



| Cronfa - Swansea University Open Access Repository | | | | |
|--|--|--|--|--|
| This is an author produced version of a paper published in: International Journal of Nutrition, Pharmacology, Neurological Diseases | | | | |
| Cronfa URL for this paper: http://cronfa.swan.ac.uk/Record/cronfa45998 | | | | |
| Paper: Abu Bakar, M., McKimm, J. & Haque, M. Otitis media and biofilm. <i>International Journal of Nutrition, Pharmacology, Neurological Diseases, 8</i> (3), 70-78. http://dx.doi.org/10.4103/ijnpnd.ijnpnd_28_18 | | | | |
| Released under the terms of a Creative Commons Attribution Non-Commercial Share Alike License. | | | | |

This item is brought to you by Swansea University. Any person downloading material is agreeing to abide by the terms of the repository licence. Copies of full text items may be used or reproduced in any format or medium, without prior permission for personal research or study, educational or non-commercial purposes only. The copyright for any work remains with the original author unless otherwise specified. The full-text must not be sold in any format or medium without the formal permission of the copyright holder.

Permission for multiple reproductions should be obtained from the original author.

Authors are personally responsible for adhering to copyright and publisher restrictions when uploading content to the repository.

http://www.swansea.ac.uk/library/researchsupport/ris-support/

| | _ | _ | • | |
|---|---|-----|-----|--------------|
| 1 | Δ | Rev | /16 | 338 / |
| _ | ~ | ,,, | ,,, | . vv |

| 2 | Chronic Tonsillitis and | Riofilms: A Bri | of Overview of | Treatment Modalities |
|---|-------------------------|-----------------|----------------|----------------------|

| 3 | List | of A | uth | org |
|---|------|------|------|------|
| J | LIJL | UI 7 | uuii | UI 3 |

- 4 Muhamad bin Abu Bakar ¹, Judy McKimm ², Seraj Zohurul Haque ³, Md. Anwarul Azim
- 5 Majumder ⁴, Mainul Haque ⁵
- 1. Professor and Dean, Unit of Otolaryngology, Faculty of Medicine and Defence Health,
 Universiti Pertahanan Nasional Malaysia (National Defence University of Malaysia), Kem
 Sungai Besi, 57000 Kuala Lumpur, Malaysia. Cell Phone: + 60 19 275 3930. Email:
- 9 <u>muhamadbakar@upnm.edu.my</u>
- Professor of Medical Education and Director of Strategic Educational Development,
 Program Director MSc in Leadership for the Health Professions, Swansea University
 School of Medicine, Grove Building, Swansea University, Singleton Park, Swansea, Wales
 SA2 8PP, UK. Email: j.mckimm@swansea.ac.uk
- Final Year Medical Student, University of Dundee, Mackenzie Building, Ninewells
 Hospital & Medical School, Kirsty Semple Way, Dundee DD2 4BF, United Kingdom.
- 16 Email: szzhague@gmail.com
- Director, Medical Education, Faculty of Medical Sciences, The University of the West
 Indies, Cave Hill Campus, Barbados, West Indies. Email:
- 19 <u>azim.majumder@cavehill.uwi.edu</u>
- Professor, Unit of Pharmacology, Faculty of Medicine and Defence Health, Universiti
 Pertahanan Nasional Malaysia (National Defence University of Malaysia), Kem Sungai
 Besi, 57000 Kuala Lumpur, Malaysia, Email: runurono@gmail.com
- 23 Running Title:
- 24 Chronic Tonsillitis and Biofilm
- 25 Key Words:
- 26 Chronic Tonsillitis, Biofilm, Treatment, Modalities
- 27 Address of Correspondence:
- 28 Mainul Haque
- 29 Faculty of Medicine and Defence Health, Universiti Pertahanan Nasional Malaysia (National
- 30 Defence University of Malaysia), Kem Sungai Besi, 57000 Kuala Lumpur, Malaysia, Email:
- 31 <u>runurono@gmail.com</u>
- 32 Word Counts Text: 3082
- 33 Number of References: 149

Review Article

35 Chronic Tonsillitis and Biofilms: A Brief Overview of Treatment Modalities

Abstract

Recurrent tonsillitis is described as when an individual suffers from several attacks of tonsillitis per year. Chronic and recurrent tonsillitis both cause repeated occurrences of inflamed tonsils which have a significant impact on a patients' quality of life. Numerous children suffer from recurrent tonsillitis, and sore throats and these illnesses become part of their life. Antimicrobials can provide temporary relief, but in many cases, tonsillitis recurs. Scientists working at Washington University School of Medicine in St. Louis identified the cause of such recurrent infections as microorganisms which often create biofilms and a repository of infection in the wet and warm folds of the tonsils. This review will discuss different treatment modalities, their advantages and disadvantages and new treatment options focusing on biofilms. All treatment options should be selected based on evidence and individual need.

Tonsillitis

Tonsillitis is an inflammation of the pharyngeal tonsils. The inflammation may affect other areas of the back of the throat, including the adenoids and the lingual tonsils. Acute tonsillitis is an infection of the tonsils triggered by one of several types of bacteria or viruses and peritonsillar abscesses can also occur. Chronic tonsillitis is a tenacious infection of the tonsils which may result in tonsil stones. Recurrent tonsillitis ensues when an individual suffers from several incidents of tonsillitis per year. Both chronic and recurrent tonsillitis involve repeated occurrences of inflamed tonsils which can impact severely on a patients' quality of life. ^{1, 2} Children very often suffer from tonsillitis although it is seldom observed below the age of 2 years. Tonsillitis due to Streptococcus bacteria classically happens in children aged between 5-15 years, while viral tonsillitis is more prevalent in younger children. ³ Multiple studies report that the average prevalence of carrier status of school children for group A Streptococcus is 15.9%. ^{4,5}

Epidemiology of Tonsillitis

Numerous children so often suffer from recurrent tonsillitis and sore throats that these illnesses become their part of life. For example, one study indicates that approximately 30% of peritonsillar abscesses require a tonsillectomy ⁶ and another indicates that recurrent tonsillitis is reported in 11.7% and 12.1% of Norwegian and Turkish children respectively. ⁷ Many of these patients are prescribed antimicrobials which typically provide temporary relief, but then the tonsillitis recurs. ⁸ Scientists working at Washington University School of Medicine in St. Louis identified that recurrent infections are exacerbated by the creation of biofilms in the wet and warm folds of the tonsils by microorganisms which act as a repository of infection. ⁹ A study utilizing an innovative imagining technique in single sections of human mucosal tissue reports the presence of biofilms in 70.8% chronic tonsillitis patients. ¹⁰ Another study revealed that biofilms were recognized on the surface epithelium of tonsils and adenoids in many of the patients who were waiting for adenotonsillectomy due to chronic tonsillitis and adenoiditis. ¹¹ Such biofilms are also observed in other otorhinolaryngology related infections such as chronic rhinosinusitis and chronic otitis media with effusion. ^{12, 13}

A Brief Overview Regarding Biofilms

Biofilms are systematized communities of microorganisms embedded in a hydrated matrix of extracellular polymeric substances causing diverse of persistent infections, including dental plaque, cystic fibrosis, urinary tract infections, osteomyelitis, and ear infections. ^{9, 14, 15} Biofilm formations is a prehistoric prokaryotic strategy of a microorganism to exist and grow in antagonistic settings through building innovative communities through several processes. ¹⁶⁻¹⁹ The Dutch scientist (commonly known as the Father of Microbiology) Antonie van Leeuwenhoek used his primitive but effective microscope to observed Biofilms as early as 1674 and describes aggregates of animalcules scraped from human tooth surfaces. ^{20, 21} The English phrase *'survival of the fittest'* arose from Darwinian evolutionary theory and describes one of the mechanisms of natural selection. ^{22, 23} Bacterial biofilm formations are a form of 'survival of the fittest' under adverse conditions including chemical or antimicrobial treatment. ^{24, 25} The formation of biofilms by bacteria has four potential advantages: "i. *Protection from harmful conditions in the host*, ii. *Sequestration to a nutrient-rich area*, iii. *Utilization of cooperative*

