1 Diffusive CH₄ fluxes from aquaculture ponds using floating

- 2 chambers and thin boundary layer equations
- 3 Ping Yang^{a,b,1}, Jiafang Huang^{a,b,1}, Hong Yang^{c,d,e,1}, Josep Peñuelas^{f,g}, Kam W.
- 4 Tangh, Derrick Y.F. Lai^{t*}, Dongqi Wang^j, Qitao Xiao^k, Jordi Sardans^{f,g,**}, Yifei
- 5 Zhang^{a,b}, Chuan Tong^{a,b,***}
- 6 aKey Laboratory of Humid Subtropical Eco-geographical Process of Ministry of Education,
- 7 Fujian Normal University, Fuzhou 350007, P.R. China
- 8 bSchool of Geographical Sciences, Fujian Normal University, Fuzhou 350007, P.R. China
- 9 *College of Environmental Science and Engineering, Fujian Normal University, Fuzhou 350007,
- 10 P.R. China
- 11 dCollaborative Innovation Center of Atmospheric Environment and Equipment Technology,
- 12 Jiangsu Key Laboratory of Atmospheric Environment Monitoring and Pollution Control
- 13 (AEMPC), School of Environmental Science and Engineering, Nanjing University of Information
- 14 Science and Technology, Nanjing 210044, China;
- 16 *U.K.*
- 17 CSIC, Global Ecology Unit CREAF-CSIC-UAB, Bellaterra, Catalonia, Spain
- 18 gCREAF, Cderdanyola del Vallès, Catalonia, Spain
- 19 hDepartment of Biosciences, Swansea University, Swansea SA2 8PP, U. K.
- 21 Shatin, New Territories, Hong Kong SAR, China
- ¹School of Geographical Sciences, East China Normal University, Shanghai 200241, China
- 23 Key Laboratory of Watershed Geographic Sciences, Nanjing Institute of Geography and
- 24 Limnology, Chinese Academy of Sciences, Nanjing, 210008, China
- 25 *Correspondence to: Derrick Y.F. Lai
- 26 **Email:** dyflai@cuhk.edu.hk
- ***Correspondence to:** Jordi Sardans

- 28 **Email:** j.sardans@creaf.uab.cat
- 29 **Correspondence to: Chuan Tong
- 30 **Email:** tongch@fjnu.edu.cn
- ¹Ping Yang, Jiafang Huang, and Hong Yang contributed equally to this work.

32 HIGHLIGHTS

- 33 Aquaculture ponds emit CH₄
- Large variations in diffusive CH₄ fluxes are estimated by different thin boundary
- layer (TBL) models
- Methane fluxes measured by chambers and match those estimated by only some
- 37 TBL models

ABSTRACT

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Static floating chambers (FCs) are the conventional method to measure CH₄ fluxes 39 across the water-air interface in ponds, while thin boundary layer (TBL) modelling is 40 increasingly used to estimate CH₄ fluxes. In this study, both FCs measurements and 41 TBL models of gas transfer velocity were used to determine CH₄ evasion from 42 aquaculture ponds in southeastern China. The surface water CH₄ concentrations 43 ranged from 0.4 to 9.1 μ mol L⁻¹ with an average of 4.8 \pm 0.8 μ mol L⁻¹. CH₄ flux was 44 always positive, indicating the ponds as a persistent CH₄ source to air. Mean CH₄ flux 45 based on different TBL models showed large variations, ranging between 19 and 316 46 umol m⁻² h⁻¹. Compared against the direct measurement FCs, three TBL models 47 developed for the open sea, flowing estuarine system and lentic ecosystem (TBL_{W92a}, 48 49 TBL_{RC01}, and TBL_{CL98}, respectively) overestimated CH₄ emission by 40-200%, while the wind tunnel-based TBL model (TBL_{LM86}) underestimated CH₄ emission. Two TBL 50 models developed for lakes (TBL_{W92b} and TBL_{CW03}) gave estimates similar to FCs. 51 Keywords: Methane fluxes; Thin boundary layer models; Floating chambers; 52 Water-air interface; Shallow aquaculture pond; Subtropical estuary 53

1. Introduction

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Methane (CH₄) emissions from inland and coastal aquatic systems are potentially 55 significant sources of atmospheric CH₄ (Bastviken et al., 2011; Musenze et al., 2014; 56 Yang et al., 2011). CH₄ release from open water can be via diffusion and/or ebullition 57 (bubbling) (Bastviken et al., 2004). Diffusive fluxes across the water-air interface are 58 usually determined by using static floating chambers (FCs) or thin boundary layer 59 (TBL) models. The FCs approach determines CH₄ fluxes based on the change in CH₄ 60 concentrations in the chamber headspace over time. The TBL approach calculates the 61 62 CH₄ flux from piston velocity and gas concentration in the water (Natchimuthu et al., 63 2017; Zhao et al., 2019). Previous studies have used either one of the two approaches to quantify CH₄ fluxes from aquatic ecosystems (e.g., Musenze et al., 2014; 64 Natchimuthu et al., 2016; Wang et al., 2017; Welti et al., 2017). However, detailed 65 comparison of the two methods is rare (e.g., Duchemin et al., 1999; Matthews et al., 66 2003), particularly for small pond ecosystems. 67 Recent studies have shown that very small ponds (area <0.001 km²) are hotspots 68 for CH₄ emission (Holgerson, 2015; Holgerson and Raymond, 2016; Wik et al., 2016; 69 70 Yuan et al., 2019). However, the scalability of these measurements are largely 71 constrained by the lack of rigorous quantifications of the area, number, and spatial distribution of small ponds globally (Jonsson et al., 2008; Zhao et al., 2019) and the 72 different flux measurement methods between studies. In particular, the lack of 73 74 consensus on gas flux measurement methods remains a major source of uncertainty in greenhouse gas assessment. For instance, the TBL_{LM86}, TBL_{Wan92a} and TBL_{Wan92b}, 75

TBL_{RC01}, TBL_{CL98}, and TBL_{CW03} models, which were developed by Liss and Merlivatt 76 (1986), Wanninkhof (1992), Raymond and Cole (2001), Cole and Caraco (1998), and 77 78 Crusius and Wanninkhof (2003), respectively, are widely adopted wind-based models to estimate CH₄ transfer velocities and fluxes. Among these TBL models, the 79 80 TBL_{LM86}, TBL_{Wan92a}, and TBL_{RC01} models were developed for wind tunnels, open sea, 81 and flowing estuarine systems, respectively, while TB_{Wan92b} , TBL_{CL98} and TBL_{CW03} models were developed for the lentic ecosystem (e.g., lake). It is unclear to what 82 extent these different models are transferable to other aquatic ecosystems (Musenze et 83 84 al., 2014), and there is also a paucity of study comparing CH₄ fluxes by the different approaches. 85 Aquaculture ponds are an important component of the global inland aquatic 86 habitats (FAO, 2017), and the total surface area of freshwater and brackish 87 aquaculture ponds is estimated to be around 110,000 km² (Verdegem and Bosma, 88 2009). Despite the importance of aquaculture ponds for CH₄ emission (Hu et al., 2016; 89 Wu et al., 2018; Yang et al., 2015, 2019a; Yuan et al., 2019), relevant CH₄ flux data 90 are disproportionately scarce, and the published results were predominantly 91 determined by FCs rather than TBL modelling (Hu et al., 2016; Wu et al., 2018; Yang 92 et al., 2015, 2019a). In this study, FCs and TBL models were used to compare CH₄ 93 fluxes in aquaculture ponds in southeastern China. The aims were: (1) to evaluate the 94 performances of different wind-based TBL models for estimating CH₄ fluxes; (2) to 95 compare the diffusive CH₄ emissions from aquaculture ponds derived from FCs 96 measurements and TBL modellings; and (3) to assess which TBL model(s) can be used 97

to replace FCs for estimating CH₄ fluxes from ponds, with acceptable validity.

2. Materials and Methods

2.1. Study area

Our study sites are located at the central-western Shanyutan Wetlands in the Min River Estuary (MRE) in southeastern China (Figure S1, 26°00′36″–26°03′42″ N, 119°34′12″–119°40′40″ E). This area is characterized by a subtropical monsoon climate, with a multi-year annual average temperature and precipitation of 19.6 °C and 1,350 mm, respectively (Tong et al., 2010). The wetlands are dominated by a semidiurnal tide with a large tidal range (2.5-6 m) that follows a spring-neap-spring tidal cycle (Luo et al., 2014; Tong et al., 2010). The dominant vegetation are the native *Cyperus malaccensis* and *Phragmites australis*, and the invasive *Spartina alterniflora*. Over the past 10 years, much of area has been converted to aquacultural ponds (Yang et al., 2017a).

