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# **Changes in inflammatory markers after sleeve gastrectomy in subjects with impaired glucose homeostasis and type 2 diabetes**

## **Short title: Inflammatory markers after sleeve gastrectomy**

Akhila Mallipedhi MRCP<sup>1, 2</sup>; Sarah L Prior PhD<sup>1</sup>; Jonathan Barry MCh<sup>3</sup>; Scott Caplin FRCS<sup>3</sup>; John N Baxter MD<sup>3</sup>; Jeffrey W Stephens FRCP<sup>1, 2, 3</sup>

<sup>1</sup>Diabetes Research Group, Institute of Life Sciences, Swansea University, Swansea SA2 8PP, UK.

<sup>2</sup>Department of Diabetes & Endocrinology, Morriston Hospital ABM University Health Board, Swansea SA6 6NL, UK.

<sup>3</sup>Welsh Institute of Metabolic and Obesity Surgery, Morriston Hospital ABM University Health Board, Swansea SA6 6NL, UK.

## **Correspondence and reprint requests to:**

Dr. Akhila Mallipedhi

Diabetes Research Group, Institute of Life Sciences, Swansea University, Swansea SA2 8PP, UK

Tel: +44 (0)1792 704078; Fax: +44 (0)1792 703214

E-mail: mallipedhi@doctors.org.uk

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<sup>1</sup>Diabetes Research Group, Institute of Life Sciences, Swansea University, Swansea SA2 8PP, UK.

<sup>2</sup>Department of Diabetes & Endocrinology, Morriston Hospital ABM University Health Board, Swansea SA6 6NL, UK.

<sup>3</sup>Clinical Research Unit, Morriston Hospital, ABM University Health Board, Swansea SA6 6NL, UK.

<sup>4</sup>Welsh Institute of Metabolic and Obesity Surgery, Morriston Hospital ABM University Health Board, Swansea SA6 6NL, UK.

<sup>5</sup>Department of Diabetes & Endocrinology, Hywel Dda Health Board, Llanelli, Carmarthenshire SA14 8QF, UK.

# **Changes in inflammatory markers after sleeve gastrectomy in subjects with impaired glucose homeostasis and type 2 diabetes**

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**Conflict of interest:** The authors declare that there is no conflict of interest associated with this manuscript.

## **Abstract**

*Background:* Bariatric surgery is an effective treatment for morbid obesity. Obesity and type 2 diabetes are associated with chronic inflammation. There is lack of data examining the effects of sleeve gastrectomy (SG) on inflammatory biomarkers.

*Objectives:* Our aim was to study the effects of SG on specific cytokines associated with obesity including interleukin-6 (IL-6), interleukin-10 (IL-10), leptin, adiponectin and C-reactive protein (CRP) preoperatively, 1 and 6 months after surgery.

*Setting:* University Hospital, United Kingdom.

*Methods:* A non-randomized prospective study comprising of 22 participants with impaired glucose homeostasis and type 2 diabetes undergoing SG (body mass index [BMI] 50.1kg/m<sup>2</sup>, Glycated hemoglobin [HbA<sub>1c</sub>] 53 mmol/mol). Serial measurements of IL-6, IL-10, leptin, adiponectin, and CRP were performed during oral glucose tolerance testing pre-operatively, 1 and 6 months postoperatively.

*Results:* We observed significant improvements at 1 and 6 months in leptin ( $p < 0.001$ ) and CRP ( $p = 0.003$ ) following SG. We also observed a significant reduction in IL-6 at 6 months ( $p = 0.001$ ). No statistically significant differences were observed for adiponectin and IL-10.

*Conclusion:* This study is the first to examine the detailed changes in the inflammatory cytokines after SG. Our study shows significant improvements in the inflammatory biomarkers following SG in subjects with impaired glucose homeostasis and type 2 diabetes.

