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Liquid-Liquid Equilibrium Data for the Ternary Systems of Palm Oil + Ethyl Lactate + Phytonutrients (Carotenes and Tocols) at 303.15 K

Y. L. Kua, S. Gan, A. Morris, and H. K. Ng

Abstract—Experimental and thermodynamic modeling studies of liquid-liquid equilibria for two model systems of β -carotene and γ -tocotrienol in palm oil + ethyl lactate were performed at 303.15 K and atmospheric pressure in an effort to explore the potential application of ethyl lactate as a green and safe agrochemical solvent to recover carotenes and tocals from palm oil. The experimental data were correlated by Universal QuasiChemical (UNIQUAC) model while the thermodynamic data enables simulation of liquid-liquid extractors for phytonutrients recovery from palm oil as well as the prediction of energy requirement, equipment sizing and phase compositions for the oil- and solvent-rich phases. The results showed that ethyl lactate favors the extraction of phytonutrients from palm oil especially tocals due to their higher polarity and separation factors as compared to carotenes.

Index Terms—Carotene, ethyl lactate, palm oil, tocol.

I. INTRODUCTION

Carotenes are important precursors of vitamin A and are also natural antioxidants in combating several degenerative diseases including cardiovascular diseases, cancers, immunity and macular degeneration. Crude palm oil contains 500-700 ppm of carotenes with approximately 56% of β -carotene [1]. The vitamin A activity of β -carotene, in terms of retinol equivalent, is 2 times higher than other provitamin A carotenes [2]. Therefore, palm oil has 15 times more retinol equivalent than carrots and 300 times more than tomatoes [3].

Tocals, including tocopherols and tocotrienols, are commonly known as vitamin E. They protect the body against oxidative damage, skin damage and aging by UV-radiation. Additionally, they help to boost the immune system, fight against infections and prevent the occurrence of old age cataract. As compared to tocopherol, tocotrienol has been reported to be more effective due to its unsaturated chain, which facilitates cell penetration and is highly antioxidative [4]. Tocotrienol was also reported to regulate cholesterol level, and prevent cancers, stroke and fats accumulation in liver [5]-[8]. Tocopherol is commonly found in many vegetable oils including cottonseed, groundnut and olive oil while palm and rice bran oil are among the richest natural sources of tocotrienol. Palm oil has 600-1000 ppm of tocals, with 78.7% of tocotrienol [9].

Palm oil is obtained from oil palm fruits (*Elaeis guineensis/tenera*) through screw pressing at high pressure.

After oil refining through either physical or chemical processes, palm oil is the vegetable oil most consumed in the world, being 35.05% in 2015/16 [10]. Oil palm is the highest oil-producing plant with a yield of approximately 4 ton of oil per hectare per year. It is mainly planted, produced and exported from Indonesia and Malaysia. As compared to chemical refining process, physical refining is commonly used for palm oil due to better yield at lower cost. During physical oil refining process, carotenes are first partially removed by adsorption on activated bleaching earth, followed by high temperature steam deodorization which destroys the chromogenic properties of the remaining carotenes to produce a light yellow palm oil. Even though tocals are more thermally stable than carotenes, near to 50% of the tocals will be stripped off along with free fatty acids (FFA), sterols and squalene into palm fatty acid distillate (PFAD) during deodorization step. Thus, there is a need to recover these phytonutrients from the oil before further refining process.

As both the extracted carotenes and tocals will be used as food fortifiers, the use of non-toxic, non-corrosive and non-carcinogenic solvents which are safe for human consumption is crucial. While many of the commonly used petrochemical solvents are known to pose certain degree of toxicity, ethyl lactate is a suitable candidate solvent because it is novel, green and safe. It is produced from carbohydrate feedstocks from the corn and soybean industries and it is present naturally in foods such as wine, beer, chicken and fruits. It is also non-ozone depleting, non-hazardous air pollutant and it is biodegradable into harmless compounds such as carbon dioxide (CO₂) and water. The US Environmental Protection Agency (USEPA) approved the solvent as a Significant New Alternatives Policy Program (SNAP) solvent while US Food and Drug Administration (USFDA) has approved its direct use in food and pharmaceutical products. It is generally recognized as a safe (GRAS) solvent. Ethyl lactate exerts polarity in the range of acetonitrile and n-hexane. It is capable of forming intra- and inter-molecular hydrogen bonding [11]. Also, it has the ability to form Van der Waals interactions in oils [12]. As a result, ethyl lactate can dissolve in both aqueous and hydrocarbon environments and is capable of extracting compounds of a wide range of polarity [13].

