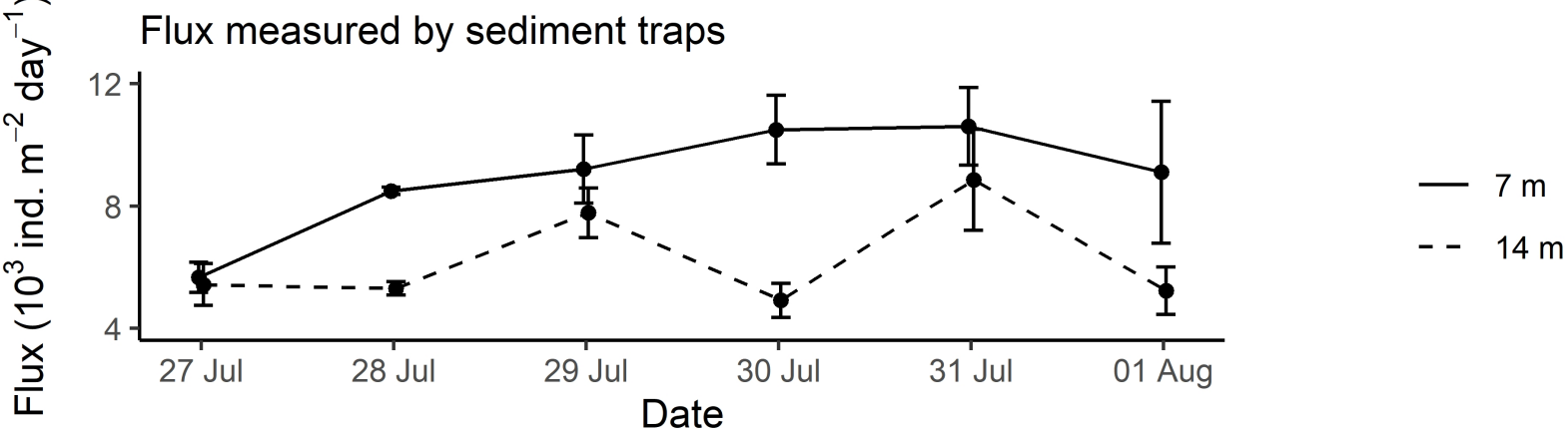
Bosmina coregoni

A**B****C****D**

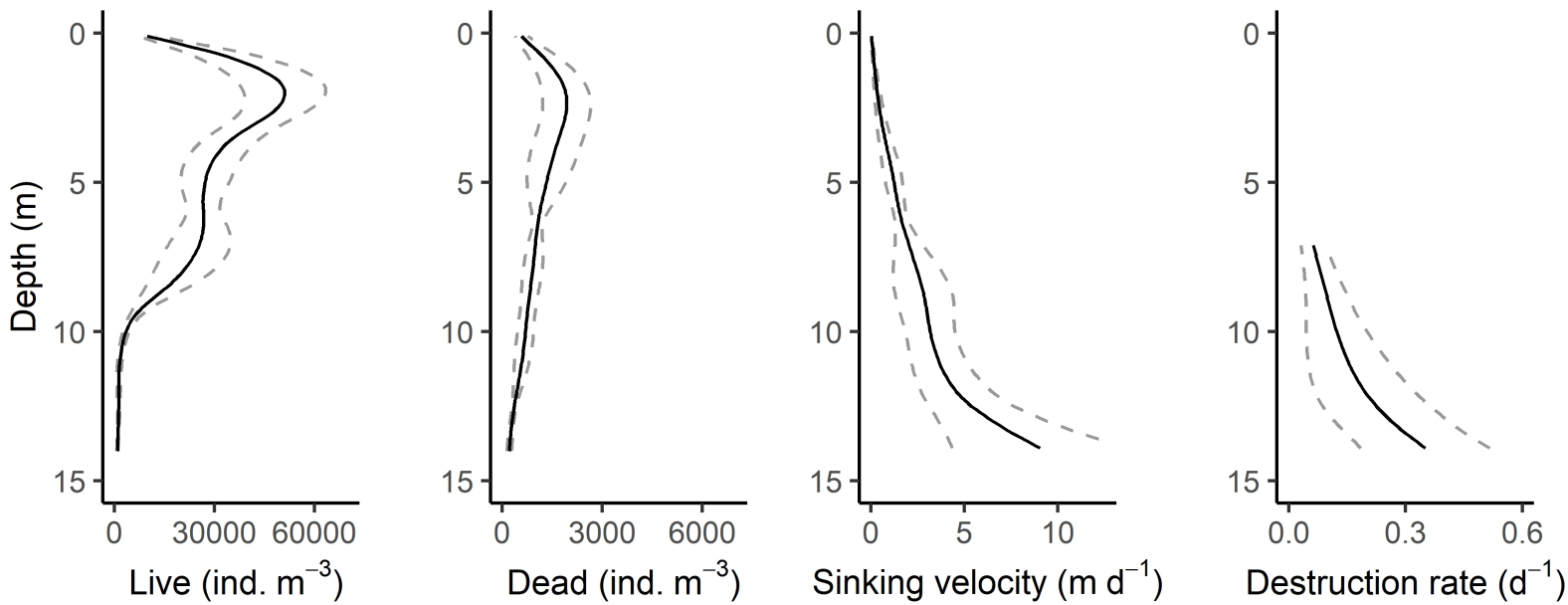
Diaphanosoma brachyurum

A**B****C****D**