2.2. Aquaculture pond management

Small and shallow aquaculture earthen ponds (area of 0.8–2.5 ha and depth of 1.1–1.8 m) are a key feature in the MRE, covering a total area of around 234 ha in the Shanyutan Wetland (Yang et al., 2017b). Semi-intensive production is concentrated between June and November, which yields a single annual crop of shrimps. The ponds are filled with brackish water (average salinity of 2.0–8.5‰) from the MRE. The shrimps are fed twice daily (at 07:00 and 16:00 hr) with commercial aquatic feed pellets containing 42% protein. Three to five paddlewheel aerators operate four times

a day (07:00–09:00, 12:00–14:00, 18:00–20:00, and 00:00–03:00 hr) to provide oxygen. For this study, three ponds separated by <10 m (see Table S1 for basic characteristics) (Zhang et al., 2019) were selected for the measurements. Additional details about the shrimp pond system and management can be found in Yang et al. (2017b).

2.3. Determination of dissolved CH₄ concentration

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Field campaigns were carried out at the three ponds between June and November 2017 following the main aquaculture operation. In each pond, water and gas samples were collected at three sites along a foot-bridge that extended ~10 m from the embankment to the pond center. Samples were collected two or three times each month in each pond for a total of 15 times. The total number of samples was 3 ponds \times 3 sites \times 15 times = 135. To measure the dissolved CH₄ concentrations, surface water (at a depth of ~20 cm) was collected by a homemade water sampler and transferred into 55-mL gas-tight glass serum bottles that had been flushed with pond water 2-3 times. A 0.2 mL aliquot of saturated HgCl₂ solution was added to each bottle to inhibit bacterial activity of water sample (Borges et al., 2018; Hu et al., 2018), and the bottle was immediately sealed with a butyl rubber stopper and an aluminum screw cap to exclude air bubbles. Sample bottles were transported back to the laboratory in an ice-packed cooler. Dissolved CH₄ concentrations were measured within 2 d of collection using the headspace equilibration method: Approximately 25 mL of water in each bottle was displaced by N₂ gas (>99.999% purity) to create headspace. The bottle was then shaken vigorously for 20 min and left at room

- 141 temperature for 30 min to attain equilibrium between the air and the water phases (Cotovicz et al., 2016). Afterward, approximately 10 mL of the headspace was 142 143 extracted and injected into a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) equipped with a flame ionization detector (FID) to determine the CH₄ concentration. 144 145 Standard CH₄ gas (at 2, 8, 500, 1000, and 10,000 ppm) was used to calibrate the FID. 146 Dissolved CH₄ concentration was calculated based on the volume of water, headspace air and gas solubility coefficient for the specific water temperature and salinity (Farías 147 et al., 2017; Wanninkhof, 1992; Xiao et al., 2017). 148
- 149 *2.4. Determination of diffusive CH* $_4$ flux across the water-air interface
 - 2.4.1. Measurement using floating chambers

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- This study used a modified chamber placed on a floating buoy (Figure S2). The floating inverted opaque chamber was made from plastic basin (polyethylene/plexiglas®) with a volume and area of 5.2 L and 0.1 m², respectively. The chamber was covered with aluminum tape to minimize internal heating by sunlight (Natchimuthu et al., 2016; Yang et al., 2019). A thin gauze (pore diameter 0.001 mm) covering the opening minimized the entry of bubbles into the chamber (Figure S2). A fan inside the chamber mixed the headspace air during the sampling. In order to quantify the potential contribution of CH₄ ebullition flux from the ponds, total CH₄ fluxes were also determined using floating chamber without gauze.
- The chamber was deployed for a period of 45 min and headspace air samples
 being extracted at 15-min intervals (0, 15, 30, 45 min) using 60-mL syringes equipped

with three-way stopcocks. The gas samples were immediately transferred into pre-evacuated airtight gas sampling bags (Dalian Delin Gas Packing Co., Ltd., China), transported to the laboratory, and analyzed within 48 h using a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) equipped with a FID, following the method of Tong et al. (2010). The detection limit for CH₄ was 0.3 ppm, and the relative standard deviations of the measurements were $\leq 2.0\%$ in 24 h.

CH₄ emission flux (F_{CH4} , μ mol m⁻² hr⁻¹) was calculated from the slope of the regression between headspace CH₄ concentration and time (Yang et al., 2019). Generally, if r^2 of the regression is > 0.90, the CH₄ emission is considered as diffusion only (Bastviken et al., 2010; Zhu et al., 2016). If r^2 is < 0.90, the emission is considered as a combination of ebullition and diffusion. The floating chambers with gauze (FCs-G) and without gauze (FCs-NG) showed distinctly linear ($r^2>0.9$) and nonlinear $(r^2 < 0.9)$ increases in methane concentration, respectively; therefore, the contribution of ebullition could be calculated as the difference between the FCs-G and the FCs-NG measurements.

177 2.4.2. Estimation using thin boundary layer models

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178 Saturation (S) of CH₄ in pond water was calculated as (Hu et al., 2018):

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$$S = C_{water}/C_{Ws} = C_{water}/(\alpha \times C_{air}) \times 100\%$$
 (Eq. 1)

where C_{water} is dissolved CH₄ concentration in pond water; C_{Ws} is the saturated CH₄ 180 concentration (µmol L⁻¹); Cair is the atmospheric CH₄ concentration (µmol mol⁻¹) at the sampling site; and α is the Bunsen coefficient (Wanninkhof, 1992). 182

Diffusive flux of CH₄ (F, µmol m⁻² hr⁻¹) across the water-air interface can be

described by a theoretical diffusion model (Musenze et al., 2014):

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$$F = k \times (C_{\text{water}} - C_{eq})$$
 (Eq. 2)

- where C_{water} (µmol L⁻¹) is the measured dissolved CH₄ concentration in surface water,
- C_{eq} (µmol L⁻¹) is the dissolved CH₄ concentration in equilibrium with the air above,
- and k is the gas transfer velocity (cm h⁻¹). The k value was parameterized as a function
- of wind speed and normalized for surface water temperature $(T, {}^{\circ}C)$ using a Schmidt
- number (*Sc*) derived from Eq. 3 (Wanninkhof, 1992):

$$Sc = 2039.20 - 120.31T + 3.4209T^2 - 0.040437T^3$$
 (Eq. 3)

- 192 This study evaluated the variations in CH₄ fluxes estimated by eight widely used
- wind-based models developed for different environments, including wind tunnels,
- open sea, estuarine systems and lakes, as follows:
- 195 LM86 (Liss and Merlivatt 1986)

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$$F_{LM\,86} = 0.17 U_{10} (Sc / 600)^{-2/3} (C_{water} - C_{eq})$$
 $0 < U_{10} \le 3.6$ (Eq. 4)

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$$F_{LM\,86} = (2.85U_{10} - 9.65)(Sc/600)^{-1/2}(C_{water} - C_{eq})$$
 $3.6 < U_{10} \le 13$ (Eq. 5)

198 W92a (Wanninkhof, 1992)

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$$F_{W92a} = 0.31U_{10}^2 (Sc/660)^{-1/2} (C_{water} - C_{eg})$$
 (Eq. 6)

200 RC01 (Raymond and Cole, 2001)

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$$F_{RC01} = 1.91 exp(0.35U_{10})(Sc/600)^{-1/2}(C_{water} - C_{eq})$$
 (Eq. 7)

202 CL98 (Cole & Caraco, 1998)

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$$F_{CL98} = [2.07 + (0.215 \times U_{10}^{1.7})](Sc/600)^{-2/3}(C_{water} - C_{eq})$$
 (Eq. 8)

204 W92b (Wanninkhof, 1992)

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$$F_{W92b} = 0.45U_{10}^{1.64} (Sc/600)^{-1/2} (C_{water} - C_{eq})$$
 (Eq. 9)

206 CW03 (Crusius & Wanninkhof, 2003)

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$$F_{CW \, 03} = 0.72 U_{10} (Sc \, / \, 600)^{-2/3} (C_{water} - C_{eq})$$
 $U_{10} < 3.7$ (Eq. 10)