Ethyl lactate has been reported to extract various phytonutrients mainly from solid matrix [13], [14]-[19]. From oil samples, [20] and [21] reported the liquid-liquid equilibrium data on the recovery of squalene and tocopherol from olive oil. Literature on the liquid-liquid equilibrium data of oil-deacidification systems by alcoholic solvents have been extensively published. Some of the papers also

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considered the partition of nutraceutical compounds by assuming that they are present at infinite dilution [22]-[24]. The distribution coefficients were reported instead of the liquid-liquid equilibrium data and these compounds are preferably retained in the oil rather than being extracted.

In this paper, ternary systems of palm oil + ethyl lactate + β -carotene and γ -tocotrienol were studied for the first time in order to examine the potential of ethyl lactate as a novel, green and safe solvent to extract the phytonutrients from palm oil in liquid-liquid extractors. Cloud points and upper critical solution temperature (UCST) of the binary system of palm oil + ethyl lactate are reported to examine the mutual miscibility of the system. Subsequently, the experimental data is used to adjust the binary interaction parameters of the UNIQUAC model in order to predict the liquid-liquid phase equilibria behavior of the pseudoternary systems.

II. MATERIAL AND METHODS

A. Chemicals

(S)-(-)-ethyl lactate (99% purity) and n-hexane (HPLC grade) were obtained from Merck. β -carotene (99.4% purity) originated from Calbiochem while γ -tocotrienol (97% purity) were bought from Davos Life Science (Singapore). Acetonitrile (HPLC grade) and dichloromethane (99.8% purity) were obtained from Labscan. 2-propanol (HPLC grade) came from Fisher while palm oil (Brand Saji) was purchased from local grocery store in Malaysia.

B. Cloud Point Determination

Cloud point (or transition temperature) is the temperature at which the solution turns turbid as an indication of single- to two-phase transition for binary or higher level system. A total of 14.5 g of solution was prepared in a glass sample bottle by weighting the corresponding amounts of palm oil and ethyl lactate on a precise analytical balance with 0.0001 g accuracy. The solution was agitated in a temperature-controlled water bath with a calibrated digital thermometer inserted. The solution was first heated up until it became clear (single phase). Next, the water bath was cooled gradually until the cloud point was observed. Three measurements were taken with difference of no more than 0.3°C.

C. Liquid-Liquid Equilibria Measurements

A total of 14.5 g of solution was prepared in a glass sample bottle by weighting the corresponding amounts of palm oil + ethyl lactate + β -carotene and γ -tocotrienol on a precise analytical balance with 0.0001 g accuracy. The solution was agitated in a temperature-controlled water bath with a calibrated digital thermometer inserted to monitor the temperature at 303.15 \pm 0.5 K for 3 h. After that, the solution was left to settle in an oven at 303.15 K for 24 h. Temperature as low as 303.15 K was chosen in order to preserve the heat- and light-sensitive phytonutrients. The top (oil-rich) and bottom (solvent-rich) phase were then carefully withdrawn by gas-tight needle syringe (SGE Analytical Science) for high performance liquid chromatography (HPLC) analysis of β -carotene and γ -tocotrienol. Ethyl lactate content in both phases was determined gravimetrically after solvent evaporation. At least three experimental data were obtained to adjust the parameters of UNIQUAC model.

D. HPLC Analysis

An Agilent 1260 Infinity Series HPLC system was used along with a quaternary pump, an autosampler, a variable wavelength detector (VWD) and ChemStation software for system control and data collection. β -carotene was eluted from Purospher STAR RP-18 encapped column (5 μ m; 4.6 \times 250 mm). The mobile phase consisted of 85% acetonitrile and 15% dichloromethane at 1.5 mL/min. The sample was diluted by ethyl lactate and 20 μ L of analyte was injected per analysis. The detector was set at 450 nm and the total run time was 25 min. γ -tocotrienol was eluted from Zorbax Rx-SIL column (5 μ m; 4.6 \times 250 mm). The mobile phase consisted of 99% of n-hexane and 1% of 2-propanol at 0.8 mL/min. The sample was re-dissolved in n-hexane and 20 μ L of analyte was injected per analysis. The detector was set at 292 nm and the total run time was 15 min. All the mobile phase and analyte were filtered through 0.45 μ m nylon (polar) or polytetrafluoroethylene (PTFE) (non-polar) membrane before use.