Biofilms normally grow as biofilms and planktonic cultures are an in vitro artifact". ²⁶ Microbial biofilms were identified as a major cause of many human infections, present in more than 65-80% of all human bacterial infections. ^{14, 27-30} Thereafter biofilm pose "a serious problem for public health because of the increased resistance of biofilm-associated organisms to antimicrobial agents and the potential for these organisms to cause infections in patients with indwelling medical devices". 31 Biofilm formations is generally considered to arise in four core stages: (1) bacterial attachment to a surface, (2) microcolony formation, (3) biofilm maturation and (4) detachment (also called dispersal) of bacteria which may then colonize new areas. 32 Multiple research reported that the process of biofilm formation is categorized by five stages. ³³⁻³⁵ (1) Microbial cells attach to surfaces reversibly. ³⁶ (2) Microbial cells then attach to surfaces irreversibly. 37 (3) Cells adsorbed on surfaces and grow into microcolonies, their physical dimensions estimated tens or hundreds of microns in diameter. ³⁸ (4) There microbial fraternity grows into a three-dimensional configuration and settle down into a biofilm as cells replicate and the extracellular polymeric substances (EPS) accumulates. ³⁹ (5) Bacterial cells detach biofilm and disperse into the bulk fluid, where they act free swimming bacteria or and form new biofilms. ^{16, 17} Biofilm formations were depicted in Figure 1 and 2.

Distinct Features of Biofilm Bacteria

Bacteria found inside biofilms have distinct features different from those of free-swimming (planktonic) bacteria of the same classes and possess a very high level of resistance to commonly-used antimicrobial remedies, biocides and antiseptics, and the host immune response. 40-42 Older, mature and impenetrable biofilms are consistently more resistant to antimicrobials than younger, less dense biofilms. 42 Bacterial cells residing in the outermost parts of the biofilm are more vulnerable to the host's defenses and antimicrobials, although these microorganisms possess numerous defensive mechanisms. The biofilm is formed of various microbial communities that create a complex three-dimensional physical barrier which hinders the diffusional penetration of antimicrobials. 17, 43, 44 The exterior layer of biofilm metabolic activity alters the local pH to be more acidic and creates anoxic zones that help to degrade antimicrobials. 45-48 The biofilm also creates nutrient-depleted areas which act on

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

microbes to put them into a stationary or dormant phase, which may also contribute towards antibiotic resistance. ^{49, 50} The extracellular matrix of the biofilm secretes polymers that bind and deactivate antimicrobials, forming an antibiotic "sink". ⁵¹ These properties of biofilms (inadequate diffusion of nutrients, restricted antimicrobial transmission and the alteration of the environment to produce a more hostile environment) combine to produce a widespread resistance and tolerance to antimicrobials. ^{16, 43-56} In addition, microbes entrenched in a biofilm can exist even in high concentrations of bactericidal antimicrobials although they are abundantly sensitive to those antimicrobials in culture plates under planktonic conditions. ⁵⁷ This complex phenomenon is known as the "recalcitrance of biofilm bacteria toward antibiotics" ⁵⁸ and microorganisms found in biofilms can be up to 500-1,000 times more tolerant to antibacterial compounds than their planktonic counterparts. ⁵⁹⁻⁶² Additionally, many studies reported that as soon as a biofilm is rooted and fixed, microbes develop resistance to several categories of physicochemical aggression, including UV light, heavy metals, low pH, changes in hydration or salinity, and phagocytosis. ⁶³⁻⁶⁷

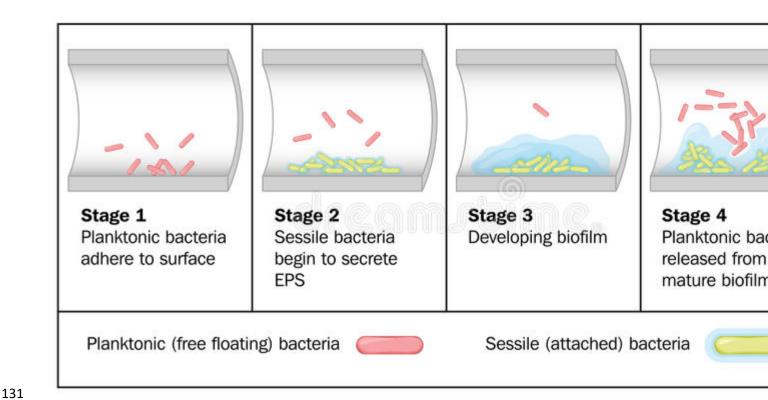
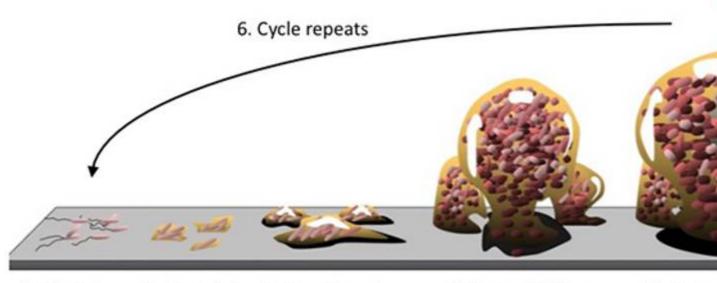


Figure 1: Showing Four Different Stages of Biofilm Development. Image was download from images for a Available

https://www.google.com/search?q=copyright+free+biofilm+image&tbm=isch&tbo=u&source=univ&sa=X&ved
 AhULMO8KHUXIALMQsAQIMA&biw=1280&bih=615&dpr=1.5 [Accessed April 16, 2018]

132



Single free floating bacteria land on surface

2. Bacterial cells aggregate and attach

Growth and division of bacteria for biofilm formation

 Mature biofilm formation 5. Part of disperses to free flo bacteria fo coloniz

137138

139

140

Figure 2: Showing Five Stages of Biofilm Development. Image was download from images for copyright fre https://www.google.com/search?q=copyright+free+biofilm+image&tbm=isch&tbo=u&source=univ&sa=X&vedAhULMo8KHUXIALMQsAQIMA&biw=1280&bih=615&dpr=1.5 [Accessed April 16, 2018]

Recurrent Tonsillitis and Tonsillectomy

Chronic tonsillitis affecting equally both children and adults is a serious health problem ^{68, 69} and whilst the definition of severe recurrent tonsillitis varies, a quantity of severity is described as five or more episodes of true tonsillitis a year, symptoms for at least a year, and episodes that are disabling and prevent normal functioning. ^{70, 71} In one study, the lifetime prevalence of recurrent tonsillitis is described as 11.7% (95% confidence interval, 11.0%-12.3%) with a significant preponderance of females. ⁷ Recurrent tonsillitis is typically treated by either surgery or, when the patient does not meet tonsillectomy benchmarks or there are surgical or medical contraindications, by medical antimicrobial intervention. ^{72, 73}

Whilst tonsillectomy (surgical removal of the tonsils, with or without adenoidectomy) as a treatment modality has been practiced for over 100 years for children, much controversy exists around its value. As for example, in 1951 the British Medical Journal reported that "it is better to delay a decision than to hurry it, and above all to avoid operating on tonsils which have been recently inflamed". ⁷⁴ One study suggests that 0.6 episodes of any type of a sore throat were reported in the first year after surgery compared to medical intervention ⁷⁵ and another reported that surgery could lead to life-threatening complications. A Swedish cohort study reports that among post-tonsillectomy patients 20 years later, there was a higher incidence of "chronic, immune-mediated diseases ... in the operated group", with a statistically significant relationship between post-tonsillectomy and chronic disease, with a relative risk at 9.41 and a confidence interval from 1 (1.13 < RR < 78.14). ⁷⁶ However, another research study focusing on adults found that tonsillectomy promotes and improves long-term health and quality of life, thus saving health resources. ⁷⁷