## **Keywords**

Obesity; Type 2 diabetes mellitus; Sleeve Gastrectomy; Inflammation

## **Introduction**

Systemic inflammation is closely associated with obesity and type 2 diabetes mellitus (T2DM). Prospective studies have shown that chronic low grade systemic inflammation predicts the future risk of impaired glucose tolerance (IGT), T2DM<sup>(1)</sup> and cardiovascular disease (CVD)<sup>(2)</sup>. Certain subgroups of the population that are at a high risk of developing T2DM have elevated inflammatory markers. These include overweight adults and children, women with polycystic ovary syndrome, certain ethnic origins (e.g. Pima Indians) and participants with a family history of T2DM<sup>(3)</sup>. Interestingly, these groups often also share the common feature of obesity. The Finnish and American Diabetes Prevention studies both showed a 58% reduction in progression from IGT to T2DM, by diet and exercise<sup>(4, 5)</sup>. In addition, several of the studies looking at CVD risk reduction with statins (WOSCOPS) and angiotensin-1 converting enzyme inhibitors (HOPE, CAPP) showed a 25-30% reduction in the development of T2DM<sup>(6-8)</sup>, presumably through an underlying anti-inflammatory

mechanism. These previously described results reinforce the close relationship between obesity, inflammation and T2DM.

Adipokines are cytokines secreted by adipose tissue and can modulate immune response, insulin sensitivity and energy balance. Cytokines associated with obesity include interleukin-6 (IL-6), interleukin-10 (IL-10), adiponectin, leptin and C-reactive protein (CRP) <sup>(9-11)</sup>. Changes in plasma levels of pro-inflammatory and anti-inflammatory cytokines and adipokines are related to increased adipose tissue mass <sup>(1,2)</sup>.

Bariatric surgery is effective in the treatment of obesity and T2DM and in reducing CVD risk factors <sup>(12)</sup>. This is likely to occur through a reduction in adipose tissue mass and subsequent improvements in the inflammatory milieu <sup>(13, 14)</sup>. Previous work has demonstrated malabsorptive surgery such as Laparoscopic Roux-en-Y Gastric Bypass (RYGB) is associated with significant reductions in serum levels of leptin and non-significant changes in soluble TNF- $\alpha$  receptor-1 and adiponectin levels 6 months following surgery <sup>(15)</sup>. CRP has also been observed to be significantly lower at 6 months compared to preoperative values. A significant reduction in CRP, IL-6 and an increase in adiponectin have also been observed 6 months after gastric bypass in another study <sup>(16)</sup> and is supported by the results of a recent meta-analysis <sup>(17)</sup>. Over recent years an increasing number of SG operations have been performed relative to RYGB, Bilio-pancreatic Diversion (BPD) and laparoscopic adjustable Gastric Banding (LAGB) and furthermore SG is now recognized as a stand-alone bariatric procedure with a superior safety profile <sup>(18, 19)</sup>. Prospective studies describe a reduction in CRP at 4 weeks in 11 patients following a SG <sup>(20)</sup> and in 37 participants at 9 months <sup>(21)</sup>. An improvement in CRP has also been observed in a retrospective hospital database study of 61 patients <sup>(22)</sup>. However, there is limited information within the available literature with regard to the effects of SG on other cytokines and adipokines.

Our aim was to specifically examine the temporal changes in IL-6, IL-10, leptin, adiponectin and CRP preoperatively, 1 and 6 months after SG in a sample of subjects with impaired glucose tolerance or T2DM.

## **Participants and methods**

### **Study participants**

Approval for the study was obtained from the Local Research Ethics Committee (South Wales; LREC reference 06/WMW02/7) and the Joint Scientific Research Committee at Swansea University and ABM University Health Board. Participants were identified and recruited from patients undergoing a planned bariatric surgical procedure at Welsh Institute of Metabolic and Obesity Surgery (WIMOS) at Morriston Hospital, ABM University Health Board, Swansea, Wales, UK. Entry criteria at the outset of the study included:- both sexes, age 20-40 years, BMI  $>40\text{kg/m}^2$  and physically fit for surgery. Participants with any acute concurrent illness were excluded. Participants with previously diagnosed T2DM treated with diet, oral agents, GLP-1 analogues or insulin were included. All participants underwent an oral glucose tolerance test (OGTT) prior to recruitment. Participants with impaired glucose regulation were those with either impaired fasting glycaemia (5.6-6.9 mmol/L) or impaired glucose tolerance (2-hour glucose 7.8-11.0 mmol/L) <sup>(23)</sup>.