E. Thermodynamic Modeling

UNIQUAC model is an activity coefficient equation to predict the highly non-ideal systems [25]. It is suitable for complicated system which involves polar and non-polar species in phase splitting [26]. The model is based on local composition theory that considers the effect of mixture composition towards mixture interaction. It consists of two contributions – combinatorial term to indicate the geometric significance for molecules of different sizes and shapes, and residual term to include the energetic parameters [27].

In the UNIQUAC model, the activity coefficient (γ_i) was predicted by the following equations.

$$\ln \gamma_i = \ln \gamma_i^C + \ln \gamma_i^R \quad (1)$$

$$\ln \gamma_i^C = \ln \frac{\psi_i}{x_i} + \left(\frac{Z}{2}\right) q_i \ln \frac{\theta_i}{\psi_i} + l_i - \left(\frac{\psi_i}{x_i}\right) \sum_{j=1}^C x_j l_j \quad (2)$$

$$\ln \gamma_i^R = q_i \left[1 - \ln \left(\sum_{j=1}^C \theta_j \tau_{ij} \right) - \sum_{j=1}^C \left(\frac{\theta_j \tau_{ij}}{\sum_{k=1}^C \theta_k \tau_{kj}} \right) \right] \quad (3)$$

where

$$l_i = \left(\frac{Z}{2}\right) (r_i - q_i) - (r_i - 1) \quad (4)$$

$$\psi_i = \frac{x_i r_i}{\sum_{i=1}^C x_i r_i} \quad (5)$$

$$\theta_i = \frac{x_i q_i}{\sum_{i=1}^C x_i q_i} \quad (6)$$

$$r_i = \sum_g v_{g(i)} S_g \quad (7)$$

$$q_i = \sum_g v_{g(i)} Q_g \quad (8)$$

$$\bar{Z} = 10 \quad (9)$$

$$\tau_{ij} = \exp\left(\frac{\mu_{ij} - \mu_{jj}}{RT}\right) \quad (10)$$

r_i and q_i are the volume and surface area parameters of compound i . They were computed from bond angle and bond distance and were estimated by group-contribution method using tabulated S_g and Q_g values from [28] and [29]. $v_{g(i)}$ is the number of functional group g for compound i , x_i is the mole fraction of compound i . For each binary pair, there were two binary interaction coefficients (τ_{ij}) that were adjusted by experimental data. They were determined using Microsoft solver through minimization of the objective function (OF).

$$K_i = \frac{x_i^{[1]}}{x_i^{[2]}} = \frac{\gamma_i^{[2]}}{\gamma_i^{[1]}} \quad (11)$$

$$OF = \sum_{\text{all points}} (K_x - K_y)^2 \quad (12)$$

where K_x is the distribution coefficient as obtained by experimental data while K_y is calculated from activity coefficient, which is obtained from an initial guess of three pairs of binary interaction coefficients (τ_{ij}). Superscripts [1] and [2] are solvent-rich and oil-rich phases, respectively.

For the prediction of the behavior of liquid-liquid equilibrium, the fitted and calculated UNIQUAC parameters were utilized to construct ternary diagrams for the pseudoternary systems of palm oil + ethyl lactate + β -carotene or γ -tocotrienol at 303.15 K.

III. RESULTS AND DISCUSSION

Fig. 1 shows the cloud point for the binary system of palm oil + ethyl lactate. The system exhibits UCST at approximately 0.65 mass fraction of ethyl lactate, with the critical point at 320 K. [21] reported a binary system of olive oil + ethyl lactate as the UCST was found at 311.2 K. This suggests that ethyl lactate exerts better miscibility with olive oil than with palm oil. The cloud point data is useful to determine the biphasic condition in liquid-liquid extractors. Within the bell curve, the binary system would settle into two

phases while homogeneous single phase exists outside the curve.