The decision to operate should therefore be taken with care based on an individual patient's needs and history, plus current research evidence. ^{74, 76, 78, 79} In making such decisions, secondary care doctors and family medicine practitioners need to collaborate because the decision whether a tonsillectomy is necessary is quite difficult and both the GP and the otolaryngologist must contribute equally. ⁷⁴ The GP knows about the patient's frequency, duration and severity of tonsillitis whereas the ENT specialist will evaluate symptoms relating to

nasal and Eustachian impediment, and will assess whether symptoms are due to tonsillitis or chronic sinusitis. ⁷⁴

Treatments Aimed at Disrupting Biofilms

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

Microbial biofilm formation is responsible for the development of acute to chronic infection in several diseases including cystic fibrosis; periodontitis; infective endocarditis; persistent otitis media; chronic rhinosinusitis; chronic tonsillitis; prostatitis; chronic osteomyelitis; atopic dermatitis; onychomycosis; dental caries; infectious kidney stones; and chronic wounds. 80-83 As well, biofilms formed on the any surface, living or non-living, even on clinical devices like pacemakers, implants and catheters, and very difficult to eradicate, which accentuates the clinical consequence, such as Pseudomonal infections can embroil any part of the human body. Further, the micro-organisms adaptive capability and genetic ups and downs of within the biofilm transform them resistant to all known antimicrobial medicines. Thereafter the Pseudomonal infections become real critical to be handle by the medical doctors and threatens human life. 83, 84 By and large it is thought that 99% of the biosphere's bacteria to live in biofilms. Thereafter, it is believed that microbial community gain an advantage living in this state. 85 Consequently, microbial biofilms significantly affecting human health by increasing morbidity, mortality, and healthcare cost. Biofilm not only adding to hospital-acquired infections (HAIs) by increasing chronicity and persistence, but colonizing in other areas of environment instigating corrosion, fouling of water pipes, and food and pharmaceutical decomposition.^{14, 86-88} Another study reported microbial biofilm can stick and infect all medical devices such as orthopedic prostheses and intravascular catheters and promote up to 60% of HAIs. 89

Microorganisms in biofilms are distinctively more resistant to antimicrobial agents and environmental insults and are therefore very difficult to eradicate. ^{42, 90-94} Biofilms in general (and chronic tonsillitis specifically) can therefore lead to substantial economic costs for countries and individuals, health concerns and an evolving public health problem in both high and low resource settings. ^{77, 95-100} Because of this, multiple research studies have attempted to resolve the issues of both biofilms and recurrent tonsillitis. ^{59, 61, 101-108}

The explosion of antibiotic resistance throughout the world of many microbial strains has put pressure on the research and medical communities to find an alternative strategy for the management of biofilm-mediated diseases. 61 "Perhaps new antibiotics are not the only way to combat biofilm infections if we could make ineffective older antibiotics active again." 59 This researcher developed a 2-amino-imidazoles molecule which is capable of disrupting biofilms through making a microorganism which was previously antibiotic-resistant more vulnerable to older antimicrobials. ^{59, 62} Immunotherapy (using cyclic di-nucleotides) has been effective in the management of different cancers, and this molecule has also been utilized as a therapeutic strategy for biofilm-related infections. Immunoprophylaxis and immunotherapy might therefore provide new tools to combat S. epidermidis biofilm formation. 109, 110 Recently, multiple studies revealed that a 3,5-cyclic diguanylic acid (c-di-GMP) binding protein was found in biofilm communities. ^{111, 112} BdcA (a protein that enhances biofilm dispersal), confiscates c-di-GMP and minimizes its local concentration and is partly responsible for the reduction and down-regulation of EPS of biofilms and for the up-regulation of swimming, swarming, and planktonic microbes. 111, 112 This phenomenon has been observed in Pseudomonas species and the Rhizobium mellioti biofilm communities. 111, 112 Multiple group of scientists recently reported that CdrA (an adhesin compound) which is produced by biofilms in response to high levels of c-di-GMP that binds with Psl and stabilizes biofilm structure. 38, 106, 113 Multiple research studies have identified at least three extracellular polysaccharides (Alginate, Pel and Psl) that have been important implication in structure maintenance and antibiotic resistance of biofilm. 114-123 Another study revealed that exogenous addition of D-amino acids 109 disrupted preformed biofilms by disturbing adhesive fiber interactions and was also effective in preventing biofilm formation by S. aureus and P. aeruginosa. 124-126 One-more biofilmdisassembly molecule is norspermidine which has a similar dispersal mechanism to D-amino acids by targeting the exopolysaccharides. ¹²⁵ The biofilm-inhibiting properties of norspermidine were detected in Staphylococcus Aureus and Escherichia coli pellicle biofilm. 125 Current research therefore needs to focus on the development of norspermidine, BdcA, D-amino acids, and other polyamines as a novel antibiofilm approach and medical communities should no longer depend exclusively on antimicrobials (which are increasingly ineffective with many

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

pathogenic microorganisms because of resistance) and surgery to treat infectious diseases. ^{104,} _{111, 112, 124, 125}

Other studies have identified additional ways of disrupting biofilms. Bioactive enzymes such as dispersin or Proteinase K studied in orthopedic implants made bacteria more susceptible to antibiotics and finally eradicated the biofilm by affecting polymers or proteins of the biofilm structure. 127 Several cytotoxic agents have also been found to successfully eliminate biofilms from implant surfaces, with citric acid being reported to be the most successful in eradicating biofilms on titanium surfaces. ¹²⁸ Multiple research studies have identified that an electrical current successfully detaches Staphylococcus aureus and Staphylococcus epidermis biofilms from stainless steel implants. 129-131 Another study observed that biofilms of Staphylococcus epidermis on stainless steel fasteners were successfully eradicated through pulsed electromagnetic fields in combination with gentamicin. ¹³² A new cluster of research studies have used laser-generated shockwaves to effectively break up biofilms. 133 The technique is founded on a Q-switched, ND: YAG rhythmically laser functioning at a "rep rate of 10 Hz with 1500 mJ pulses centered at 1064 nm. The laser pulses were used to create shockwave pulses in Al coated polycarbonate substrates and a resulting peak stress of greater than 50 MPa" was able to reduce 55% living microorganisms. 134 The laser technique offers another way of disrupting biofilms and is useful in the management of infected wounds, where standard treatment modalities such as topical antimicrobials or the removal of dead, damaged, or infected tissue is unsuccessful or injurious. One study found that just 4-10 seconds of the laser therapy was able to disperse biofilms from nitinol stents on 97.9% of Pseudomonas aeruginosa to single-celled planktonic microorganisms that can be more easily treated with antibiotics. 135 Another found that laser-generated shockwaves therapy quickly disrupts the biofilms in infected wounds to eliminate the microorganisms and intensify the effectiveness of topical antimicrobials in the residual biofilm. Such interventions will promote patients' quality of life by reducing healing times, morbidity, and save healthcare costs. 136

N-Acetyl-Cysteine (NAC) is an antioxidant mediator which reduces the variety of microbial bacteria on biofilm emergence and evolution, ¹³⁷ inhibits the manufacturing of the extracellular