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$$F_{CW03} = (4.33U_{10} - 13.3)(Sc / 600)^{-1/2}(C_{water} - C_{eq})$$
 $U_{10} \ge 3.7$ (Eq. 11)

- In the above equations, U_{10} was determined according to the logarithmic wind profile
- relationship (Crusius and Wanninkhof, 2003):

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$$U_{10} = U_z \left[1 + \frac{(C_{d10})^{1/2}}{K} \ln(\frac{10}{z})\right]$$
 (Eq. 12)

212 where U_z is the wind speed (m s⁻¹) at height z above the water surface (2.5 m in this 213 study), C_{d10} is the drag coefficient at 10 m above the water surface (0.0013 m s⁻¹), and 214 K is the von Karman constant (0.41). Generally, the calculation of U_{10} is sensitive of 215 the stability of the atmosphere. If the atmosphere over the aquatic systems is unstable, 216 and the equation used to calculate U_{10} needs to be adjusted. The air-water temperature 217 difference can be used to determine the atmospheric stability; if the air-water 218 temperature difference is positive, the atmosphere is considered stable. During the 219 study period, the air temperature was 0.1-3.8 °C higher than water temperature, 220 indicating that the atmosphere over the ponds was largely in the stability regime. 221 Therefore, no adjustment was needed for U_{10} , and Eq. 12 was appropriate for 222 calculating U_{10} . Some recent studies have applied surface renewal models that take

- into account both wind speed and buoyancy to determine the k values (e.g., Czikowsky
- 224 et al., 2018; MacIntyre et al., 2010; MacIntyre et al., 2018).

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- 225 2.5. Measurement of meteorological and environmental variables
- Meteorological variables including air temperature (A_T), air pressure (A_P) and precipitation were recorded using an automatic meteorological station (Vantage Pro 2, China) installed at the MRE weather station in the China Wetland Ecosystem Research Network. The distance between the automatic meteorological station and sampling ponds is about 75 m. The precision for air temperature, atmospheric pressure, and precipitation were \pm 0.2 °C, \pm 1.5 hPa, and \pm 0.4 mm min⁻¹, respectively (Yang et al., 2020). Wind speed (W_S) was measured at 1 Hz at a resolution of 0.4 m
 - Water temperature, electrical conductivity (EC), pH, dissolved oxygen (DO), total organic carbon (TOC) and total dissolved nitrogen (TDN) content of surface water (~20 cm below the water surface) were measured at each sampling site in all sampling campaigns. Water temperature and pH were measured by a portable pH/mV/Temperature meter (IQ150, IQ Scientific Instruments, USA), and EC and DO by an electrical conductivity meter (2265FS EC, Spectrum Technologies, USA) and a multiparameter water quality probe (550A YSI, USA), respectively. The relative standard deviations of EC, pH, and DO measurements were □1.0%, □1.0% and □2.0%, respectively.
 - Water samples for TOC and TDN analyses were collected using a 5-L plexiglass

water sampler, transferred to a 150-mL polyethylene bottle, and then transported to the laboratory in an ice-packed cooler. The water samples were filtered through a 0.45- μ m cellulose acetate filter (Biotrans nylon membranes); the filtrates were then analysed by a TOC analyzer (TOC-V_{CPH/CPN}, Shimadzu, Kyoto, Japan) for TOC and a flow injection analyzer (Skalar Analytical SAN⁺⁺, The Netherlands) for NO₃⁻-N. The detection limits for NO₃⁻-N and TOC were 6 μ g L⁻¹ and 4 μ g L⁻¹, respectively. The relative standard deviations of NO₃⁻-N and TOC measurements were \Box 3.0% and \Box 1.0%, respectively.

2.6. Statistical analysis

Repeated-measures analysis of variance (RM-ANOVA) was conducted to test the differences in diffusive CH₄ flux between the different approaches over the study period. Pearson correlation analysis was used to examine the relationships between (1) dissolved CH₄ concentration or CH₄ fluxes and environmental variables, and (2) diffusive CH₄ fluxes measured using FCs and those estimated using the TBL models. The coefficient of variation (CV) for CH₄ fluxes on each sampling campaign was determined by dividing the standard deviation by the mean value. Statistical analyses were conducted using the software SPSS (v. 17.0, SPSS Inc., USA) and the significance level was set at p < 0.05. Data are presented as mean \pm 1 standard error. Generalized linear modelling was conducted to compare the variables that influenced CH₄ emission flux from the different methods (i.e. FCs + 8 TBL models).

The "gls" function from the "nlme" R package (Pinheiro et al., 2018) with a saturated

model was conducted for all variables (dissolved CH₄, U_{10} , water temperature, dissolved oxygen, total dissolved carbon and dissolved nitrate). This model was run using the stepAIC function in R "MASS" package that follows the Akaike Information Criterion (AIC) (Venables and Ripley, 2002). It was used to identify the best model (lowest AIC value) in each case.

3. Results

3.1. Meteorological and environmental variables

- The average air temperature (A_T) and air pressure (A_P) during the study were 28.7±0.4 °C (range: 18.6–35.6 °C) and 1010.0±0.5 hPa (range: 985–1025 hPa), respectively. Notably, the maximal A_T appeared in July and the minimal A_P happened in August, different from the other months. W_S ranged from 0.2 to 18.8 m s⁻¹, and it varied between seasons, with a peak in July (Figure S3a). Approximately 92% of W_S fell within the range of 0.2–4.0 m s⁻¹ (Figure S3b).
- There were temporal variations in surface water characteristics during the study
 period. The mean water temperature ranged from 18.1 °C (November) to 34.4 °C

 (August) (Figure S4a), while the mean DO concentration varied between 9.4 mg L⁻¹

 (August) and 19.9 mg L⁻¹ (November) (Figure S4). The mean TOC concentration
 varied between 9.9 mg L⁻¹ (July) and 57.3 mg L⁻¹ (November) (Figure S3), while

 NO₃-N concentrations ranged from 504 μg N L⁻¹ (June) to 10.7 μg N L⁻¹ (November)

 (Figure S4).
- 285 3.2. Model estimated k values and dissolved CH₄ concentrations

- The mean k value showed considerable variations between models: k_{RC01}
- 287 $(6.5\pm0.8 \text{ cm h}^{-1}) > k_{\text{W92a}} (3.5\pm0.7 \text{ cm h}^{-1}) > k_{\text{FCs}} (3.2\pm0.4 \text{ cm h}^{-1}) > k_{\text{CL98}} (2.9\pm0.3 \text{ cm}^{-1})$
- 288 h^{-1}) > k_{CW03} (2.5±0.5 cm h^{-1}) > k_{W92b} (2.4±0.4 cm h^{-1}) > k_{LM86} (0.6±0.1 cm h^{-1}) (Figure
- 289 1).
- Dissolved CH₄ concentration varied considerably during the study period
- 291 (0.1-31.1 μmol L⁻¹), with a large increase between June and August, followed by a
- small decrease toward November (Figure 2). The water was supersaturated in CH₄ in
- 293 all ponds and on all sampling dates, with an overall mean of $4.8 \pm 0.8 \,\mu\text{mol} \,L^{-1}$ (162.0
- ± 18.4 ppmv), equivalent to 8700% saturation (range of $200 5.9 \times 10^4$ % saturation).
- 295 3.4. CH₄ flux estimates by using TBL models and FCs method
- There were considerable differences in the estimated diffusive CH₄ fluxes among
- 297 the *TBL* models (*TBL*_{RC01}: 215.9 \pm 39.2 μ mol m⁻² h⁻¹; *TBL*_{CL98}: 115.0 \pm 21.9 μ mol m⁻²
- 298 h^{-1} ; TBL_{W92a} : 102.9 \pm 19.5 μ mol m^{-2} h^{-1} ; TBL_{W92b} : 78.3 \pm 13.9 μ mol m^{-2} h^{-1} ; TBL_{CW03} :
- 299 74.9 ±13.2 μmol m⁻² h⁻¹; and, TBL_{LM86} : 19.5 ±3.7 μmol m⁻² h⁻¹) (Table 1, Figure 3 and
- Figure S5). Although there were marked variations in the flux estimates among the
- various models, results from all models showed similar temporal patterns (Figure 3).
- The largest fluxes were generally recorded between August and October, while the
- lowest fluxes were consistently recorded in June and November (Figure 3).
- Direct measurements using FCs with gauze (FCs-G) and without gauze
- 305 (FCs-NG) methods were 75.0 ± 12.5 (Figure 3) and 2231.3 ± 681.3 µmol m⁻² h⁻¹
- 306 (Figure S6; Yang et al., unpublished data), showing significant difference between the

two methods (Independent Samples T-Test, F = 118.190, p<0.001). On average, ebullitive CH₄ flux accounted for 33%-99% of the total CH₄ emissions during the study period.