Table I presents the overall composition and the corresponding tie lines for the pseudoternary systems composed of palm oil + ethyl lactate + β -carotene and γ -tocotrienol.

In the thermodynamic modeling, palm oil was represented by a single component, which is composed of glycerol backbone with an equal number of palmitic and oleic acid side chains. This simplification was based on the fact that the two major fatty acids in palm oil are palmitic (43.7%) and oleic acids (39.9%). As β -carotene and γ -tocotrienol are the major carotene and tocol in palm oil, they were used to model carotenes and tocols in palm oil, respectively. This approach assumes that the different triglycerides, carotenes and tocols behave similarly in the liquid-liquid system under analysis.

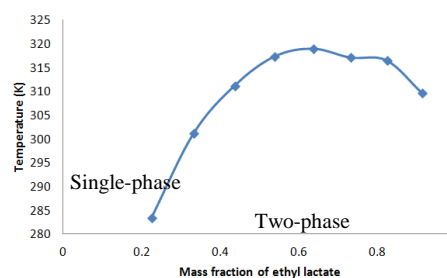


Fig. 1. Cloud point with respect to mass fraction of ethyl lactate for the binary mixture of palm oil + ethyl lactate at atmospheric pressure.

TABLE I: LIQUID-LIQUID EQUILIBRIUM DATA FOR THE TERNARY SYSTEMS OF PALM OIL (1) + ETHYL LACTATE (2) + BETA-CAROTENE (3) AND GAMMA-TOCOTRIENOL (4) AT 303.15 K

Overall Composition			Solvent-rich phase			Oil-rich phase		
X_1	X_2	X_3	X_1	X_2	X_3	X_1	X_2	X_3
0.1065	0.8934	1.5676×10^{-4}	0.0142	0.9858	2.1067×10^{-5}	0.1878	0.8120	2.7450×10^{-4}
0.1065	0.8934	5.2398×10^{-5}	0.0139	0.9861	9.7709×10^{-6}	0.1878	0.8121	8.9773×10^{-5}
0.0572	0.9427	1.3283×10^{-4}	0.0125	0.9874	2.2093×10^{-5}	0.1941	0.8055	4.7439×10^{-4}
0.0571	0.9429	9.4545×10^{-5}	0.0127	0.9873	2.0449×10^{-5}	0.1930	0.8067	3.2160×10^{-4}
0.0796	0.9203	1.0695×10^{-4}	0.0133	0.9866	1.6490×10^{-5}	0.1900	0.8097	2.5762×10^{-4}
0.1103	0.8895	1.9644×10^{-4}	0.0147	0.9853	2.1803×10^{-5}	0.1876	0.8121	3.3885×10^{-4}
0.0794	0.9205	1.4754×10^{-4}	0.0132	0.9868	1.8161×10^{-5}	0.1895	0.8102	3.6419×10^{-4}
X_1	X_2	X_4	X_1	X_2	X_4	X_1	X_2	X_4
0.0434	0.9563	3.1460×10^{-4}	0.0109	0.9889	2.2670×10^{-4}	0.2210	0.7782	7.9415×10^{-4}
0.0611	0.9387	1.4143×10^{-4}	0.0126	0.9874	8.9130×10^{-5}	0.2205	0.7792	3.1385×10^{-4}
0.0423	0.9574	2.9283×10^{-4}	0.0114	0.9884	2.1247×10^{-4}	0.2266	0.7726	7.7083×10^{-4}
0.0569	0.9428	3.2376×10^{-4}	0.0107	0.9891	1.9704×10^{-4}	0.2209	0.7783	7.7399×10^{-4}
0.0995	0.9001	3.4604×10^{-4}	0.0115	0.9883	1.6463×10^{-4}	0.2255	0.7739	6.0434×10^{-4}

TABLE II: THE VOLUME (R_g), SURFACE (Q_g) GROUP PARAMETERS [28], [29] AND THE NUMBER OF FUNCTIONAL GROUPS IN PALM OIL, ETHYL LACTATE, BETA-CAROTENE AND GAMMA-TOCOTRIENOL