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

polysaccharide matrix ¹³⁸ and promotes the disruption of mature biofilms. ¹³³ NAC has been found to reduce Streptococcus pneumoniae and Haemophilus influenzae adhesion to human oropharyngeal epithelial cells in laboratory experiments. 138 Chronic infections raise prostaglandin levels and NAC effectively reduces these levels and helps to disrupt the biofilms. ¹³⁹⁻¹⁴² correspondingly, aspirin-like non-steroidal anti-inflammatory drugs (NSAIDs) decrease biofilm production and completely block fungal infections. ¹⁴³ NAC interacts with the sulfhydryl group of enzymes involved in EPS production or excretion, which reduces the activity of these molecules or inhibits cysteine utilization. ¹⁴⁴ NAC therefore, decreases in-vitro biofilm formation ¹⁴⁵ and research on salicylates shows a similar negative effect on the production of biofilm. ¹⁴⁶ A study which applied both found that therapeutic doses of acetylsalicylic acid (ASA) and NAC diminishes tonsillar mucosal biofilm formation in chronic or recurrent tonsillitis. 102 An Iraqi study found a strong correlation between the biofilm of Streptococcus pyogenes and recurrent tonsillitis and that three types of vinegar eradicated streptococcal biofilm remarkably: Date (100%), Apple (95.5%), and Grape (90.9%). ¹⁰⁵ A later study also demonstrated the potential of vinegar in eradicating tonsillar biofilm. 101 In a laboratory experiment, whilst washing and cleaning with a soft brush did not remove the chronic tonsillitis biofilm layer on the tonsil surface in, using a harder brush removed more biofilm. ¹⁰³ Researchers believe that the physical removal of biofilm (by brushing or using ultrasound-activated bubbles) from the tonsil surface in vivo will lead to greater effectiveness of topical antimicrobials and decrease the need for systemic antimicrobials. ¹⁰³

Conclusion

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

Recurrent or chronic tonsillitis is currently a global public health issue which can severely impair an individuals' quality of life. ^{77, 147} Microbial biofilms are a major cause of repeated tonsillitis in both pediatric and adult cohorts and more research is needed to develop new treatment strategies. ^{107,148, 149} Treatment modalities should however be based on careful selection and individual consideration of the potential impact of biofilms on cases of recurrent tonsillitis. ⁷⁴ Rather than developing or using more potent antimicrobials, doctors should ensure they are

- up-to-date with research and the treatment of biofilms, including the application of topical
- agents, the physical removal of biofilms and other innovative treatments.
- 284 Conflict of Interest
- 285 Authors declare no conflict of Interest.
- 286 Funding
- This manuscript obtained no financial support from any source.
- 288 **References**
- 289 1. American Academy of Otolaryngology. Tonsillitis. 2018. Available at 290 http://www.entnet.org/content/tonsillitis [Accessed January 6, 2018]
- 291 2. Hayes K. Chronic and Recurrent Tonsillitis: What to Know. 2017. Available at
 292 https://www.verywell.com/chronic-and-recurrent-tonsillitis-1191984 [Accessed January 6,
 293 2018]
- Shah UK. Tonsillitis and Peritonsillar Abscess. Drugs & Diseases. Otolaryngology and Facial
 Plastic Surgery. Medscape. Available at https://emedicine.medscape.com/article/871977-overview#a6 [Accessed January 6, 2018]
- 4. Pichichero ME, Casey JR. Defining and dealing with carriers of group A Streptococci.
 Contemp Pediatr. 2003 Available at
- 299 http://contemporarypediatrics.modernmedicine.com/contemporary-
- 300 <u>pediatrics/news/clinical/clinical-pharmacology/defining-and-dealing-carriers-group-stre</u>
- 301 [Accessed January 6, 2018]
- 5. Wald ER. Commentary: Antibiotic treatment of pharyngitis. Pediatr Rev 2001; 22 (8):255-303 256.
- 6. Herzon FS. Peritonsillar abscess: incidence, current management practices, and a proposal for treatment guidelines. Laryngoscope. 1995; 105(8 Pt 3 Suppl 74):1-17.
- 7. Kvestad E, Kvaerner KJ, Roysamb E, Tambs K, Harris JR, Magnus P. Heritability of recurrent tonsillitis. Arch Otolaryngol Head Neck Surg 2005; 131 (5): 383-87.
- 8. Ward D. Bacterial biofilms may be source of recurrent tonsillitis. Medicine & Health.
 Washington University in St. Louis, 2018. Available at https://source.wustl.edu/2003/09/bacterial-biofilms-may-be-source-of-recurrent-
- 311 <u>tonsillitis/</u> [Accessed January 6, 2018]

- 9. Chole RA, Faddis BT. Anatomical Evidence of Microbial Biofilms in Tonsillar Tissues a Possible Mechanism to Explain Chronicity. Arch Otolaryngol Head Neck Surg 2003; 129(6):
- 314 634-36. DOI: 10.1001/archotol.129.6.634
- 10. Kania RE, Lamers GE, Vonk MJ, Huy PT, Hiemstra PS, Bloemberg GV, Grote JJ.

 Demonstration of Bacterial Cells and Glycocalyx in Biofilms on Human Tonsils. Arch
- 317 Otolaryngol Head Neck Surg 2007; 133 (2): 115-121.
- 11. Al-Mazrou KA, Al-Khattaf AS Adherent Biofilms in Adenotonsillar Diseases in Children. Arch Otolaryngol Head Neck Surg 2008; 134(1): 20-23.
- 12. Saylam G, Tatar EC, Tatar I, Özdek A, Korkmaz H. Association of Adenoid Surface Biofilm
- Formation and Chronic Otitis Media with Effusion. Arch Otolaryngol Head Neck Surg 2010;
- 322 **136 (6)**: 550-555.
- 13. Sanderson AR, Leid JG, Hunsaker D. Bacterial Biofilms on the Sinus Mucosa of Human Subjects with Chronic Rhinosinusitis. Laryngoscope 2006;116(7):1121-6.
- 14. Costerton JW, Stewart PS, Greenberg EP. Bacterial biofilms: a common cause of persistent infections. Science 1999; 284 (5418): 1318-1322.
- 15. Donlan RM, Costerton JW. Biofilms: survival mechanisms of clinically relevant microorganisms. Clin Microbiol Rev 2002; 15 (2): 167–193
- 16. Hall-Stoodley L, Costerton JW, Stoodley P. Bacterial biofilms: from the environment to infectious disease. Nat Rev Microbiol 2004; 2 (2): 95-108.
- 17. Hall-Stoodley L, Stoodley P. Biofilm formation and dispersal and the transmission of human pathogens. Trends Microbiol 2005; 13 (1): 7-10.
- 18. Purevdorj-Gage B, Costerton WJ, Stoodley P. Phenotypic differentiation and seeding dispersal in nonmucoid and mucoid Pseudomonas aeruginosa biofilms. Microbiology 2005; 151 (Pt 5): 1569-1576.
- 19. Mai-Prochnow A, Lucas-Elio P, Egan S, Thomas T, Webb JS, Sanchez-Amat A, Kjelleberg S. Hydrogen peroxide linked to lysine oxidase activity facilitates biofilm differentiation and dispersal in several gram-negative bacteria. J Bacteriol 2008; 190 (15): 5493-5501.
- 20. Borhan WM, Dababo MA, Thompson LDR, Saleem M, Pashley N. Acute Necrotizing Herpetic Tonsillitis: A Report of Two Cases. Head Neck Pathol 2015; 9(1): 119-122. doi:10.1007/s12105-013-0516-2.
- 21. Slavkin HC. Biofilms, microbial ecology, and Antoni van Leeuwenhoek. J Am Dent Assoc. 1997; 128(4): 492-95.
- 22. Fasolo A. The Theory of Evolution and Its Impact. Springer Milan Dordrecht HeidelbergLondon, 2012.
- 23. Neumann JJ. The Role of Metaphor in The Darwin Debates: Natural Theology, Natural Selection, And Christian Production of Counter-Metaphor. Master of Arts Thesis. Texas A&M University, 2012. Available at