3.5. Environmental influences on dissolved CH₄ concentrations and fluxes

Pearson correlation analysis showed that dissolved CH₄ concentration was positively correlated with air temperature and TOC (p<0.01), and negatively with NO₃⁻-N and EC (p<0.01) (Table 2). CH₄ flux was positively correlated with air temperature (p<0.05), TOC and dissolved CH₄ concentration (p<0.01), and negatively with NO₃⁻-N (p<0.01) and EC (p<0.05) (Table 2 and Table S3). Environmental variables explained a larger proportion of variability in CH₄ flux derived from the *TBL* models (R²=0.46-0.54) than those from direct *FCs* measurements (R²=0.35) (Table S2).

4. Discussion

4.1. CH₄ supersaturation and degassing from aquaculture ponds

There are very few studies on CH₄ in small ponds, particularly, those created for aquaculture purposes. In this study, the dissolved CH₄ concentration in surface water of the aquaculture ponds ranged from 0.1 to 31.1 μmol L⁻¹ during the study period, which was higher than that observed in many small ponds in Florida (~2.2 μmol L⁻¹; Barber et al., 1988), Colorado (~1.0 μmol L⁻¹; Bastviken et al., 2004), Wisconsin and Minnesota (0.3–2.3 μmol L⁻¹; Smith and Lewis, 1992) in the USA, in Sweden (~1.3 μmol L⁻¹; Natchimuthu et al., 2014), Canada (0.5–6.7 μmol L⁻¹; Pelletier et al., 2014),

and Siberia (~2.6 µmol L⁻¹; Repo et al., 2007). In addition, CH₄ concentration in our aquaculture ponds was generally larger than that in some nutrient-enriched rivers in China, i.e. Lixiahe River (0.2–0.8 µmol L⁻¹; Wu et al., 2019), Beitang Drainage River and Dagu Drainage River (0.3-1.7 umol L⁻¹; Hu et al., 2018). Similar to other inland aquatic systems, such as lakes (e.g., Wen et al., 2016; Wik et al., 2016; Yan et al., 2018), reservoirs (e.g., Deemer et al., 2016; Musenze et al., 2014; Wang et al., 2017), rivers (e.g., Barbosa et al., 2016; Striegl et al., 2012), floodplains (Barbosa et al., 2020) and small ponds (e.g., Holgerson and Raymond, 2016; Wik et al., 2016), our aquaculture ponds were vastly supersaturated in CH₄ relative to air (2.71–599.81 times the equilibrium concentration) (Figure 2b). The small temperate ponds in the Yale Myers Forest in Connecticut, the USA, have some of the highest concentrations of CH₄ (21.0-58.9 µmol L⁻¹, equivalent to 119-2907 times the equilibrium concentration) (Holgerson, 2015). The CH₄ concentrations and supersaturation levels in our aquaculture ponds fall well within the range reported by Holgerson (2015), showing that aquaculture ponds in the subtropical estuaries are also hotspots for CH₄ production and emission. In inland aquatic ecosystems, the strong CH₄ release is likely a result of large organic matter inputs from the catchment, algae and aquatic plants that sustain high methanogenesis rates (Finlay et al., 2009; Lundin et al., 2013; Venkiteswaran et al., 2013; Yan et al., 2018), as indicated by the significant relationship between dissolved CH₄ and nutrient level (Huttunen et al., 2003; Kortelainen et al., 2001; Wen et al., 2016). The shrimp ponds in this study are semi-artificial ecosystems that are

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maintained through a daily feed supply for the production of aquatic animals. However, only a small portion of the feed input is converted into shrimp biomass, with the feed utilization efficiency of \sim 4.0–27.4% (Chen et al., 2016; Molnar et al., 2013; Yang et al., 2017b). Surface sediments in the aquaculture systems typically retain a large amount of organic matter from feces and residual feeds (Chen et al., 2016; Yang et al., 2017b) that can support high levels of CH₄ production and its subsequent release to atmosphere. Although organic matter content was not quantified in this study, our results confirmed the significantly correlation between dissolved CH₄ and TOC concentration (p<0.01; Table 2), which lends support to the notion that CH₄ supersaturation in the aquaculture ponds was related to the large input of organic matter.

361 4.2. Comparison of different TBL modelled CH₄ fluxes

Although previous studies have compared the performance of different *TBL* models in estimating diffusive CH₄ flux in inland waters (Amouroux et al., 2002; Li et al., 2015; Musenze et al., 2014; Xiao et al., 2017; Zappa et al., 2007), such comparison is scarce for shallow ponds, particularly aquaculture ponds. To the best of our knowledge, this study is the first attempt to compare the estimates of diffusive CH₄ flux using different *TBL* models over the whole aquaculture period in aquaculture ponds. Interestingly, although the patterns of temporal variations in diffusive CH₄ flux were largely consistent among the *TBL* models (Figure 3), there were clear differences in the magnitude of flux estimated from the different models (Table 1).

Notably, the mean flux estimated by the TBL_{RC01} model (215.6 µmol m⁻² h⁻¹) was an order of magnitude greater than that derived from the TBL_{LM86} model (19.4 µmol m⁻² h⁻¹, Figure 3). Moreover, CH₄ flux estimated by the TBL_{RC01} model was 2 - 3 times larger than that by the TBL_{W92a} , TBL_{CL98} , TBL_{W92b} and TBL_{CW03} models (Table 1 and Figure S5). However, there was no significant difference between the TBL_{W92a} and TBL_{CL98} models (p>0.05; Table 1 and Figure S5) or between the TBL_{W92b} and TBL_{CW03} models (p>0.05; Table 1 and Figure S5). In other inland waters (river and reservoirs), Gao et al. (2014) and Musenze et al. (2014) also found that the estimated diffusive CH₄ fluxes derived from the TBL_{RC01} model were substantially greater than those from other TBL models.

The differences in the estimated CH₄ flux between different *TBL* models were likely a result of different weighting of wind as a driver of gas transfer velocity (Musenze et al., 2014, Figure 1). Because these wind-based models were originally developed for specific environments under different conditions (Gao et al., 2014; Musenze et al., 2014), their suitability for other situations could be questioned (Bade, 2009; Musenze et al., 2014; Schilder et al., 2013). The *TBL*_{CL98} and *TBL*_{CW03} models were developed for lentic ecosystems under a range of wind speed, which most closely resemble aquaculture pond conditions. One may therefore argue that these two models would be most applicable to aquaculture ponds, although more *in situ* measurement will be needed to further increase the accuracy of the estimate.

4.3. Comparison of CH₄ fluxes derived from FCs measurement and TBL models

Previous studies have shown that CH₄ fluxes estimated by TBL models tend to be

lower than those measured by FCs (Chuang et al., 2017; Duchemin et al., 1999; Li et al., 2015; Matthews et al., 2003). This study also compared CH₄ fluxes measured by FCs and those estimated by TBL models over the aquaculture season (Table 1 and Figure S5). Although there were significant correlations between TBL model estimates and FCs measurements (p < 0.05 in all cases), the agreement between the two methods varied considerably between models (Figure 4). The TBLw92b and TBLcw03 models gave the largest r^2 values (0.82 and 0.83, respectively) and good agreements with FCs measurements (slope = 0.92 and 0.89, respectively), whereas TBL_{CL98} yielded mean estimates virtually identical to FCs measurements (slope = 1) but with larger variability around the mean ($r^2 = 0.53$) (Figures 4d-f). In contrast, TBL_{LM86} vastly underestimated FCs fluxes whereas TBL_{RC01} grossly overestimated FCs fluxes (Figures 4a,b). Approximately 80% of the diffusive CH₄ fluxes estimated by the models fell within the range measured by the FC method. Balancing the consideration of overall agreement (regression slope) and estimate

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Balancing the consideration of overall agreement (regression slope) and estimate variability (regression r^2), the TBL_{W92b} and TBL_{CW03} models appeared to give the best approximations of FCs measurements. While previous studies showed that FCs were more appropriate for determining greenhouse gas fluxes in heterogeneous environments such as lakes and reservoirs (Cole et al., 2010; Duchemin et al., 1999; Murray et al., 2015; Vachon et al., 2010; Wu et al., 2018), our results suggest that TBL_{W92b} and TBL_{CW03} models are reliable alternatives for estimating CH₄ diffusive flux in shallow aquaculture ponds.