### ***Study design***

Participants with a planned SG were recruited prospectively and consecutively from the bariatric surgical clinic. SG is a standard sleeve i.e. sleeve fashioned around a 32F bougie taken from 5cm proximal to the pylorus and up to the left crus. All participants were recruited

pre-operatively (within 1 month of surgery) and followed up postoperatively at 1 and 6 months. All participants with the help of the research nurse completed a baseline questionnaire and all clinical measurements were documented during the visits. All blood samples were collected after stopping any prescribed insulin or oral hypoglycaemic agent for 24 hours prior to an OGTT performed with 75g of glucose (122mLs of Polycal 61.9g/100mL of glucose, Nutricia Clinical Care, Trowbridge, UK).

### ***Baseline clinical and biochemical information***

At the time of screening the following clinical information was ascertained: age, gender, past medical history, treatment and duration of diabetes. Baseline clinical measurements consisted of weight, height, BMI, waist circumference, systolic and diastolic blood pressure. Baseline biochemical measurements (total cholesterol, Low density lipoprotein-cholesterol [LDL-C], High density lipoprotein-cholesterol [HDL-C] and triglycerides) were analyzed within the local hospital accredited laboratory. Glucose and lipids (Roche Modular P800 Analyzer) and insulin and C-peptide (Roche E170 Modular Analyzer) were also measured locally. All blood samples were collected on ice, centrifuged and separated within one hour of collection and subsequently stored at  $-80^{\circ}\text{C}$  until analysis. Fasting EDTA samples were collected for the measurement of cytokines during the OGTT at baseline, 1 and 6 months.

### ***Measurement of total cytokines***

Fasting plasma levels of IL-6, IL-10 and leptin were measured with high sensitivity ELISA kits (R&D Systems). Intra-assay and inter-assay variability coefficients were as follows: IL-6  $\leq 4.2\%$  and  $\leq 6.4\%$ ; IL-10,  $\leq 5.0\%$  and  $\leq 7.5\%$ ; leptin,  $\leq 3.3\%$  and  $\leq 5.4\%$ .

Fasting plasma levels of total adiponectin and CRP were measured with high sensitivity ELISA kits (Immundiagnostik AG). Intra-assay and inter-assay variability coefficients were

as follows: adiponectin,  $\leq 3.4\%$  and  $\leq 6.3\%$ ; CRP,  $\leq 6.0\%$  and  $\leq 13.8\%$ . All samples were assayed in duplicate.

### ***Statistical methods***

Statistical analysis was performed using SPSS (version 10.1, SPSS Inc., Chicago). Results for continuous variables are presented as mean and standard deviation and in graphical representation as mean and standard error. Continuous variables that did not have a normal distribution (triglyceride, adiponectin, CRP and IL-6) underwent log transformation to normalize the data for analysis and are described with the geometric mean and approximate standard deviation. For continuous variables, the mean temporal changes were compared between baseline and 1 or 6 months using a paired t-test. Categorical data were analysed using a Chi-squared test. Paired t-tests were used to compare mean differences in cytokines between baseline, 1 and 6 months for IL-10, leptin, adiponectin, CRP and IL-6 (with log transformation where appropriate). In all cases a  $p < 0.05$  was considered statistically significant.

## **Results**

### **Subject characteristics**

A total of 22 participants who underwent a SG (mean age  $48 \pm 7$  years) with impaired glucose homeostasis or T2DM (median duration of 42 month [interquartile range 21-66 months]) completed the study. Table 1 shows the temporal changes between baseline, 1 and 6 months within the group. As shown in Table 1, significant reductions were observed in weight at 1 month with a mean reduction in BMI of  $10.5 \text{ kg/m}^2$  (weight change of 31.1kg) at 6 months.



We also observed a significant reduction in the systolic blood pressure at 1 month but no significant changes in total cholesterol, LDL-C or triglyceride concentrations. Significant changes were observed in fasting glucose, 2-hour glucose and HbA<sub>1c</sub>.