- http://oaktrust.library.tamu.edu/bitstream/handle/1969.1/ETD-TAMU-2012-05-10729/NEUMANN-THESIS.pdf?sequence=2 [Accessed January 9, 2018]
- 24. Tilahun A, Haddis S, Teshale A, Hadush T. Review on Biofilm and Microbial Adhesion. Int J Microbiol Res 2016; 7 (3): 63-73.
- 25. Brown MRW, Gilbert P. Microbiological Quality Assurance: A Guide Towards Relevance and Reproducibility of Inocula. CRC Press, Boca Raton, New York, 1995.
- 26. Jefferson KK. What drives bacteria to produce a biofilm? FEMS Microbiol Lett 2004 15; 236(2):163-73. DOI: 10.1016/j.femsle.2004.06.005
- 27. Chambers JR, Sauer K. The MerR-Like Regulator BrlR Impairs Pseudomonas aeruginosa Biofilm Tolerance to Colistin by Repressing PhoPQ. J Bacteriol 2013; 195(20): 4678-4688. doi:10.1128/JB.00834-13.
- 28. Joo H-S, Otto M. Molecular basis of in-vivo biofilm formation by bacterial pathogens. Chem Biol 2012; 19(12): 1503-1513. doi: 10.1016/j.chembiol.2012.10.022.
- 29. Lebeaux D, Chauhan A, Rendueles O, Beloin C. From in vitro to in vivo Models of Bacterial Biofilm-Related Infections. Pathogens 2013; 2(2): 288-356. doi:10.3390/pathogens2020288.
- 30. Costerton JW. Introduction to biofilm. Int J Antimicrob Agents 1999; 11 (3-4): 217-21.
- 31. Donlan RM. Biofilm Formation: A Clinically Relevant Microbiological Process. Clin Infect Dis 2001; 33(8): 1387-92. DOI: 10.1086/322972
- 32. Landini P, Antoniani D, Burgess JG, Nijland R. Molecular mechanisms of compounds affecting bacterial biofilm formation and dispersal. Appl Microbiol Biotechnol 2010; 86(3): 813-23. doi: 10.1007/s00253-010-2468-8.
- 33. Renner LD, Weibel DB. Physicochemical regulation of biofilm formation. MRS Bull 2011; 36(5): 347-355. doi:10.1557/mrs.2011.65.
- 34. Banerjee P, Singh M, Sharma V. Biofilm Formation: A Comprehensive Review. Int J Pharm Res Health Sci 2015; 3 (2): 556-560.
- 35. Sauer K, Camper AK, Ehrlich GD, Costerton JW, Davies DG. Pseudomonas aeruginosa Displays Multiple Phenotypes during Development as a Biofilm. J Bacteriol 2002; 184(4): 1140-1154. doi:10.1128/jb.184.4.1140-1154.2002.
- 36. Thomas WE, Nilsson LM, Forero M, Sokurenko EV, Vogel V. Shear-dependent 'stick-and-roll' adhesion of type 1 fimbriated Escherichia coli. Mol Microbiol. 2004; 53 (5): 1545-57. DOI: 10.1111/j.1365-2958.2004.04226.x
- 37. Flemming HC, Wingender J. The biofilm matrix. Nat Rev Microbiol 2010; 8 (9): 623-33. doi: 10.1038/nrmicro2415.
- 38. Borlee BR, Goldman AD, Murakami K, Samudrala R, Wozniak DJ, Parsek MR. Pseudomonas 384 aeruginosa uses a cyclic-di-GMP-regulated adhesin to reinforce the biofilm extracellular 385 matrix. Mol Microbiol 2010; 75 (4): 827-842. doi:10.1111/j.1365-2958.2009.06991.x.

- 39. Alpkvist E, Picioreanu C, van Loosdrecht MC, Heyden A. DJ. Three-dimensional biofilm model with individual cells and continuum EPS matrix. Biotechnol Bioeng 2006; 94 (5): 961-79. doi:10.1002/bit.20917
- 389 40. Hentzer M, Givskov M. Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. J Clin Invest 2003; 112(9): 1300-1307. doi:10.1172/JCI200320074.
- 41. Chole RA, Faddis BT. Evidence for Microbial Biofilms in Cholesteatomas. Arch Otolaryngol Head Neck Surg. 2002; 128 (10): 1129-33.
- 42. Stewart PS. Antimicrobial Tolerance in Biofilms. Microbiol Spectr 2015; 3(3): 10.
- 43. McConoughey SJ, Howlin R, Granger JF, Manring MM, Calhoun JH, Shirtliff M, Kathju S, Stoodley P. Biofilms in periprosthetic orthopedic infections. Future Microbiol 2014; 9(8): 987-1007. doi:10.2217/fmb.14.64.
- 44. Stoodley P, Sauer K, Davies DG, Costerton JW. Biofilms as complex differentiated communities. Annu Rev Microbiol 2002; 56: 187–209.
- 45. Huang CT, Yu FP, Mcfeters GA, Stewart PS. Nonuniform spatial patterns of respiratory activity within biofilms during disinfection. Appl Environ Microbiol 1995; 61(6): 2252-2256.
- 46. de Beer D, Stoodley P. Relation between the structure of an aerobic biofilm and mass transport phenomena. Water Sci Technol 1995; 32 (8): 11-18.
- 47. de Beer D, Stoodley P, Lewandowski Z. Measurement of local diffusion coefficients in biofilms by microinjection and confocal microscopy. Biotechnol Bioeng 1997; 53 (2): 151-158.
- 48. Stoodley P, Wefel J, Gieseke A, DeBeer D, von Ohle C. Biofilm plaque and hydrodynamic effects on mass transfer, fluoride delivery, and caries. J Am Dent Assoc 2008; 139 (9): 1182-1190.
- 49. Walters MC, Roe F, Bugnicourt A, Franklin MJ, Stewart PS. Contributions of antibiotic penetration, oxygen limitation, and low metabolic activity to tolerance of Pseudomonas aeruginosa biofilms to Ciprofloxacin and Tobramycin. Antimicrob Agents Chemother 2003; 47 (1): 317-32.
- 50. Fux CA, Wilson S, Stoodley P. Detachment characteristics and oxacillin resistance of Staphylococcus aureus biofilm emboli in an in vitro catheter infection model. J Bacteriol 2004; 186 (14): 4486-4491
- 417 51. Hoiby N. Recent advances in the treatment of Pseudomonas aeruginosa infections in cystic 418 fibrosis. BMC Med 2011; 9: 32.
- 52. Anwar H, Strap JL, Costerton JW. Establishment of aging biofilms: a possible mechanism of bacterial resistance to antimicrobial therapy. Antimicrob Agents Chemother 1992; 36 (7): 1347–1351.