In addition to diffusive flux from the water column, bottom sediment could also

contribute to CH₄ emission via ebullition, especially in eutrophic, shallow aquaculture ponds. This is illustrated by the differences in the measured CH₄ flux using FCs with and without gauze in our aquaculture ponds (Figure S6). The CH₄ flux measured by FCs without gauze (2231.3 \pm 681.3 μ mol m⁻² h⁻¹) were one to two orders of magnitude higher than that by FCs with gauze (75.0 \pm 12.5 μ mol m⁻² h⁻¹) (Figure S6); from this ebullition was estimate to contribute 96.6% to the total CH₄ emissions. Overall, our results showed that ebullition was the primary path of CH₄ emission in aquaculture ponds, and that ebullitive flux vs. diffusive flux could be easily resolved with a simple design of FCs with a detachable gauze.

4.4. Implications of the comparison between different methods

The FCs method is the popular technique for measuring CH₄ emissions due to its ability to detect low fluxes and the simplicity of its operating principle (Bastviken et al., 2015; Lorke et al., 2015; Musenze et al., 2014; Podgrajsek et al., 2014). However, the FCs method requires time-consuming manual operation, which limits the frequency of measurements and can be difficult to deploy in remote areas (Acosta et al., 2017; Morin et al., 2017). Improvement of the global CH₄ budget would require high-resolution emission data covering large time and spatial scales, which obviously is difficult to achieve with the FCs method.

Large-scale estimates of aquatic CH₄ emissions using *TBL* models has been gaining popularity (Holgerson and Raymond, 2016; Martinez-Cruz et al., 2016; Musenze et al., 2014; Wang et al., 2017) due to their simplicity, practicality and low cost. There are, however, different *TBL* models to choose from, and the large

differences in the model performances (Figure 4) mean that selecting the appropriate model(s) would be critical, or otherwise large errors would occur when upscaling the results from small ponds to the regional/ global scale. Our results suggest that TBL_{W92b} and TBL_{CW03} models could be used as effective and convenient alternatives to FCs in shallow aquaculture ponds.

4.5. Limitation and future research

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The FCs method is a common method to measure CH₄ fluxes from aquatic ecosystems. However, FCs may create microenvironments that affect the boundary layer conditions through, for instance, blockage of wind, change of atmospheric pressure at the measurement point, and change in the gas transfer rate through pressure build-up (Duchemin et al., 1999; Matthews et al., 2003; Musenze et al., 2014). For example, the turbulence resulted from the chamber walls can enhance the efficiency of gas exchange and increase gas fluxes during low wind conditions (Matthews et al., 2003; Xiao et al., 2016). TBL models rely on the gas transfer velocity coefficient (k_x), which itself is estimated from some empirical wind-based models. Effects of artificial aeration, which is commonly done in aquaculture ponds, on k_x are unknown. More importantly, the TBL models ignore the effect of buoyancy fluxes near the air-water interface on k_x . An alternative is the surface renewal model (SRM), which considers both wind speed

The use of eddy covariance (EC) technique is increasingly popular as it can

and buoyancy (e.g., Czikowsky et al., 2018; MacIntyre et al., 2010; MacIntyre et al.,

provide a better characterization of the variation in CH₄ fluxes through quasi-continuous measurements (Acosta et al., 2017; Morin et al., 2017; Xiao et al., 2014; Zhao et al., 2019). However, its application in small water bodies (e.g., ponds) is limited by footprint contamination (Zhao et al., 2019). Developing a practical and effective way to reduce the flux footprint and the contamination from gaseous sources outside the water body will allow broader application of EC method in the future.

Different methods have their own limitations; careful comparison and cross calibration would be needed to increase the overall accuracy of these methods and to improve the global CH₄ budget.

5. Conclusions

Despite the large CH₄ emission potential from small ponds, there are few studies comparing the different methods to estimate CH₄ fluxes across the water-air interface. In this study, FCs and TBL models were used to estimate CH₄ fluxes from aquaculture ponds. Our results indicate that dissolved CH₄ concentrations in the subtropical shallow aquaculture ponds were on average ~87 times oversaturated relative to the ambient air, and thus the ponds acted as strong atmospheric CH₄ sources. The high organic matter loading contributed to CH₄ supersaturation in the ponds. This study for the first time compared the CH₄ fluxes measured directly by floating chambers (FCs) and those estimated by thin boundary layer (TBL) models (TBL_{LM86} , TBL_{W92a} , TBL_{RC01} , TBL_{CL98} , TBL_{W92b} , and TBL_{CW03}). The model estimates of diffusive CH₄ fluxes were highly variable, and were overall 27 - 300% larger than those measured by FCs. The TBL_{W92b} and TBL_{CW03} models provided a robust and simple alternative to FCs in

estimating diffusive CH₄ fluxes. Our results suggest that the comparison of different methods and selection of the most appropriate method(s) should be a high research priority to improve the accuracy of greenhouse gas fluxes from aquaculture ponds and other aquatic ecosystems.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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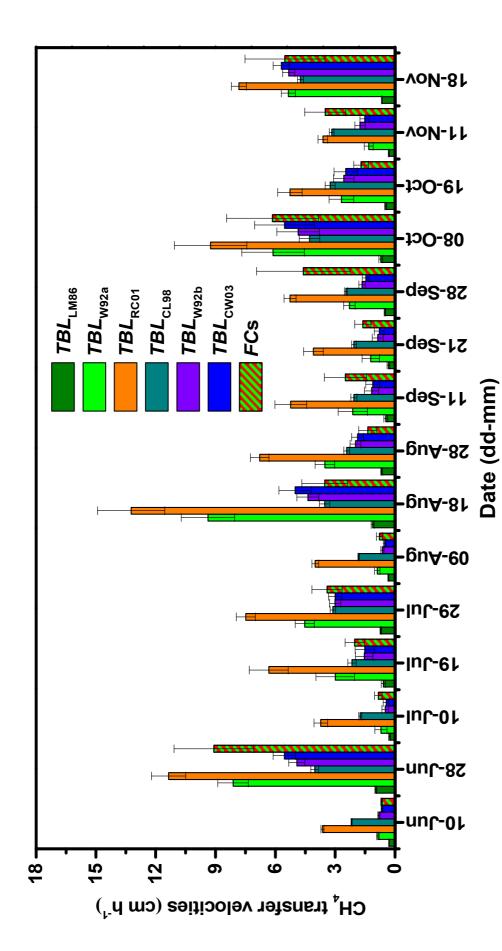


Figure 1. Temporal variation in CH4 transfer velocities from the aquaculture ponds during the aquaculture period in the Min River Estuary. Values represent the means of nine replicates samples, while the vertical lines indicate standard errors.

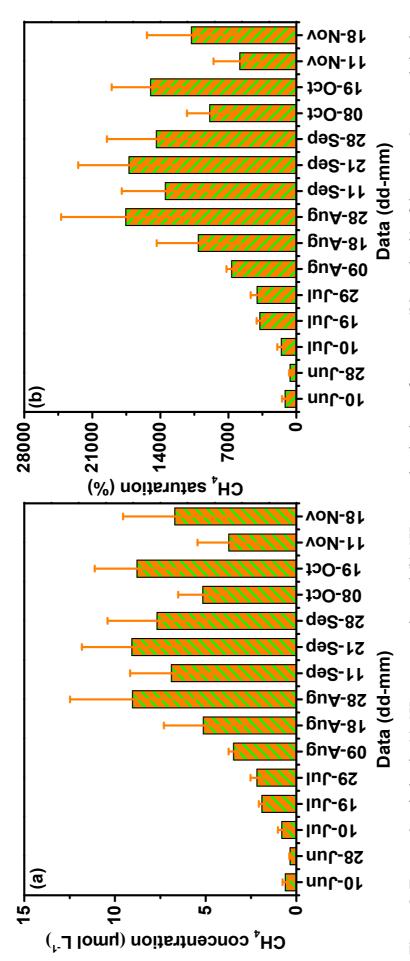
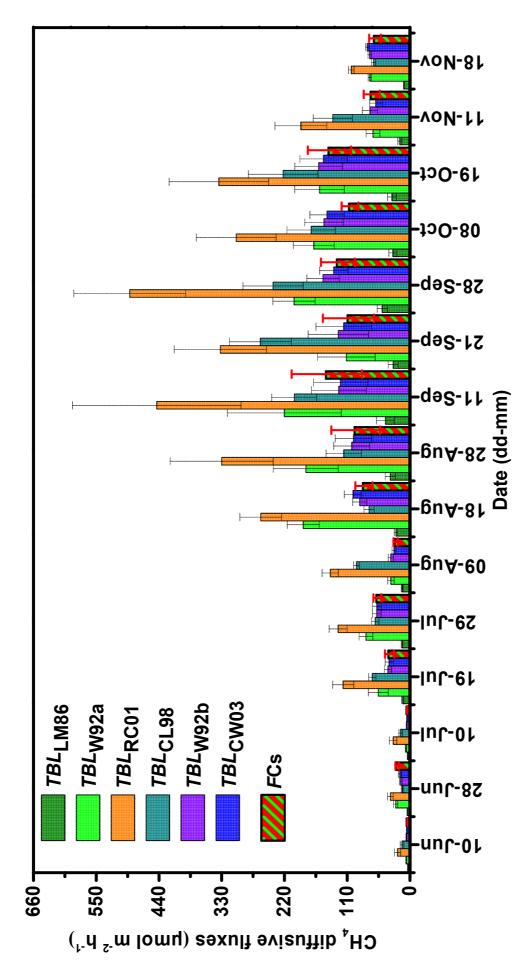