### **Temporal changes in inflammatory cytokines following sleeve gastrectomy**

The temporal changes in fasting inflammatory cytokines between baseline, 1 and 6 months are shown in Figure 1. Plasma leptin decreased by 34.4% and 43.1% respectively at 1 and 6 months postoperatively. Compared to baseline, significant reductions of 67.5% and 44.8% were observed for CRP respectively. At 6 months there was a significant reduction (31.6%) in plasma IL-6. There was a non-significant increase of adiponectin at 6 months ( $p=0.15$ ). No significant changes were observed in IL-10.

### **Discussion**

Bariatric surgery in addition to weight loss effectively reduces morbidity and mortality in severely obese individuals with favorable effects on T2DM, hypertriglyceridaemia and hypertension. These effects are likely to be related to an improvement in the inflammatory profile caused by the rapid and significant reductions in fat mass following surgery<sup>(16)</sup>. SG has gained recent popularity as an independent bariatric procedure<sup>(18, 19)</sup>. We observed significant improvements in leptin and CRP at 1 month postoperatively following SG and this was maintained at 6 months. We also observed a reduction in IL-6 at 1 month which was statistically significant at 6 months. No differences were observed for adiponectin and IL-10 following SG. Previous studies have only described changes in CRP as early as 4 weeks following a SG<sup>(20-22)</sup>.

We also examined the correlations between the change in BMI with the inflammatory biomarkers. At 1 month, there was a significant correlation between greater weight loss and

increased plasma IL-10 ( $r=0.50$ ,  $p=0.008$ ) and an inverse association between increased adiponectin and IL-6 ( $r=-0.50$ ,  $p=0.04$ ), such that an increase in adiponectin was associated with reduction in IL-6. These changes would be consistent with a reduction in adipose tissue mass being responsible for increased levels of the anti-inflammatory adipokine adiponectin and a reduction in the proinflammatory cytokine IL-6. This is in line with previous studies which have demonstrated that adipose tissue is a major source of IL-6 with as much as 30% of circulating IL-6 being derived from adipose tissue (24). Circulating levels of IL-6 are raised in insulin resistant states such as obesity<sup>(25)</sup>, IGT<sup>(26)</sup> and T2DM<sup>(27-29)</sup> and previous prospective studies have shown plasma IL-6 to correlate with BMI, percentage fat mass and fasting insulin levels. Weight reduction by diet and exercise has also been shown to reduce plasma IL-6 levels<sup>(30)</sup>.

This study is the first to examine the temporal relationship of these inflammatory biomarkers at 1 and 6 months following SG. This current study contributes to the available literature supporting the role of SG for the treatment of T2DM, and pro-inflammatory conditions associated with morbid obesity. One limitation of the study was that the sample group was heterogeneous comprising of subjects with impaired glucose tolerance and type 2 diabetes. Furthermore, the duration of diabetes was variable within the study group (median 42 months [interquartile range 21-66 months]) and that this might have an effect on the levels of inflammatory markers pre-operatively. Furthermore, the duration of diabetes was obtained from primary care medical records and this is therefore an estimate as diabetes may be present for sometime before the diagnosis is made<sup>(31)</sup>.

Further prospective studies are required to examine the effects of SG in relation to biomarkers of inflammation and oxidative stress in relation to inflammatory mediated complications of obesity.

**Table 1: Baseline and end of study clinical and biochemical measurements within the SG group**

Measurement	Baseline	1 month	<sup>a</sup> P	6 months	<sup>b</sup> P
Weight (kg)	146.6 (29.6)	129.4 (26.9)	<0.001	115.5 (24.4)	<0.001
BMI (kg/m <sup>2</sup> )	50.1 (6.6)	44.0 (6.6)	<0.001	39.6 (6.2)	<0.001
Waist (cm)	138 (18)	128 (17)	<0.001	118 (18)	<0.001
Systolic BP (mmHg)	131 (18)	123 (14)	0.04	128 (20)	0.12
Diastolic BP (mmHg)	76 (11)	74 (9)	0.63	73 (14)	0.13
Cholesterol (mmol/L)	4.3 (1.0)	4.4 (1.1)	0.58	4.7 (1.2)	0.09
LDL-C (mmol/L)	2.4 (0.8)	2.7 (1.0)	0.16	2.8 (0.9)	0.05
HDL-C (mmol/L)	1.2 (0.3)	1.1 (0.3)	0.02	1.3 (0.3)	0.10
Triglyceride (mmol/L) <sup>c</sup>	1.4 (0.4)	1.4 (0.2)	0.92	1.1 (0.2)	0.15
HbA <sub>1c</sub> (%)	7.0 (1.7)	6.1 (0.8)	0.005	5.7 (0.8)	0.002
HbA <sub>1c</sub> (mmol/mol)	53 (18.6)	43 (8.7)	0.005	39 (8.7)	0.002
Fasting glucose (mmol/L)	7.6 (3.6)	5.4 (0.9)	0.02	5.0 (1.0)	0.08
2-hour glucose (mmol/L)	11.6 (5.9)	7.8 (3.4)	0.002	5.4 (2.2)	<0.001