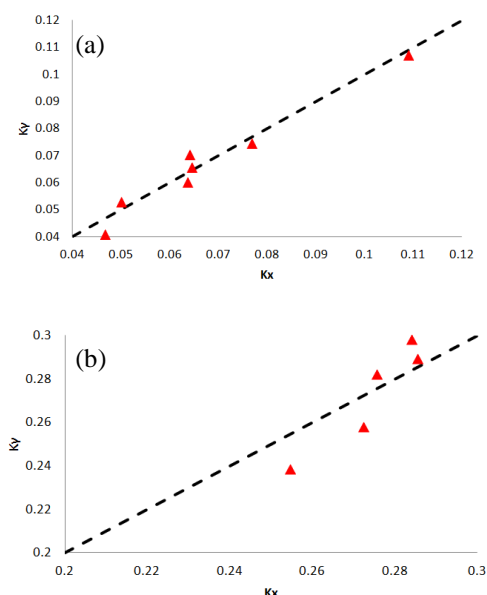
Functional group (g)	R_g	Q_g	Palm oil	Ethyl lactate	β -carotene	γ -tocotrienol
CH ₃	0.9011	0.848	3	2	8	4
CH ₂	0.6744	0.540	42	-	5	8
CH	0.4469	0.228	-	1	1	-
C	0.2195	0	-	-	2	-
CH = CH	1.1167	0.867	1.5	-	5	-
CH = C	0.8886	0.676	-	-	4	3
ACH	0.5313	0.400	-	-	1	1
AC	0.3652	0.120	-	-	1	2
ACCH ₃	1.2663	0.968	-	-	2	2
OH	1.0000	1.200	-	1	-	-
ACOH	0.8952	0.6800	-	-	-	1
CH ₃ CO	1.6724	1.4880	-	-	-	1
CH ₂ COO	1.6764	1.420	3	1	-	-

TABLE III: MOLECULAR WEIGHT, VOLUME AND SURFACE PARAMETERS FOR PALM OIL, ETHYL LACTATE, B-CAROTENE AND γ -TOCOTRIENOL

Component i	Molecular weight (g/mol)	r_i	q_i
Palm oil	846	37.7324	30.785
Ethyl lactate	118	4.9255	4.544
β -carotene	536	24.0337	19.207
γ -tocotrienol	410	18.0273	14.484

TABLE IV: BINARY INTERACTION PARAMETERS FOR SYSTEMS COMPRISING PALM OIL, ETHYL LACTATE, BETA-CAROTENE AND GAMMA-TOCOTRIENOL. VALUES AS OBTAINED BY GAMMA-TOCOTRIENOL MODEL ARE REPRESENTED IN BRACKETS.

Component i	Component j	τ_{ij}	τ_{ji}
Palm oil	Ethyl lactate	0.7705	1.0039
		(0.7242)	(1.0405)
Palm oil	β -carotene	0.03704	0.0029
		(0.0435)	(2.8781)
Ethyl lactate	β -carotene	0.01224	1.0193
Ethyl lactate	γ -tocotrienol	(0.0099)	(4.6452)


 Fig. 2. The comparison of K_x and K_y for β -carotene (a) and γ -tocotrienol (b).

The volume (R_g), surface (Q_g) group parameters and the number of functional groups in palm oil, ethyl lactate, β -carotene and γ -tocotrienol are tabulated in Table II. The volume and surface area parameters as calculated through functional-group contribution method for each of the component (palm oil, ethyl lactate, β -carotene and γ -tocotrienol) are tabulated in Table III.

The most reliable procedure for the determination of UNIQUAC model parameters involves a fit to available experimental data over a wide range of liquid compositions. For the binary system of palm oil + ethyl lactate, the interaction parameters (τ_{ij}) were determined as the first guess for the ternary systems. It is not uncommon for the OF in such nonlinear parameter estimation problems to be non-convex and thus have several local minima. Because the methods typically used to solve such problems, like the one used in the present study, are local methods that are strongly dependent on the initial estimates applied, they provide no guarantee that the global optimum has been found. Thus, the sets of the binary interaction parameters obtained likely

represent just one of the possible sets of parameters [20]. The binary interaction parameters as adjusted by the experimental data through model fitting are tabulated in Table IV. The OF for β -carotene and γ -tocotrienol models were obtained at 0.000149 and 0.0104, respectively, which were very near to zero. The comparison of K_x and K_y is presented in Fig. 2. The deviations were calculated at 5.7% and 4.1% for β -carotene and γ -tocotrienol, respectively.