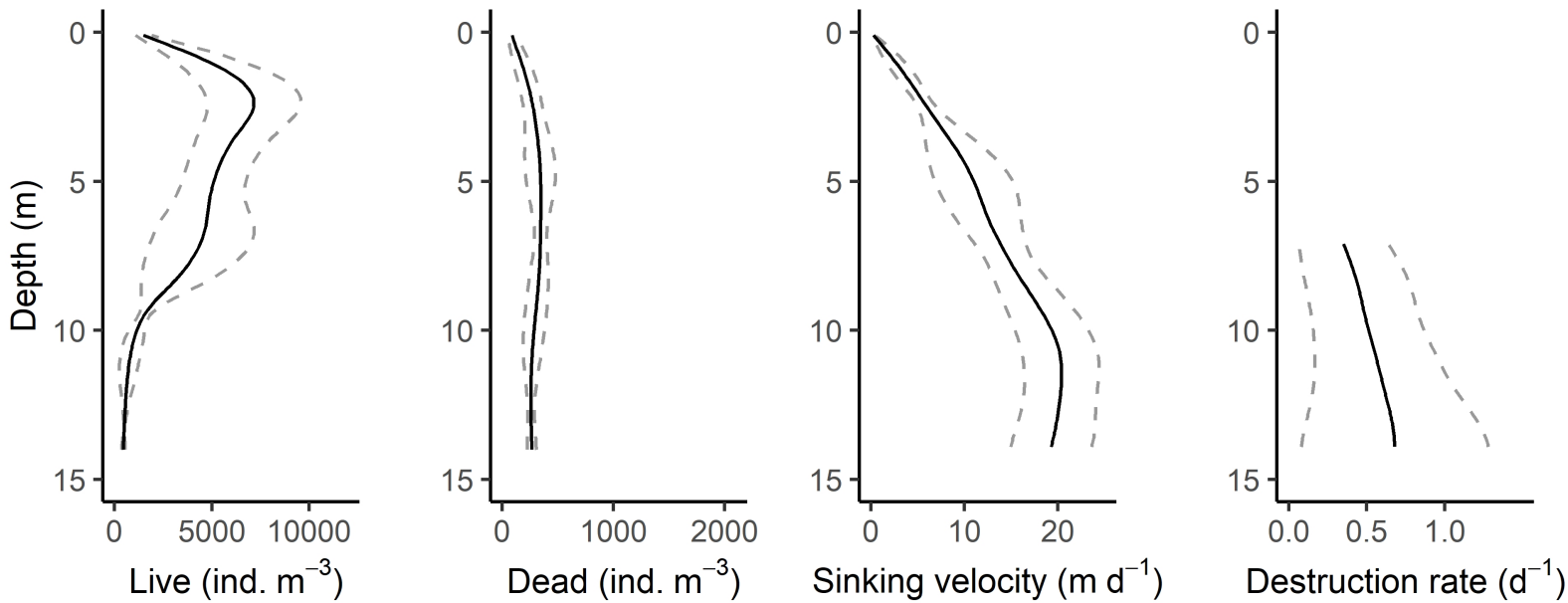
Chydorus cf. sphaericus

A**B****C****D**

Daphnia cucullata



Bosmina coregoni



Diaphanosoma brachyurum