Figure 2. Temporal variation in (a) CH₄ concentration and (b) CH₄ saturation in the surface water (20 cm depth) of the aquaculture ponds in the Min River Estuary during the aquaculture period. Values represent the means of nine replicates samples, while the vertical lines indicate standard

10 /0 1

errors.



during the aquaculture period from the aquaculture ponds in the Min River Estuary. Values represent the means of nine replicates samples, while Figure 3. Temporal variation in CH₄ diffusive fluxes measured with the floating chamber method and the gas transfer velocity model methods the vertical lines indicate standard errors.

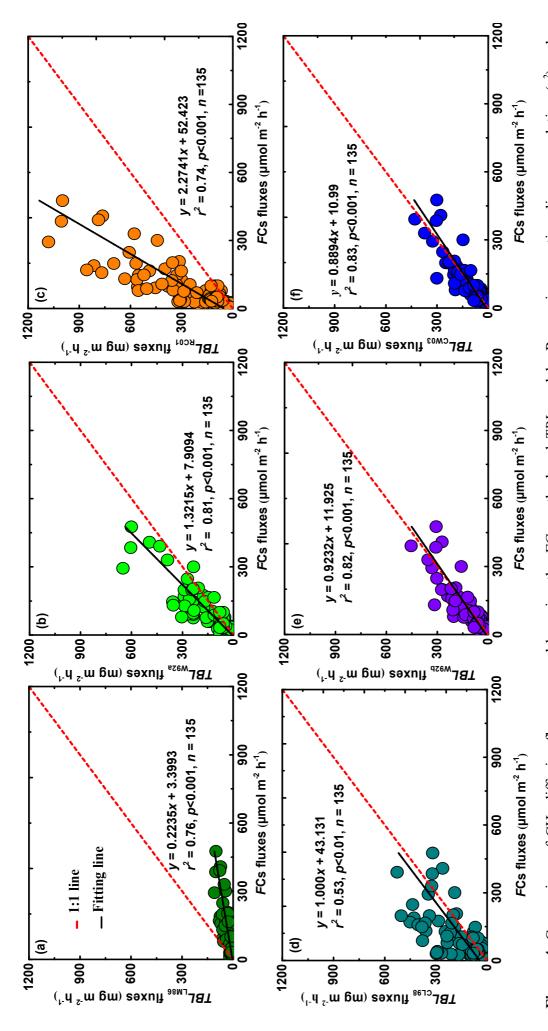


Figure 4. Comparison of CH₄ diffusive flux measured by using the FCs method and TBL models. Regression equation, linear correlation (r²) and significance (p) are also shown. Parameter bounds on the regression coefficients are 95% confidence intervals.

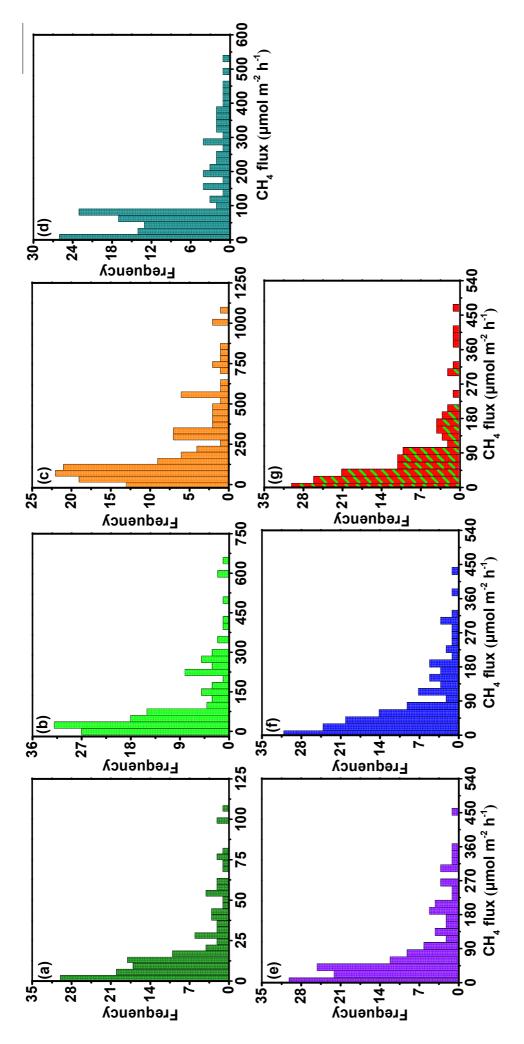


Figure 5. Frequency distribution of CH₄ diffusive fluxes from (a) TBL_{LM86}, (b) TBL_{W92a}, (c) TBL_{RC01}, (d) TBL_{CL98}, (e) TBL_{W92b}, (f) TBL_{CW03}, and (g) FCs measurements at the aquaculture ponds in the Min River Estuary during the aquaculture period. 16

Table 1

Summary of the TBL and FCs methods applied to measure CH4 diffusive fluxes from the aquaculture ponds in Min River Estuary during the

aquaculture period.

	TBL methods						170 m 200
	$TBL_{ m LM86}$	$TBL_{ m W92a}$	$TBL_{ m RC01}$	TBL_{CL98}	$TBL_{ m W92b}$	$TBL_{ m CW03}$	rcs method
Minimum (μ mol m ⁻² h ⁻¹)	9.0	1.3	5.6	1.9	1.9	1.3	1.3
Maximum (μ mol m ⁻² h ⁻¹)	108.8	650.0	1079.4	527.5	454.4	428.8	476.3
Average (μ mol m ⁻² h ⁻¹)	19.4	103.1	215.6	115.0	78.1	75.0	71.9
Standard deviation	23.1	130.6	236.3	122.5	91.3	6.98	88.8
Coefficient of variation	1.18	1.27	1.09	1.06	1.16	1.15	1.24

Table 2

- Pearson correlation coefficients for dissolved CH₄ concentration, CH₄ diffusive fluxes 9
- and environmental variables from the aquaculture ponds in Min River Estuary during 9
- the aquaculture period^a. Bold numbers denote correlation coefficients for significant _
- relationships. ∞

Environmental variables	Environmental variables Dissolved CH ₄ concentration CH ₄ diffusive fluxes	CH4 diffusive fluxes
Meteorological parameters		
Air temperature	0.214*	0.203*
Wind speed (W_S)	NS	0.281*
Atmospheric pressure	NS	NS
Water parameters		
Water temperature	NS	NS
Dissolved oxygen (DO)	NS	NS
TOC concentration	0.312**	0.296**
NO ₃ N concentration	-0.401**	-0.392**
Electrical conductivity (EC) -0.361**	-0.361**	-0.185*

^a The symbols * and ** indicate significant correlations at the 0.05 and 0.01 levels, respectively. n = 135 for

environmental variables and CH4 diffusive fluxes from the aquaculture ponds. CH4 diffusive fluxes were directly 9 10 111

measured using floating chambers method.