Fig. 3 shows the comparison between the experimental liquid-liquid equilibrium data and their values as simulated by UNIQUAC thermodynamic model. As can be observed from the figure, good representation of liquid-liquid equilibrium is achieved in β -carotene model. As for the model of γ -tocotrienol, deviations are greater in the solvent-rich phase at higher concentration of γ -tocotrienol when UNIQUAC model parameters underestimated the amount of γ -tocotrienol extracted. An alternative thermodynamic model, non-random two-liquid (NRTL) may be considered for better accuracy even at high γ -tocotrienol concentration. In [24], the comparison of UNIQUAC and NRTL models were presented, it was found that UNIQUAC underestimated the extraction of FFA at high concentration even though no difference was found for tocopherols by assuming infinite dilution in cottonseed oil and ethanol. The average percentage of deviations was calculated at 4.10% and 9.98%, respectively, by (13).

$$100 \sqrt{\frac{\sum_{n=1}^N \sum_{i=1}^C (x_{i,n}^{I,exp} - x_{i,n}^{I,calc})^2 + (x_{i,n}^{II,exp} - x_{i,n}^{II,calc})^2}{2NC}} \quad (13)$$

where N represents the number of tie lines while C represents the number of compounds.

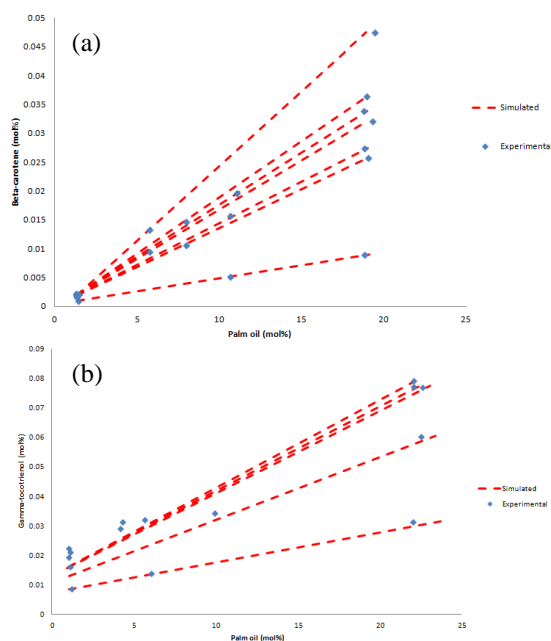

 Fig. 3. The experimental and simulated liquid-liquid equilibrium data for the systems of palm oil + ethyl lactate + β -carotene (a) and γ -tocotrienol (b).

Fig. 4 shows the separation factors as plotted against β -carotene and γ -tocotrienol initial concentration in palm oil. Separation factor is defined by (14) and it is a measure of the system selectivity towards the partition of β -carotene and γ -tocotrienol from oil. It was found that the separation factors are reducing for β -carotene but increasing for γ -tocotrienol.

This suggests that the separation factor improved with lower concentration of carotenes and higher concentration of tococls in palm oil using ethyl lactate as the solvent. This is similar to the results reported by [21] when the separation factors increased with the concentration of α -tocopherol in olive oil using ethyl lactate. The average separation factor as obtained in this work (5.4) is slightly higher than [21] at 4.9. This may be due to the use of higher temperature at 303.15 K as compared to [21] at 298.15 K. As compared to γ -tocotrienol, the average separation factor of β -carotene is lower at 0.9. Similar results were obtained by [23] as more tocopherols were extracted than carotenoids from palm oil using ethanol as the solvent.

$$\text{Separation factor} = \frac{K_{\beta\text{-carotene or } \gamma\text{-tocotrienol}}}{K_{\text{oil}}} \quad (14)$$

With the use of ethyl lactate as the solvent, the tendency towards the extraction of tocotrienol is greater than carotene. Even though both carotene and tocotrienol are non-polar in nature, tocotrienol is relatively more polar due to the presence of the hydroxyl and triene groups. Therefore, it has higher solubility into ethyl lactate, which is polar. By dissolving excessive β -carotene in ethyl lactate at 303.15 K, it was found that the saturation concentration was 310.15 mg/L, which is quite low. Thus, it is the maximum extractive capability of ethyl lactate towards β -carotene. Any increase of the β -carotene concentration will not enhance but reduce the separation factor.