Mean and standard deviation shown for continuous variables

<sup>a</sup>P-value comparing baseline with 1 months.

<sup>b</sup>P-value comparing baseline with 6 months.

<sup>c</sup>log transformed for analysis. Geometric mean and approximate standard deviation shown for log- transformed data.

In the group there were 15 females and 7 males.

BMI= Body mass index, LDL-C= Low density lipoprotein-Cholesterol, HDL-C= High

density lipoprotein-Cholesterol, HbA<sub>1c</sub>= Glycated hemoglobin

## Figure legends

### Figure 1: Temporal changes in inflammatory biomarkers following sleeve gastrectomy

Mean and standard error is shown.

\*P<0.05: Significant changes relative to baseline.

Figure 1a: Interleukin-10.

Figure 1b: Leptin

Figure 1c: Adiponectin

Figure 1d: C-reactive protein

Figure 1e: Interleukin-6

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Figure 1a: Interleukin-10

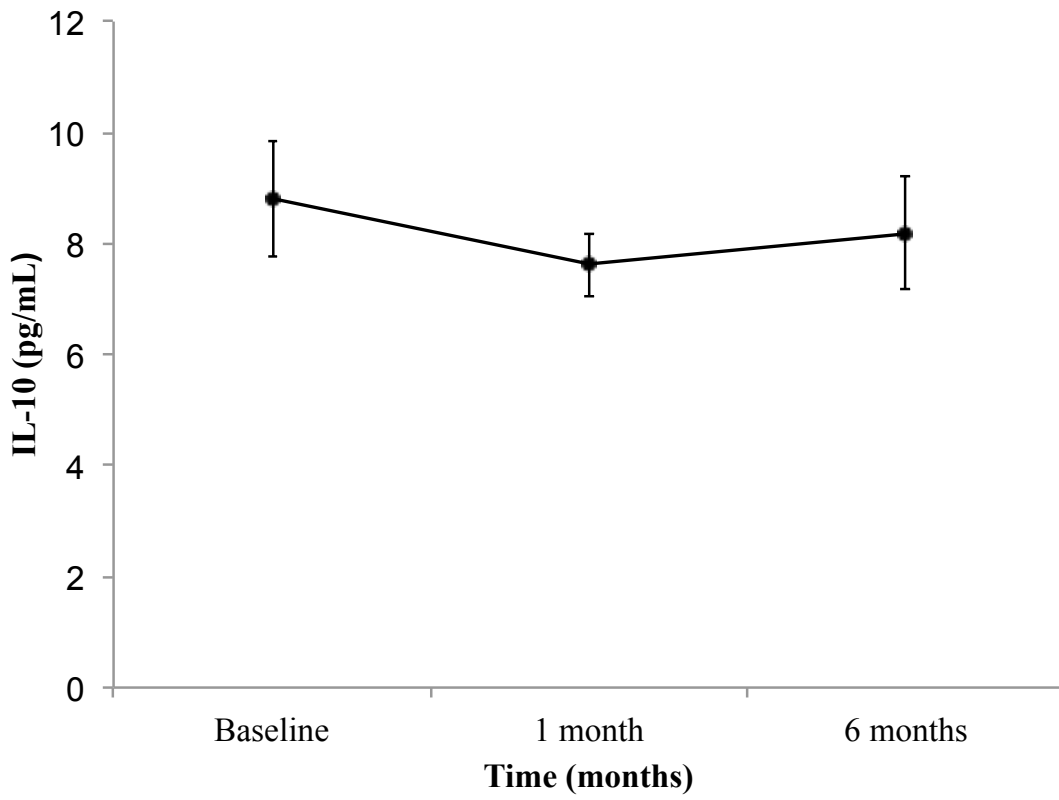


Figure 1b: Leptin

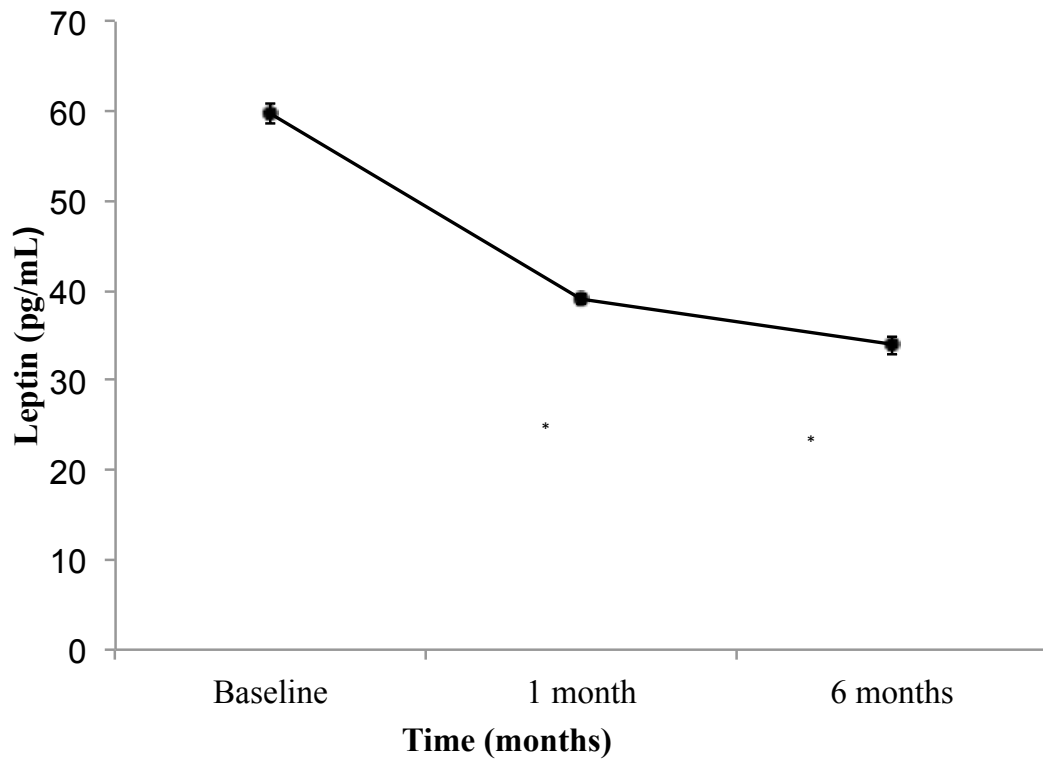


Figure 1c: Adiponectin

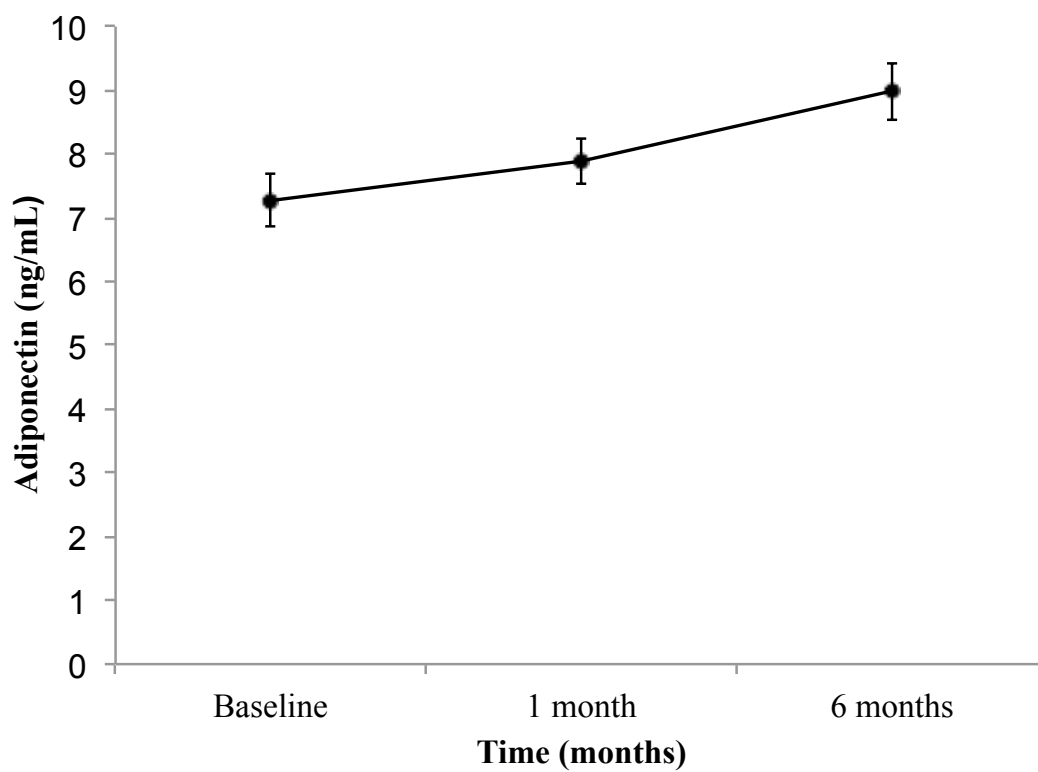


Figure 1d: C-reactive protein

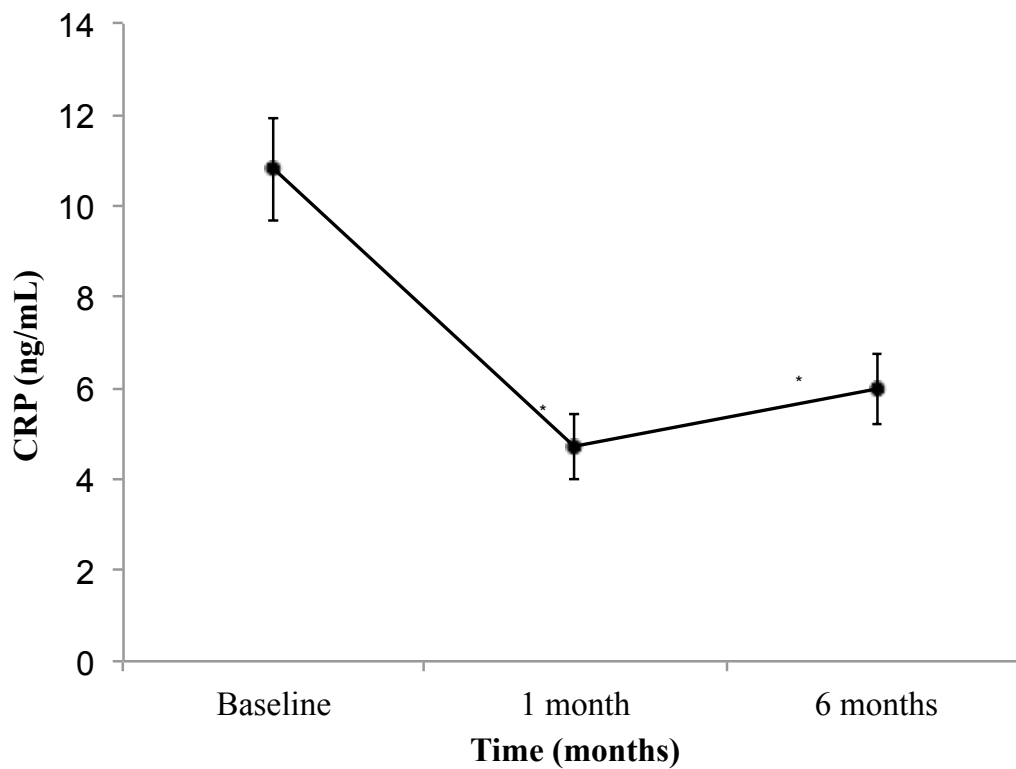




Figure 1e: Interleukin-6