- 53. Borriello G, Werner E, Roe F, Kim AM, Ehrlich GD, Stewart PS. Oxygen limitation contributes to antibiotic tolerance of Pseudomonas aeruginosa in biofilms. Antimicrob Agents Chemother 2004; 48 (7): 2659-2664.
- 54. Brown MR, Allison DG, Gilbert P. Resistance of bacterial biofilms to antibiotics: a growthrate related effect? J Antimicrob Chemother 1988; 22 (6): 777-783.
- 55. Shah D, Zhang Z, Khodursky A, Kaldalu N, Kurg K, Lewis K. Persisters: a distinct physiological state of E. coli. BMC Microbiol 2006; 12: 53.
- 56. Stewart PS, Costerton JW. Antibiotic resistance of bacteria in biofilms. Lancet 2001; 358 (9276): 135-138.
- 57. Anderl JN, Franklin MJ, Stewart PS. Role of antibiotic penetration limitation in Klebsiella pneumoniae biofilm resistance to ampicillin and ciprofloxacin. Antimicrob. Agents Chemother 2000; 44 (7): 1818-1824. Doi.org/10.1128/AAC.44.7.1818-1824.2000
- 58. Lebeaux D, Ghigo J-M, Beloin C. Biofilm-Related Infections: Bridging the Gap between Clinical Management and Fundamental Aspects of Recalcitrance toward Antibiotics. Microbiol Mol Biol Rev 2014; 78(3): 510-543. doi:10.1128/MMBR.00013-14.
- 59. Potera C. Antibiotic Resistance: Biofilm Dispersing Agent Rejuvenates Older Antibiotics. Environ Health Perspect 2010; 118(7): A288.
- 60. Sedlacek MJ, Walker C. Antibiotic resistance in an in vitro subgingival biofilm model. Oral Microbiol Immunol 2007; 22(5): 333-339. doi:10.1111/j.1399-302X.2007.00366.x.
- 61. Worthington RJ, Richards JJ, Melander C. Small molecule control of bacterial biofilms. Org Biomol Chem 2012; 10 (37): 7457-7474. doi:10.1039/c2ob25835h.
- 443 62. Rogers SA, Huigens RW, Cavanagh J, Melander C. Synergistic Effects between Conventional 444 Antibiotics and 2-Aminoimidazole-Derived Antibiofilm Agents. Antimicrob Agents 445 Chemother. 2010; 54 (5): 2112-2118. doi:10.1128/AAC.01418-09.
- 446 63. Espeland EM, Wetzel RG. Complexation, stabilization, and UV photolysis of extracellular 447 and surface-bound glucosidase and alkaline phosphatase: implications for biofilm 448 microbiota. Microb Ecol 2001; 42 (4): 572-585.
- 64. Le Magrex-Debar E, Lemoine J, Gelle MP, Jacquelin LF, Choisy C. Evaluation of biohazards in dehydrated biofilms on foodstuff packaging. Int J Food Microbiol 2000; 55 (1-3): 239-243.
- 65. Leid JG, Shirtliff ME, Costerton JW, Stoodley P. Human leukocytes adhere to, penetrate, and respond to Staphylococcus aureus biofilms. Infect Immun 2002; 70 (11): 6339-6345.
- 453 66. McNeill K, Hamilton IR. Acid tolerance response of biofilm cells of Streptococcus mutans. 454 FEMS Microbiol Lett 2003; 221 (1): 25-30.
- 455 67. Teitzel GM, Parsek MR. Heavy metal resistance of biofilm and planktonic Pseudomonas aeruginosa. Appl Environ Microbiol 2003; 69 (4): 2313-2320.
- 68. Wagner S, Jung H, Nau F, Schmitt H. Relevance of infectious diseases in a pediatric practice.
 Klin Padiatr 1993; 205 (1): 14-17

- 69. Potera C. Forging a link between biofilms and disease. Science 1999; 283 (5409): 1837-460 1839.
- 70. Management of sore throat and indications for tonsillectomy. National Clinical Guideline
 No 34. Scottish Intercollegiate Guidelines Network, Royal College of Physicians, 9 Queen
 St., Edinburgh EH2 1JQ. Available at
 http://www.sdl.academic.chula.ac.th/Sore%20Throat/Sign.pdf [Accessed January 11, 2018]
- 465 71. McKerrow WS. Recurrent tonsillitis. Am Fam Physician 2002; 66 (9): 1735-1736.
- 72. El Hennawi DED, Geneid A, Zaher S, Ahmed MR. Management of recurrent tonsillitis in children. Am J Otolaryngol 2017; 38(4): 371-374. doi: 10.1016/j.amjoto.2017.03.001.
- 468 73. Georgalas CC, Tolley NS, Narula A. Tonsillitis. BMJ Clin Evid 2009; 2009: 0503. 2014.
- 74. Gale AH. Refresher Course for General Practitioners Pros and Cons of Tonsillectomy. Br Med J 1951; 1 (4698): 133-135. doi: https://doi.org/10.1136/bmj.1.4698.
- 75. Burton MJ, Glasziou PP, Chong LY, Venekamp RP. Tonsillectomy or adenotonsillectomy versus non-surgical treatment for chronic/recurrent acute tonsillitis. Cochrane Database Syst Rev 2014; 19 (11): CD001802. doi: 10.1002/14651858.CD001802.pub3.2014
- 76. Johansson E, Hultcrantz E. Tonsillectomy--clinical consequences twenty years after surgery? Int J Pediatr Otorhinolaryngol 2003; 67 (9): 981-88.
- 77. Senska G, Atay H, Pütter C, Dost P. Long-Term Results from Tonsillectomy in Adults. Dtsch Arztebl Int 2015; 112(50): 849-855. doi:10.3238/arztebl.2015.0849.
- 78. Stuck BA, Windfuhr JP, Genzwürker H, Schroten H, Tenenbaum T, Götte K. Tonsillectomy in Children. Dtsch Arztebl Int 2008; 105 (49): 852-861. doi:10.3238/arztebl.2008.0852.
- 79. Windfuhr JP. Indications for tonsillectomy stratified by the level of evidence. GMS Curr Top Otorhinolaryngol Head Neck Surg 2016; 15: Doc09. doi:10.3205/cto000136.
- 482 80. Aparna MS, Yadav S. Biofilms: microbes and disease. Braz J Infect Dis 2008; 12(6): 526-30.
- 483 81. Soto SM. Importance of Biofilms in Urinary Tract Infections: New Therapeutic Approaches.
 484 Adv Biol 2014; 2014: Article ID 543974, 1-13.
- 485 82. Zhao G, Usui ML, Lippman SI, James GA, Stewart PS, Fleckman P, Olerud JE. Biofilms and Inflammation in Chronic Wounds. Adv Wound Care 2013; 2(7): 389-399.
- 487 83. Reyes-Darias JA, Krell T. Riboswitches as Potential Targets for the Development of Anti-Biofilm 488 Drugs. Curr Top Med Chem 2017; 17 (17): 1945-1953. doi: 10.2174/1568026617666170407163517. 489
- 490 84. Sharma G, Rao S, Bansal A, Dang S, Gupta S, Gabrani R. Pseudomonas aeruginosa biofilm: 491 potential therapeutic targets. Biologicals 2014; 42(1): 1-7. doi: 492 10.1016/j.biologicals.2013.11.001.
- 493 85. Donlan RM. New approaches for the characterization of prosthetic joint biofilms. Clin 494 Orthop Relat Res 2005; 437: 12–9. doi:10.1097/01.blo.0000175120.66051.29

- 495 86. Figueiredo AMS, Ferreira FA, Beltrame CO, Côrtes MF. The role of biofilms in persistent 496 infections and factors involved in ica-independent biofilm development and gene 497 regulation in Staphylococcus aureus. Crit Rev Microbiol. 2017; 43(5): 602-620. doi: 498 10.1080/1040841X.2017.1282941.
- 499 87. Zumstein V, Betschart P, Albrich WC, Buhmann MT, Ren Q, Schmid HP, Abt D. Biofilm 500 formation on ureteral stents - Incidence, clinical impact, and prevention. Swiss Med Wkly 501 2017; 147: w14408. doi: 10.4414/smw.2017.14408.
- 502 88. Vickery K, Hu H, Jacombs AN, Bradshaw DA, Deva AK. A review of bacterial biofilms and 503 their role in device associated infection. Healthcare Infection 2013; 18 (2): 61-66. Doi: 504 org/10.1071/HI12059
- 89. Bryers JD. Medical biofilms. Biotechnol Bioeng 2008; 100(1): 1–18. doi:10.1002/bit.21838
- 90. Lewis K. Riddle of Biofilm Resistance. Antimicrob Agents Chemother 2001; 45(4): 999-1007.
 doi:10.1128/AAC.45.4.999-1007.2001.
- 508 91. Spoering AL, Lewis K. Biofilms and Planktonic Cells of Pseudomonas Aeruginosa Have 509 Similar Resistance to Killing by Antimicrobials. J Bacteriol 2001;183(23):6746-6751. 510 doi:10.1128/JB.183.23.6746-6751.2001.
- 92. Davey ME, O'toole GA. Microbial Biofilms: from Ecology to Molecular Genetics. Microbiol Mol Biol Rev 2000; 64(4): 847-867.
- 93. Bridier A, Briandet R, Thomas V, Dubois-Brissonnet F. Resistance of bacterial biofilms to disinfectants: a review. Biofouling 2011; 27(9): 1017-32. doi: 10.1080/08927014.2011.626899.
- 94. El Khatib M, Tran QT, Nasrallah C, Lopes J, Bolla JM, Vivaudou M, Pagès JM, Colletier JP. Providencia stuartii form biofilms and floating communities of cells that display high resistance to environmental insults. PLoS One 2017; 23; 12(3): e0174213. doi: 10.1371/journal.pone.0174213.
- 95. Zhou G, Shi Q-S, Huang X-M, Xie X-B. The Three Bacterial Lines of Defense against Antimicrobial Agents. Boix E, ed. Int J Mol Sci 2015; 16 (9): 21711-21733. doi:10.3390/ijms160921711.
- 96. Fish KE, Osborn AM, Boxall J. Characterizing and understanding the impact of microbial biofilms and the extracellular polymeric substance (EPS) matrix in drinking water distribution systems. Environ Sci Water Res Technol 2016; 2: 614-630. DOI: 10.1039/C6EW00039H
- 527 97. Sadekuzzaman M, Yang S, Mizan MFR, Ha SD. Current and Recent Advanced Strategies for 528 Combating Biofilms. Comp Rev Food Sci Food Safety 2015; 14 (4): 491-509. DOI: 529 10.1111/1541-4337.12144
- 98. Zhao X, Zhaoa F, Wang J, Zhong Z. Biofilm formation and control strategies of foodborne pathogens: food safety perspectives. RSC Adv 2017; 7: 36670-36683. DOI: 10.1039/C7RA02497E