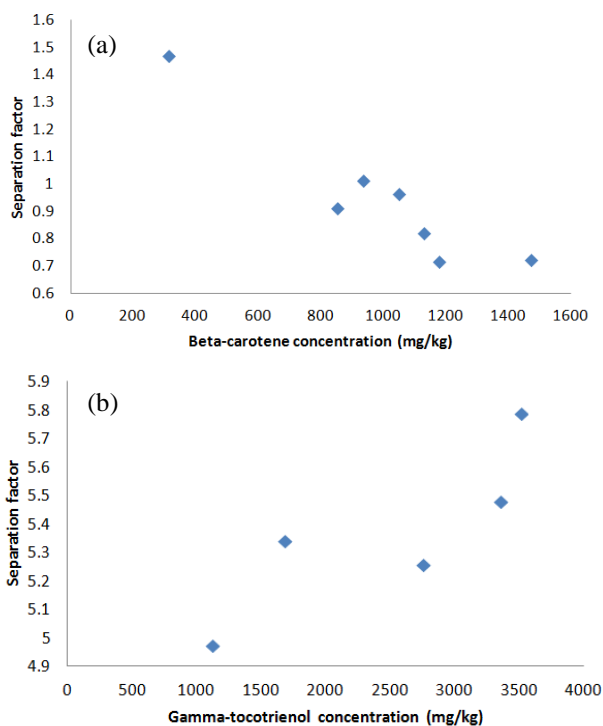


Fig. 4. The separation factor with respect to the initial concentration of β -carotene (a) and γ -tocotrienol (b) in palm oil.

Even though the increase of the mass % of ethyl lactate did not improve the distribution coefficients for both carotene and tocotrienol, the % mass recovery by (15) could be enhanced, provided the mixture (with higher mass % of ethyl lactate) is still within the biphasic condition (within the curve of Fig. 1). As refer to Fig. 5, the increase of the mass % of ethyl lactate promoted the % mass recovery for both carotene

and tocotrienol. For comparison, mass recovery of 16% and 48% of carotene and tocotrienol were achieved at 70 mass% of ethyl lactate. Again, more tocotrienol was extracted due to its higher polarity than carotene.

$$\% \text{ mass recovery} = \frac{\text{mass of } \beta\text{-carotene or } \gamma\text{-tocotrienol extracted (mg)}}{\text{mass of } \beta\text{-carotene or } \gamma\text{-tocotrienol in starting solution (mg)}} \quad (15)$$

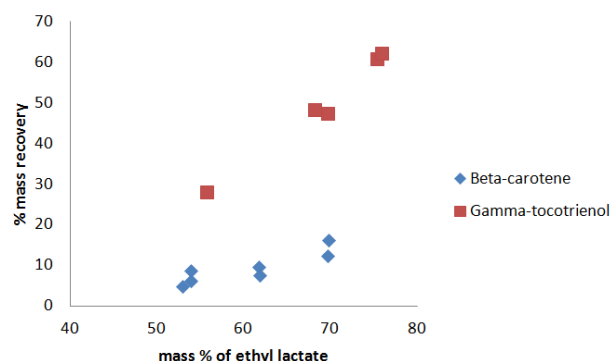


Fig. 5. The percentage mass recovery of β -carotene and γ -tocotrienol with increasing mass percentage of ethyl lactate.

IV. CONCLUSION

UNIQUAC thermodynamic model was demonstrated for the first time for the two ternary systems of palm oil + ethyl lactate + β -carotene and γ -tocotrienol. The presented data enables the design of liquid-liquid extractors for palm carotenes and tococls recovery using ethyl lactate. The studies in this work showed the feasibility of ethyl lactate as a safe and green agrochemical solvent to extract carotenes and tococls from palm oil. The average separation factor of 0.9 and 5.4 were reported for β -carotene and γ -tocotrienol, respectively. Also, recovery of 16% of β -carotene and 48% of γ -tocotrienol were achieved in a single stage at 70 mass% of ethyl lactate. As the implementation of multistage is not uncommon for liquid-liquid extraction, the yield will be further improved. NRTL could be considered to model γ -tocotrienol system in an effort to improve the accuracy at higher concentration.

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