Supporting Information

- 2 Title: Diffusive CH4 fluxes from aquaculture ponds using floating
- 3 chambers and thin boundary layer equations
- 4 Ping Yang^{a,b,1}, Jiafang Huang^{a,b,1}, Hong Yang^{c,d,e,1}, Josep Peñuelas^{f,g}, Kam W. Tang^h,
- 5 Derrick Y.F. Lai^{i*}, Dongqi Wang^j, Qitao Xiao^k, Jordi Sardans^{f,g,**}, Yifei Zhang^{a,b},
- 6 Chuan Tonga,b,***
- 7 *Key Laboratory of Humid Subtropical Eco-geographical Process of Ministry of Education, Fujian
- 8 Normal University, Fuzhou 350007, P.R. China
- 9 bSchool of Geographical Sciences, Fujian Normal University, Fuzhou 350007, P.R. China
- 10 *College of Environmental Science and Engineering, Fujian Normal University, Fuzhou 350007, P.R.
- 11 China
- dCollaborative Innovation Center of Atmospheric Environment and Equipment Technology, Jiangsu
- 13 Key Laboratory of Atmospheric Environment Monitoring and Pollution Control (AEMPC), School of
- 14 Environmental Science and Engineering, Nanjing University of Information Science and Technology,
- 15 Nanjing 210044, China;
- ^oDepartment of Geography and Environmental Science, University of Reading, Reading RG6 6AB,
- 17 *U.K.*
- 18 CSIC, Global Ecology Unit CREAF-CSIC-UAB, Bellaterra, Catalonia, Spain
- 19 CREAF, Cderdanyola del Vallès, Catalonia, Spain
- 20 hDepartment of Biosciences, Swansea University, Swansea SA2 8PP, U. K.
- ¹Department of Geography and Resource Management, The Chinese University of Hong Kong, Shatin,
- 22 New Territories, Hong Kong SAR, China
- ¹School of Geographical Sciences, East China Normal University, Shanghai 200241, China
- 24 Key Laboratory of Watershed Geographic Sciences, Nanjing Institute of Geography and Limnology,
- 25 Chinese Academy of Sciences, Nanjing, 210008, China
- ***Correspondence to:** Derrick Y.F. Lai
- 27 **Email:** dyflai@cuhk.edu.hk
- ***Correspondence to:** Jordi Sardans

- 29 **Email:** j.sardans@creaf.uab.cat
- 30 **Correspondence to: Chuan Tong
- 31 **Email:** tongch@fjnu.edu.cn
- ¹Ping Yang, Jiafang Huang, and Hong Yang contributed equally to this work.

33 Supporting Information Summary

- No. of pages: 16 No. of figures: 4 No. of tables: 5
- 35 Page S5: Figure S1. Location of the study area and sampling sites at aquaculture
- 36 ponds in Min River Estuary, Southeast China.
- Page S6: Figure S2. Schematic diagram for the gas sampling device of CH₄ diffusive
- 38 flux across the water-air interface. Numbers 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, and 12
- 39 represents chambers body, Neoprene floats, thin gauze, mooring anchor, sampling
- 40 tube, 60-mL plastic syringes equipped with three-way stopcocks, fixed rope, valve
- body, valve body, gas collecting hole, ribbon, and handle, respectively.
- 42 **Page S7:** Figure S3. (a) Temporal variation in the wind speed (W_S) , and (b) frequency
- distribution of wind speed at the shrimp ponds in the Min River Estuary during the
- 44 aquaculture period.
- 45 Page S8: Figure S4. Temporal variation in (a) water temperature, (b) dissolved
- oxygen, (c) TOC, and (b) N-NO_x in the surface water (20 cm depth) of the
- 47 aquaculture ponds in the Min River Estuary during the aquaculture period. *Error bars*
- 48 represent standard error (n = 9). Data are after Yang et al. [unpublished data] for
- 49 reference and review only.
- Page S9: Figure S5. Boxplots of CH₄ diffusive fluxes estimated using the TBL and
- 51 FCs methods at the aquaculture ponds in Min River Estuary during the aquaculture
- 52 period. The letters above the boxes represent the LSD (Least Significant Difference)
- test results, and different letters mean significant difference at 0.05 level. The centre
- line and square represent the median value and area-weighted average.
- Page S10: Figure S6. Comparison of CH₄ fluxes measured using the FCs with gauze
- 56 (FCs-G) and without gauze (FCs-NG) from the aquaculture ponds in the Min River
- 57 Estuary during the aquaculture period. Data are after Yang et al. [unpublished data]
- 58 for reference and review only. Values represent the means of nine replicates samples,
- 59 while the vertical lines indicate standard errors.
- 60 Page S11: Table S1. Characteristics of the three aquaculture ponds in the Min River
- 61 Estuary.

- Page S12: Table S2. The best GLS model (lowest AIC values) with the CH₄ fluxes as
- 63 functions of environmental values.
- Page S15: Table S3. Pearson correlation coefficients for environmental variables,
- dissolved CH₄ concentration and CH₄ diffusive fluxes from the aquaculture ponds in
- 66 Min River Estuary during the aquaculture period.

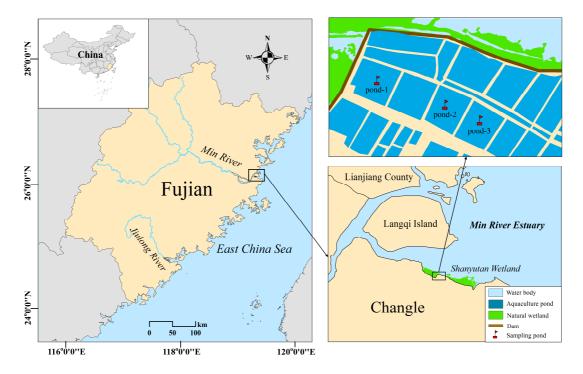


Figure S1. Location of the study area and sampling sites at aquaculture ponds in Min
 River Estuary, Southeast China.

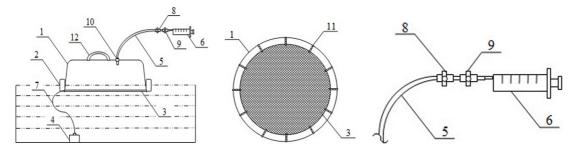


Figure S2. Schematic diagram for the gas sampling device of CH₄ diffusive flux across the water-air interface. Numbers 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, and 12 represents chambers body, Neoprene floats, thin gauze, mooring anchor, sampling tube, 60-mL plastic syringes equipped with three-way stopcocks, fixed rope, valve body, valve body, gas collecting hole, ribbon, and handle, respectively.



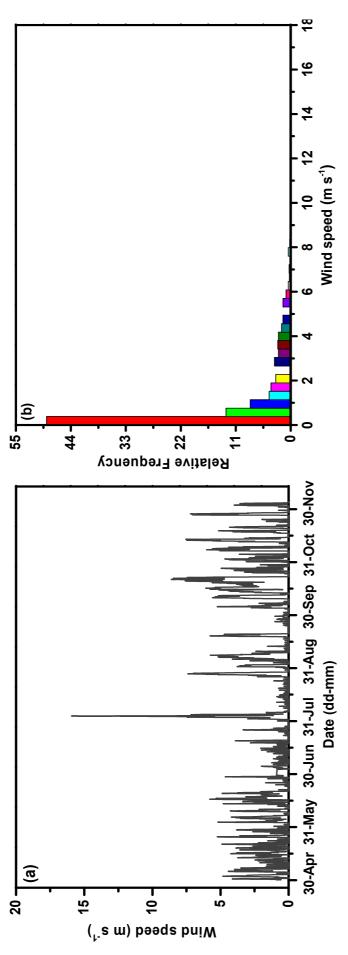


Figure S3. (a) Temporal variation in the wind speed (Ws), and (b) frequency distribution of wind speed at the shrimp ponds in the Min River

Estuary during the aquaculture period.