- 533 99. Duarte VM, McGrath CL, Shapiro NL, Bhattacharrya N. Healthcare costs of acute and chronic tonsillar conditions in the pediatric population in the United States. Int J Pediatr Otorhinolaryngol 2015; 79(6): 921-5. doi: 10.1016/j.ijporl.2015.04.019.
- 536 100. Stelter K. Tonsillitis and sore throat in children. GMS Curr Top Otorhinolaryngol Head 537 Neck Surg 2014; 13: Doc07. doi: 10.3205/cto000110
- 538 101. Al-Saadi MAK, Abdul-Lateef LA, Kareem MA. Detection of biofilm formation and effect of 539 vinegar on biofilm of Streptococcus pyogenes isolated from patients with tonsillitis. Int J 540 Pharm Tech Res 2016; 9 (9): 236-242.
- 541 102. Bulut F, Meric F, Yorgancilar E, Nergiz Y, Akkus M, Nergiz S, Nasir Y. Effects of N-acetyl-542 cysteine and acetylsalicylic acid on the tonsil bacterial biofilm tissues by light and electron 543 microscopy. Eur Rev Med Pharmacol Sci 2014; 18(23): 3720-5.
- 544 103. Ciftci Z, Develioglu O, Arbak S, Ozdoganoglu T, Gultekin E. A new horizon in the 545 treatment of biofilm-associated tonsillitis. Ther Adv Respir Dis 2014; 8(3): 78-83. DOI: 546 10.1177/1753465814529177
- 547 104. Connaughton A, Childs A, Dylewski S, Sabesan VJ. Biofilm Disrupting Technology for 548 Orthopedic Implants: What's on the Horizon? Front Med 2014; 1: 22. doi: 549 10.3389/fmed.2014.00022
- 105. Ismael NF. "Vinegar" as Anti-Bacterial Biofilm formed by Streptococcus pyogenes Isolated from Recurrent Tonsillitis Patients, in vitro. Jordan J Biol Sci 2013; 6 (3): 191-197.
- 552 106. Kostakioti M, Hadjifrangiskou M, Hultgren SJ. Bacterial Biofilms: Development, 553 Dispersal, and Therapeutic Strategies in the Dawn of the Postantibiotic Era. Cold Spring 554 Harb Perspect Med 2013; 3(4): a010306. doi: 10.1101/cshperspect.a010306.
- 107. Römling U, Balsalobre C. Biofilm infections, their resilience to therapy and innovative treatment strategies. J Intern Med 2012; 272(6): 541-61. doi: 10.1111/joim.12004.
- 557 108. Wu H, Moser C, Wang H-Z, Høiby N, Song Z-J. Strategies for combating bacterial biofilm 558 infections. Int J Oral Sci 2015; 7(1): 1-7. doi: 10.1038/ijos.2014.65
- 109. Topalian SL, Weiner GJ, Pardoll DM. Cancer immunotherapy comes of age. J Clin Oncol 2011; 29 (36): 4828-36.
- 110. Van Mellaert L, Shahrooei M, Hofmans D, Eldere JV. Immunoprophylaxis and immunotherapy of Staphylococcus epidermidis infections: challenges and prospects. Expert Rev Vaccines 2012; 11 (3): 319–34.
- 111. Ma Q, Guishan Z, Wood TK. Escherichia coli BdcA controls biofilm dispersal in Pseudomonas aeruginosa and Rhizobium meliloti. BMC Res Notes 2011; 4: 447.
- 112. Ma Q, Yang Z, Pu M, Peti W, Wood TK. Engineering a novel c-di-GMP-binding protein for biofilm dispersal. Environ Microbiol 2011; 13 (3): 631–642.
- 113. Ha D-G, O'Toole GA. c-di-GMP and its effects on biofilm formation and dispersion: a Pseudomonas aeruginosa review. Microbiol Spectrum. 2015; 3 (2): 10. doi:10.1128/microbiolspec.MB-0003-2014.

- 571 114. Friedman L, Kolter R. Genes involved in matrix formation in Pseudomonas aeruginosa 572 PA14 biofilms. Mol Microbiol 2004; 51 (3): 675-690.
- 573 115. Friedman L, Kolter R. Two genetic loci produce distinct carbohydrate-rich structural 574 components of the Pseudomonas aeruginosa biofilm matrix. J Bacteriol 2004; 186 (14): 575 4457-4465.
- 576 116. Jackson KD, Starkey M, Kremer S, Parsek MR, Wozniak DJ. Identification of psl, a locus 577 encoding a potential exopolysaccharide that is essential for Pseudomonas aeruginosa 578 PAO1 biofilm formation. J Bacteriol 2004; 186 (1\$): 4466-4475.
- 117. Matsukawa M, Greenberg EP. Putative exopolysaccharide synthesis genes influence Pseudomonas aeruginosa biofilm development. J Bacteriol 2004; 186 (14): 4449-4456.
- 118. Ma L, Jackson KD, Landry RM, Parsek MR, Wozniak DJ. Analysis of Pseudomonas aeruginosa conditional psl variants reveals roles for the psl polysaccharide in adhesion and maintaining biofilm structure post attachment. J Bacteriol 2006; 188 (23): 8213-8221.
- 584 119. Vasseur P, Vallet-Gely I, Soscia C, Genin S, Filloux A. The pel genes of the Pseudomonas 585 aeruginosa PAK strain are involved at early and late stages of biofilm formation. 586 Microbiology 2005; 151 (Pt 3): 985-997.
- 120. Colvin KM, Irie Y, Tart CS, Urbano R, Whitney JC, Ryder C, Howell PL, Wozniak DJ, Parsek MR. The Pel and Psl polysaccharides provide Pseudomonas aeruginosa structural redundancy within the biofilm matrix. Environ Microbiol 2012; 14(8): 1913-28. doi:10.1111/j.1462-2920.2011.02657.x.
- Wei Q, Ma LZ. Biofilm Matrix and Its Regulation in Pseudomonas aeruginosa. Int J Mol
 Sci 2013; 14(10): 20983-21005. doi:10.3390/ijms141020983.
- 593 122. Franklin MJ, Nivens DE, Weadge JT, Howell PL. Biosynthesis of the Pseudomonas 594 Aeruginosa Extracellular Polysaccharides, Alginate, Pel, and Psl. Front Microbiol 2011; 2: 595 167. doi:10.3389/fmicb.2011.00167
- 596 123. Limoli DH, Jones CJ, Wozniak DJ. Bacterial Extracellular Polysaccharides in Biofilm 597 Formation and Function. Microbiol Spectr 2015; 3(3): 10. doi:10.1128/microbiolspec.MB-598 0011-2014.
- 599 124. Fujii N. D-amino acids in living higher organisms. Orig Life Evol Biosph 2002; 32(2): 103-600 27.
- 125. Kolodkin-Gal I, Romero D, Cao S, Clardy J, Kolter R, Losick R. D-amino acids trigger biofilm disassembly. Science 2010; 328 (5978): 627–629.
- 603 126. Cava F, Lam H, de Pedro MA, Waldor MK. Emerging knowledge of regulatory roles of D-604 amino acids in bacteria. Cell Mol Life Sci 2010; 68 (5): 817-831.
- 605 127. Campoccia D, Montanaro L, Arciola CR. A review of the biomaterials technologies for 606 infection-resistant surfaces. Biomaterials 2013; 34 (34): 8533–54. doi: 607 10.1016/j.biomaterials.2013.07.089