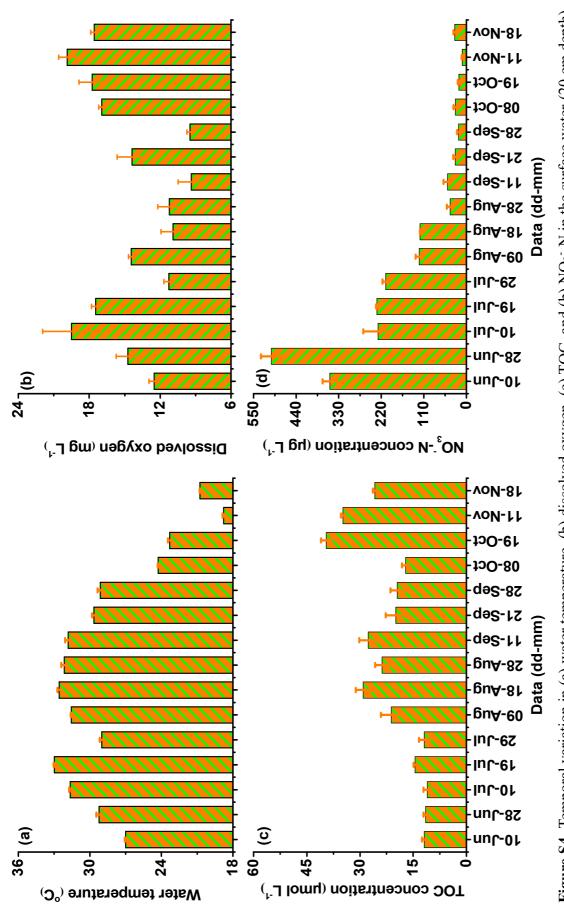


Figure S4. Temporal variation in (a) water temperature, (b) dissolved oxygen, (c) TOC, and (b) NO₃-N in the surface water (20 cm depth) of the aquaculture ponds in the Min River Estuary during the aquaculture period. Error bars represent standard error (n = 9). Data are after Yang et al. [unpublished data] for reference and review only

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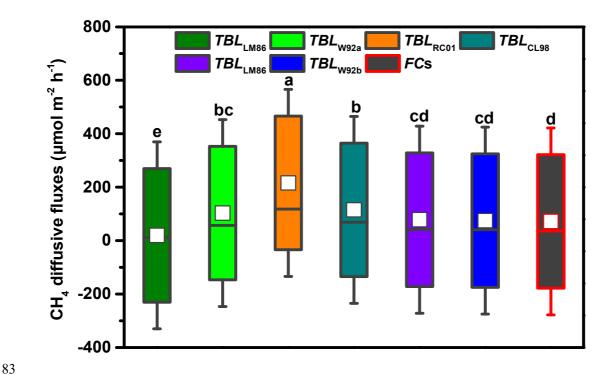


Figure S5. Boxplots of CH₄ diffusive fluxes estimated using the *TBL* and *FCs* methods at the aquaculture ponds in Min River Estuary during the aquaculture period. The letters above the boxes represent the LSD (Least Significant Difference) test results, and different letters mean significant difference at 0.05 level. The centre line and square represent the median value and area-weighted average.

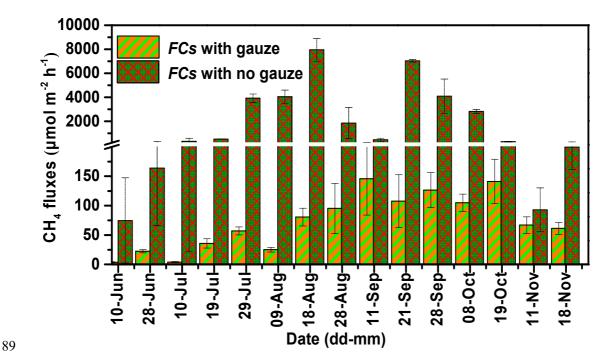


Figure S6. Comparison of CH₄ fluxes measured using the *FCs* with gauze (*FCs*-G) and without gauze (*FCs*-NG) from the aquaculture ponds in the Min River Estuary during the aquaculture period. Data are after Yang et al. [*unpublished data*] for reference and review only. Values represent the means of nine replicates samples, while the vertical lines indicate standard errors.

75 **Table S1** Characteristics of the three aquaculture ponds in the Min River Estuary. *.

	1	1	<u> </u>
Parameters	Pond-I	Pond-II	Pond-III
Shrimp species	Litopenaeus vannamei	Litopenaeus vannamei	Litopenaeus vannamei
Water depth (m)	1.3(0.3-1.7)	1.7(0.5-2.1)	1.5(0.3-1.8)
Water salinity (‰)	3.6(1.5-6.8)	2.2(1.7-5.2)	2.8(1.7-5.4)
Surface area (m ²)	21426.94	18412.89	19112.71
Stocking density (PL m ⁻²) ^a	150	120	119
Survival rate (%) ^a	45	62	59
Feed conversion rateb	3.5	2.3	2.5

^{96 *}Based on Zhang et al. (2019).

a The data for the stocking density, survival rate, and yield were provided by the farmers; b Feed conversion rate =
 dry weight of feeds added / wet weight of shrimps produced.

Table S2 The best GLS model (lowest AIC values) with the CH4 fluxes as functions of environmental values.

Variables							
	Model	Model R^2 and	Model independent factors statistics	endent fact	ors statistics		
		<i>P</i> -value					
$TBL_{ m LM86}$	$gls(CH4flux \sim disCH4 + U10 + watertemp + NO3N,$	$R^2 = 0.47$	Coefficients:				
	data=dades, method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	-0.396	0.166	-2.38	0.019
			disCH4	0.0300	0.00399	7.54	<0.0001
			U10	0.0515	0.0167	3.08	0.0025
			watertemp	0.0187	0.00553	3.39	0.00093
			NO3N	-0.786	0.178	-4.41	<0.0001
$TBL_{ m W92a}$	gls(CH4flux ~ disCH4 + U10 + watertemp + NO3N,	$R^2 = 0.50$	Coefficients:				
	data=dades, method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	-3.52	0.945	-3.73	<0.0001
			disCH4	0.141	0.0227	6.20	<0.0001
			U10	0.614	0.0951	6.45	<0.0001
			watertemp	0.122	0.0314	3.89	<0.0001
			NO3N	4.64	1.014	-4.58	<0.0001
$TBL_{ m RC01}$	gls(CH4flux \sim disCH4 + U10 + watertemp + NO3N,	$R^2 = 0.54$	Coefficients:				
	data=dades, method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	-3.32	1.64	-2.03	0.045
			disCH4	0.331	0.0393	8.42	<0.0001
			U10	0.464	0.165	2.81	0.0057
			watertemp	0.178	0.0546	3.26	0.0014
		Č	NO3N	-8.18	1.76	-4.65	<0.0001
$TBL_{ m CL98}$	gls(CH4flux ~ disCH4 + NO3N, data=dades,	$R^2=0.53^{12}$	Coefficients:				

	method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	1.45	0.212	98.9	<0.0001
			disCH4	0.181	0.0203	8.95	<0.0001
			NO3N	-3.59	0.862	-4.16	<0.0001
TBL_{W92b} g	gls(CH4flux ~ disCH4 + U10 + NO3N, data=dades,	$R^2 = 0.46$	Coefficients:				
ш	method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	0.348	0.244	1.43	0.16
			disCH4	0.108	0.0163	6.61	<0.0001
			U10	0.287	0.0676	4.24	<0.0001
			NO3N	-2.90	0.695	4.18	0.0001
TBL _{CW03} gi	gls(CH4flux ~ disCH4 + U10 + NO3N, data=dades,	$R^2 = 0.48$	Coefficients:				
ш	method ="REML")	P < 0.0001		Estimate	Std. Error	t-value	p-value
			(Intercept)	0.178	0.230	0.774	0.44
			disCH4	0.0988	0.0153	6.46	<0.00019
			U10	0.340	0.0634	5.35	< 0.0001
			NO3N	-2.79	0.652	-4.27	<0.0001
FCs	gls(CH4flux \sim disCH4 + U10 + N03N, data=dades,	$R^2 = 0.35$		Estimate	Std. Error	t-value	p-value
ш	method ="REML")	P < 0.0001	(Intercept)	0.258	0.263	86.0	0.33
			disCH4	0.0784	0.0175	4.47	<0.0001
			U10	0.324	0.0728	4.46	<0.0001
			NO3N	-2.71	0.748	-3.63	0.00041

Table S3 Pearson correlation coefficients for environmental variables, dissolved CH4 concentration and CH4 diffusive fluxes from the aquaculture ponds in Min River Estuary during the aquaculture period.

	TBLw92b TBLCw03	
	TBL_{CL98}	
	$TBL_{ m RC01}$	
luxes	$TBL_{ m W92a}$	
CH4 diffusive fi	$TBL_{ m LM86}$	
To the state of th	Environmental variables	

The symbols * and ** indicate significant correlations at the 0.05 and 0.01 levels, respectively. NS indicates non-significant.

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