- 128. Ntrouka VI, Slot DE, Louropoulou A, Van der Weijden F. The effect of chemotherapeutic agents on contaminated titanium surfaces: a systematic review. Clin Oral Implants Res 2011; 22(7): 681-90. doi:10.1111/j.1600-0501.2010.02037.x
- 611 129. Ercan B, Kummer KM, Tarquinio KM, Webster TJ. Decreased Staphylococcus aureus 612 biofilm growth on anodized nanotubular titanium and the effect of electrical stimulation. 613 Acta Biomater 2011; 7(7): 3003-12. doi:10.1016/j.actbio.2011.04.002
- 130. Del Pozo JL, Rouse MS, Euba G, Kang CI, Mandrekar JN, Steckelberg JM, Patel R. The electricidal effect is active in an experimental model of Staphylococcus epidermidis chronic foreign body osteomyelitis. Antimicrob Agents Chemother 2009; 53(10):4064–8. doi:10.1128/AAC.00432-09
- 618 131. van der Borden AJ, van der Mei HC, Busscher HJ. Electric block current induced 619 detachment from surgical stainless steel and decreased viability of Staphylococcus 620 epidermidis. Biomaterials 2005; 26(33): 6731–5. doi:10.1016/j.biomaterials.2004.04.052
- 132. Pickering SA, Bayston R, Scammell BE. Electromagnetic augmentation of antibiotic efficacy in infection of orthopedic implants. J Bone Joint Surg Br 2003; 85 (4): 588–93. doi:10.1302/0301-620X.85B4.12644
- Hansen EN, Zmistowski B, Parvizi J. Periprosthetic joint infection: what is on the horizon? Int J Artif Organs 2012; 35 (10): 935-50. doi:10.5301/ijao.5000145
- 134. Taylor ZD, Navarro A, Kealey CP, Beenhouwer D, Haake DA, Grundfest WS, Gupta V.
 Bacterial biofilm disruption using laser-generated shockwaves. Conf Proc IEEE Eng Med Biol
 Soc 2010; 2010: 1028-32. doi: 10.1109/IEMBS.2010.5627726.
- 135. Kizhner V, Krespi YP, Hall-Stoodley L, Stoodley P. Laser-generated shockwave for clearing medical device biofilms. Photomed Laser Surg 2011; 29 (4): 277-82. doi:10.1089/pho.2010.2788
- 136. Francis NC, Yao W, Grundfest WS, Taylor ZD. Laser-Generated Shockwaves as a Treatment to Reduce Bacterial Load and Disrupt Biofilm. IEEE Trans Biomed Eng 2017; 634 64(4): 882-889. doi: 10.1109/TBME.2016.2581778. Epub 2016 Jun 15.
- 137. Schwandt LQ, Van Weissenbruch R, Stokroos I, Van Der Mei HC, Busscher HJ, Albers FW.
 Prevention of biofilm formation by dairy products and N-acetylcysteine on voice prostheses in an artificial throat. Acta Otolaryngol 2004; 124 (6): 726-731.
- 638 138. Riise GC, Qvarfordt I, Larsson S, Eliasson V, Andersson BA. Inhibitory effect of N-639 acetylcysteine on adherence of Streptococcus pneumoniae and Haemophilus influenzae to 640 human oropharyngeal epithelial cells in vitro. Respiration 2000; 67: 552-558.
- 139. Ricciotti E, FitzGerald GA. Prostaglandins and Inflammation. Arterioscler Thromband Vasc Biol 2011; 31(5): 986-1000. doi:10.1161/ATVBAHA.110.207449.
- 140. Kiecolt-Glaser JK. Stress, Food, and Inflammation: Psychoneuroimmunology and Nutrition at the Cutting Edge. Psychosom Med 2010; 72(4): 365-369. doi:10.1097/PSY.0b013e3181dbf489.

- 646 141. Hsieh C-C, Hsieh S-C, Chiu J-H, Wu Y-L. Protective Effects of N-acetylcysteine and a 647 Prostaglandin E1 Analog, Alprostadil, Against Hepatic Ischemia: Reperfusion Injury in Rats. J 648 Tradit Complement Med 2014; 4(1): 64-71. doi:10.4103/2225-4110.124351.
- 649 142. Blasi F, Page C, Rossolini GM, Pallecchi L, Matera MG, Rogliani P, Cazzola M. The effect 650 of N-acetylcysteine on biofilms: Implications for the treatment of respiratory tract 651 infections. Respir Med 2016; 117: 190-7. doi: 10.1016/j.rmed.2016.06.015.
- 652 143. Witkin SS, Jeremias J, Ledger WJ. A localized vaginal allergic response in women with 653 recurrent vaginitis. J Allergy Clin Immunol 1988; 81 (2): 412-416.
- 654 144. Alem MA, Douglas LJ. Effects of aspirin and other nonsteroidal anti-inflammatory drugs 655 on biofilms and planktonic cells of Candida albicans. Antimicrobial Agents Chemother 656 2004; 48 (1): 41-47.
- 657 145. Pe´Rez-Giraldo C, Rodriguez-Benito A, Moran FJ, Hurtado C, Blanco MT, 658 Gomez-Garcia AC. Influence of N-acetylcysteine on the formation of biofilm by 659 Staphylococcus epidermidis. J Antimicrob Chemother 1997; 39 (5): 643-646.
- 660 146. Xu XM, Sansores-Garcia L, Chen XM, Matijevic-Aleksic N, Du M, Wu KK. 661 Suppression of inducible cyclooxygenase 2 gene transcription by aspirin and sodium 662 salicylate. Proc Natl Acad Sci USA 1999; 96 (9): 5292-5297
- Torretta S, Rosazza C, Pace ME, Iofrida E, Marchisio P. Impact of adenotonsillectomy on pediatric quality of life: review of the literature. Ital J Pediatr. 2017; 43(1): 107. doi: 10.1186/s13052-017-0424-2.
- 666 148. Alasil SM, Omar R, Ismail S, Yusof MY, Dhabaan GN, Abdulla MA. Evidence 667 of Bacterial Biofilms among Infected and Hypertrophied Tonsils in Correlation with the 668 Microbiology, Histopathology, and Clinical Symptoms of Tonsillar Diseases. Int J 669 Otolaryngol 2013; 2013: 408238. doi:10.1155/2013/408238.
- Torrettaa S, Lorenzo Drago L, Marchisio P, Cappadona M, Rinaldi V, Nazzari E, Pignataro L. Recurrences in chronic tonsillitis substained by tonsillar biofilm-producing bacteria in children. Relationship with the grade of tonsillar hyperplasy. Int J Pediatr Otorhinolaryngol 2013; 77 (2): 200-204. doi.org/10.1016/j.ijporl.2012.10.018

674